

Association of *NOS3* gene variants and clinical contributors of hypoxic-ischemic encephalopathy

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Abstract

The aim of this study was to analyze the association of different clinical contributors of hypoxic-ischemic encephalopathy with *NOS3* gene polymorphisms. A total of 110 children with hypoxic-ischemic encephalopathy and 128 control children were selected for this study. Association of gender, gestational age, birth weight, Apgar score, cranial ultrasonography, and magnetic resonance imaging findings with genotypic data of six haplotype-tagging single nucleotide polymorphisms and the most commonly investigated rs1800779 and rs2070744 polymorphisms was analyzed. The TGT haplotype of rs1800779, rs1800779, and rs2070744 polymorphisms was associated with hypoxic-ischemic encephalopathy. Children with the TGT haplotype were infants below 32 weeks of gestation and they had the most severe brain damage. Increased incidence of the TT genotype of the *NOS3* rs1808593 SNP was found in the group of hypoxic-ischemic encephalopathy patients with medium and severe brain damage. The probability of brain damage was twice as high in children with the TT genotype than in children with the TG genotype of the same polymorphism. Furthermore, the T allele of the same polymorphism was twice as frequent in children with lower Apgar scores. This study strongly suggests associations of *NOS3* gene polymorphism with intensity of brain damage and severity of the clinical picture in affected children.

Key words: Clinical contributors; Hypoxic-ischemic encephalopathy; *NOS3* gene polymorphisms

Introduction

Hypoxic-ischemic encephalopathy (HIE), a serious condition affecting infants in the perinatal period, is the most common cause of neurologic deficits in children. It may occur in premature and full-term children as a result of an intrauterine infection and/or chorioamnionitis, premature rupture of membranes, maternal trauma and shock (1). Lack of oxygen, caused by impairment of cerebral blood flow, starts a cascade of harmful biochemical processes at the cellular level, resulting in serious brain damage (2-4).

One of the mechanisms of cell-structure degradation is mediated by activation of a nitric oxide synthase (NOS) and by formation of free radicals of oxygen and nitrogen (5-7). Exposure to mild ischemia activates endothelial nitric oxide synthase (*NOS3*), a Ca^{2+} -dependent enzyme;

small amounts of NO are produced, causing vasodilatation, and the consequent maintenance of cerebral blood flow prevents neuronal injury. NO catalyzed by *NOS3* also affects other processes such as inhibition of platelet and leukocyte adhesion to endothelium, inhibition of platelet aggregation, relaxation and inhibition of cell proliferation of blood vessel smooth muscle, stimulation of angiogenesis, anti-inflammatory action, and prevention of oxidative damage (8). Production of NO in endothelial blood vessel cells is regulated by the *NOS3* gene (OMIM: 163729). Several studies reported that *NOS3* gene polymorphisms may alter enzymatic function and therefore may be associated with HIE (9-11).

Our previous study showed genotype and haplotype association of *NOS3* gene polymorphisms with HIE.

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Received February 21, 2014. Accepted June 2, 2014. First published online August 15, 2014.

Genotypic tests of six haplotype-tagging single nucleotide polymorphisms (SNPs; rs3918186, rs3918188, rs1800783, rs1808593, rs3918227, rs1799983), and the most commonly investigated rs1800779 SNP, showed an association of rs1808593 polymorphism with HIE. The TG genotype of rs1800783-rs1800779 polymorphisms was found less frequently, and the TT genotype more frequently, in a group of affected patients than in a control group. We also observed an association of the TG haplotype with HIE (12).

Severity of clinical presentation and degree of brain damage can be influenced by a genetic predisposition to development of the disease. The aim of this study was to analyze the association of different clinical contributors that define the severity of disease and patient condition with *NOS3* gene polymorphisms that we investigated in our previous study.

Material and Methods

A total of 110 unrelated full-term or preterm children (69 boys and 41 girls) with HIE (HIE group) were recruited for the study. The study included all HIE patients treated in the University Hospital Split, Croatia between 1992 and 2008, and included children who met at least one of the following criteria: fetal heart rate during delivery <100 or >160/min, Apgar score ≤ 7 at 5 min, arterial pH ≤ 7.1 , neonatal neurological sequelae classified by Sarnat and Sarnat (13), or positive hypoxic-ischemic brain damage found on repeated cranial ultrasonography at different time periods. Perinatal hypoxic-ischemic brain injuries of newborns were detected by cranial ultrasonography. With cranial ultrasonography, we determined the grade of periventricular leukomalacia (PVL) according to criteria described by Weisglas-Kuperus (14): grade I PVL, prolonged periventricular flare present for 7 days or more; grade II PVL, prolonged periventricular flare with small, localized frontoparietal cysts; grade III PVL, extensive periventricular cystic lesions involving occipital and frontoparietal white matter; grade IV PVL, areas of extensive subcortical cystic lesions.

Hypoxic-ischemic brain damage in full-term infants was also confirmed by serial cranial ultrasonography performed at 3, 7, 14, and 21 days after delivery, followed by once per month until closing the anterior fontanel. Ultrasonography was performed by two independent experts.

Final diagnosis and grade of hypoxic-ischemic brain damage in the HIE group was determined by magnetic resonance imaging (MRI) at the age of 2 years, when the processes of proliferation, migration, organization, and myelination were completed. The altered signal of the white matter, appearance of cystic changes, changes in the size of the lateral ventricle anterior horns, extension of subarachnoid space and thinning of the corpus callosum were followed (Table 1) (15,16). Each parameter that was

observed was scored from 1-3 points: grade 1, total scores of up to 5 points were considered normal findings, grade 2, moderate injury, included total scores of 6-10 points, and grade 3, severe brain injury, comprised total scores of 11-15 points.

For the two groups studied, demographic data and other characteristics such as gender, gestational age (≥ 37 , 32 to 36, <32 weeks), and birth weight were collected from the medical documentation. For the data analysis, study participants were divided into four sub-groups according to birth weight (≤ 1000 , 1001-1499, 1500-2499, and ≥ 2500 g.) and Apgar score (≤ 5 , 6-7, 8-9, and 10). Patients with congenital malformations of the central nervous system (CNS), metabolic diseases, chromosomal abnormalities, with infections, and patients with CNS trauma, were excluded. We also excluded infants from multiple gestations, and infants of mothers who had preeclampsia and placental abruption.

The HIE group consisted of 80 patients (72.7%) with hypoxic-ischemic injury and 30 patients (27.3%) with hypoxic-ischemic injury accompanied by intracranial hemorrhage. The control group consisted of 128 apparently healthy full-term and preterm born children (60 boys and 68 girls), with normal brain ultrasound findings and/or normal neurological status after the second year of life. The HIE and the control subject populations were the same as in our previous publication (12).

Additionally, another frequently investigated polymorphism, rs2070744, was genotyped in both the HIE and control groups. Real-time polymerase chain reaction (PCR) was performed using the ABI PRISM 7500 Sequence Detection System (Applied Biosystems, USA) following the manufacturer's protocol.

The study was approved by the Ethics Committee of University Hospital Split and School of Medicine, University of Split, and informed consent was obtained from patients' parents prior to the blood sampling.

Statistical analysis

Quality control analysis of the obtained rs2070744 genotypes was performed. Genotype frequencies, Hardy-Weinberg equilibrium (HWE), and minor allele frequencies (MAF) for all samples were analyzed using HaploView 4.1 (<http://haploview.software.informer.com/4.1/>). An 80% study power at $\alpha = 0.05$ was calculated using Quanto (<http://biostats.usc.edu/Quanto.html>), and results are reported as odds ratios (ORs). When $OR > 1$, the genotype confers sensitivity to the effects of exposure (17). Case-control association analysis as well as cis-position haplotype analysis of 8 SNPs was carried out using Haploview 4.1.

Chi-square tests were used to analyze genotypic and allelic distribution for each polymorphism examined by gestational age, gender, Apgar score, birth weight, and MRI findings. The Yates correction was performed when necessary.

Table 1. Scoring of perinatal hypoxic-ischemic brain injury according to magnetic resonance imaging.

	Grade 1 (normal findings)	Grade 2 (moderate brain injury)	Grade 3 (severe brain injury)
White matter signal abnormality	Normal signal	Focal altered signal (one region)	Changed signal to 2 or more regions
Size of lateral ventricle anterior horns	Proper size up to 8 mm	Mild to moderate dilatation, 8-10 mm	Significant dilatation >10 mm
Cystic change	No	Focal cystic changes	Extensive cystic changes in white matter
Corpus callosal thinning	Normal	Thinned <3 mm focal	Thinned <3 mm multifocal
Subarachnoid space	Proper size up to 4 mm	Slightly extended, 4-8 mm	Moderate to very extended \geq 8 mm

Logistic regression (Statistica 7.0; <http://statistica.software.informer.com/7.0/>) was performed to calculate OR in order to determine incidence of hypoxic-ischemic brain damage with respect to gestational age, gender, Apgar score, birth weight, and haplotype. Multiple logistic regression with regard to haplotype distribution was also performed in order to prove logistic regression results. Chi-square tests were also performed to compare the degree of brain damage established by MRI with regard to gender, gestational age, birth weight, and Apgar score.

Results

The genotype call rate for the investigated rs2070744 polymorphisms was 100%, and the genotypes in the case and control groups fit HWE. The rs2070744 polymorphisms showed strong linkage disequilibrium with earlier investigated rs1800783 and rs1800779 polymorphisms ($r^2=0.96$ and $r^2=0.99$, respectively). Allelic and genotype analysis of rs2070744 polymorphism did not show any association with HIE. The study had 80% statistical power to detect ($\alpha=0.05$) an effect with an OR=1.76 for rs2070744, assuming an additive model. In addition, the cis TGT haplotype of rs1800783, rs1800779, and rs2070744 polymorphisms of the NOS3 gene was found in only five of the HIE patients and was not found in the control group.

Genotype distribution analysis of the six haplotype-tagging SNPs (rs3918186, rs3918188, rs1800783, rs1808593, rs3918227, rs1799983) and the most commonly investigated rs1800779 and rs2070744 SNPs were determined in the case and control groups and evaluated according to clinical contributors (i.e., gestational age, gender, Apgar score, birth weight, and MRI findings). The analysis showed that the TT genotype of rs1808593 polymorphism was twice as common as TG and GG in the HIE group in patients with moderate brain injury (grade 2; TT=26, TG=11, GG=1) and severe brain injury (grade 3; TT=41, TG=25, GG=6) than in the control group (grade 1; TT=59, TG=66, GG=3; $\chi^2=12.36$, $P=0.015$). In children with normal MRI findings (grade 1), the most

common rs1808593 genotype was TG.

Comparing the genotype distribution of the same rs1808593 polymorphisms between normal MRI findings (grade 1) and pathological MRI findings regardless of the severity of damage (grades 2 and 3), a statistically significant association ($\chi^2=6.572$, $P=0.0104$) was found, with a higher incidence of the TT genotype in children with abnormal MRI findings.

The allele distribution analysis of all investigated polymorphisms showed a statistically significant more frequent occurrence of the T allele of the rs1808593 polymorphism in children with low Apgar scores. The T allele was more frequent in all groups, as expected; but the difference between T and G allele frequency was more significant as Apgar score was lower ($\chi^2=10.4$, $P=0.015$; Table 2).

The genotype distribution analysis of polymorphism rs3918186, showed that AA was twice as frequent as AT in children with normal Apgar scores (AS=10) but that the difference was not statistically significant ($\chi^2=9.4$, $P=0.152$; Table 3). Furthermore, analysis of HIE children with TGT haplotype polymorphisms rs1800783, rs1800779, rs2070744 of the NOS3 gene showed that they were premature infants of <32 weeks of gestation with the most severe degree of brain damage (grade 3) on the MRI findings.

Logistic regression analysis showed that the OR for incidence of hypoxic-ischemic brain damage was up to two times higher in male than in female children ($P=0.01$; Table 4). The OR for the occurrence of disease was 1.92 times higher in children with Apgar scores <5 than in those with scores of 10 ($P<0.001$). Also, the OR for the occurrence of disease was 1.32 times higher in children with lower birth weight ($P=0.03$) and 1.62 times higher in children with lower gestational age ($P=0.003$). Since the TGT haplotype was found in only five patients with HIE, logistic regression analysis was performed only for the two predominant haplotype combinations, TAT and AGC, and showed no significant difference in occurrence ($P=0.573$; Table 4).

After applying multiple logistic regression with regard to haplotype distribution, the difference in occurrence of

Table 2. Allele distribution analysis of *NOS3* rs1808593 polymorphism and Apgar score.

Apgar score	Total alleles	T allele	G allele	χ^2	P
≤5	166 (100%)	130 (78%)	36 (22%)	10.4	0.015
6-7	142 (100%)	105 (74%)	37 (26%)		
8-9	70 (100%)	45 (64%)	25 (36%)		
10	98 (100%)	74 (75%)	44 (45%)		

Data are reported as number with percent in parentheses. The χ^2 test was used for analyses.

disease was maintained for gender (OR=1.98, 95% CI=1.1-3.5, P=0.021) and Apgar score (OR=2.13, 95%CI=1.5-3.0, P<0.001).

There was a 90% probability for children of a lower gestational age to have very severe brain damage (MRI grade 3), and for children with greater gestational age (≥ 37 weeks) to have a 90% probability to have a milder form of damage (grade 2; Table 5).

There was a 90% probability for lower birth weight children to have severe brain damage (grade 3). Also, there was almost 90% probability for children with a birth weight >2500 g to have a milder form of brain damage (grade 2, Table 6).

Discussion

The results of this study strengthen the association of *NOS3* gene polymorphisms with HIE and also provides evidence of their association with certain clinical contributors and intensity of brain damage. We found the *TGT* haplotype of *NOS3* gene polymorphisms rs1800783, rs1800779, and rs2070744 in five patients with HIE, with no such haplotype in the control group. This finding highlights even more our previous finding of an rs1800783-rs1800779 TG haplotype association with HIE (12).

NO is an important and well-characterized vasodilator, with homeostasis of NO essential in maintaining vascular tone in the systemic and cerebral circulation (8,10). Several earlier studies investigating possible causes of cerebral palsy indicated that genes that are associated with vascular disease or inflammation, and with the fetal inflammatory response, may also be linked with cerebral palsy development (18,19). Polymorphisms rs1800783,

rs1800779, and rs2070744 are located in the upstream and promoter region of the *NOS3* gene that may influence mRNA transcription and can reduce gene expression; this may further lead to impaired production or reduced bioavailability of NO, thus causing susceptibility to severe brain injury (20). Lower levels of *NOS3* expression may lead to reduced protective effects in endothelial cells, thus predisposing to the occurrence of stroke (11).

This study also showed that children with the *TGT* haplotype were premature infants below 32 weeks of gestational age and had the most severe brain damage (grade 3) on MRI. It is interesting that we observed rs1808593 TT genotype association with HIE in our previous study (12). In this study, we further confirmed and elucidated the significance of this polymorphism in children affected by HIE. We found a higher incidence of the TT genotype of *NOS3* rs1808593 polymorphism in those patients with medium and severe brain damage.

We analyzed the genotypic distribution of polymorphism rs1808593 with regard to MRI findings and found twice the prevalence of the TT genotype in the group of children with grade 2 and grade 3 brain injury. Thus, prevalence of the TT genotype was increased in children with abnormal MRI findings regardless of the degree of damage, while among children with normal MRI findings (control group), the TG genotype was predominant.

Also, this study showed a frequent occurrence of the T allele of rs1808593 polymorphism in children with low Apgar scores. Although the T allele was more frequent in all groups, the difference in frequency between T and G allele was significant in children with low Apgar scores. Such a detailed analysis of genotypic and allelic correlations with clinical contributors allows us to suggest that

Table 3. Genotype distribution analysis of *NOS3* gene polymorphism rs3918186 and Apgar score.

Apgar score	Genotype AA	Genotype AT	Genotype TT	χ^2	P
≤5	65 (35%)	18 (40%)	0 (0%)	9.4	0.152
6-7	56 (30%)	15 (33%)	0 (0%)		
8-9	27 (14%)	7 (16%)	1 (20%)		
10	40 (21%)	5 (11%)	4 (80%)		

Data are reported as number with percent in parentheses. The χ^2 test was used for analyses.

Table 4. Odds ratios for occurrence of hypoxic-ischemic encephalopathy according to gender, Apgar score, body weight, gestational age and haplotype distribution.

	Control group	HIE group	χ^2	P	OR (95%CI)	P
Gender						
M	60 (47%)	70 (64%)	6.7	0.01	1.98 (1.18-3.34)	0.01
F*	68 (53%)	40 (36%)				
Apgar score						
≤5	33 (26%)	50 (46%)	29.3	<0.001	1.92 (1.48-2.5)	<0.001
6-7	32 (25%)	39 (36%)				
8-9	21 (16%)	14 (12%)				
10*	42 (33%)	7 (6%)				
Body weight						
≤1000	12 (9%)	14 (13%)	8.9	0.03	1.32 (1.0-1.7)	0.03
1001-1499	29 (23%)	28 (25%)				
1500-2499	34 (27%)	42 (38%)				
≥2500*	53 (41%)	26 (24%)				
Gestational age						
≤31	45 (35%)	52 (47%)	11.6	0.003	1.62 (1.2-2.2)	0.003
32-36	34 (27%)	38 (35%)				
≥37*	49 (38%)	20 (18%)				
Haplotype						
TAT	109 (86%)	91 (88%)	0.1	0.713	1.25 (0.57-2.7)	0.573
AGC*	18 (14%)	12 (12%)				

Data are reported as number with percent in parentheses. HIE: hypoxic-ischemic encephalopathy; OR: odds ratio; CI: confidence interval. * Reference value. The χ^2 test and logistic regression were used for analyses.

the presence of the T allele seems to indicate a susceptibility for greater brain injury, and a lower Apgar score at birth.

Polymorphism rs1808593 is located in the twenty-third intron of the NOS3 gene. Intron polymorphisms may have a potential effect on gene transcription or mRNA stability, and thus affect gene expression, and consequently, enzyme function (21). Yoon et al. (22) showed an association of the NOS3 gene polymorphism rs1808593 with NO plasma concentration in patients with coronary artery disease. Association of G894T polymorphism (rs1799983) and 10G/T intron-23 (rs1808593) polymorphism and their haplotype combination with childhood hypertension, especially in females, were also shown (23). On the other hand, the AA genotype of polymorphism rs3918186 was twice as prevalent among children with Apgar scores of 10, which might indicate its protective function in brain damage.

This study also found an association of low Apgar scores <5 at 5 min after birth with hypoxic-ischemic injury, which is in line with previous findings (24). A recently published study showed that the prevalence of cerebral palsy in children with an Apgar score <3 was 130-times higher than in children with a score of 10 (25). Similar observations have been reported in children with extremely low birth weights (<1000 g), who were more likely to develop HIE than children with higher birth weights. It should be emphasized that the children of low birth weight are generally premature, as gestational age and birth weight are closely correlated (26).

We also found that boys had a higher risk of HIE than girls, which is consistent with the observations of other authors. Thorngren-Jerneck et al. (27) also reported that boys have a higher risk of developing cerebral palsy than girls (1.36:1), especially if they were born prematurely

Table 5. Magnetic resonance imaging findings according to the gestational age of children with hypoxic-ischemic encephalopathy.

	GA <32	GA 32-36	GA ≥37	Total	χ^2	P
MRI grade 2	13 (34%)	15 (40%)	10 (26%)	38 (100%)	4.614	0.099
MRI grade 3	39 (54%)	23 (32%)	10 (14%)	72 (100%)		
Total	52 (47%)	38 (35%)	20 (18%)	110 (100%)		

Data are reported as number with percent in parentheses. GA: gestational age; MRI: magnetic resonance imaging. The χ^2 test was used for analyses.

Table 6. Magnetic resonance imaging findings according to the birth weight in children with hypoxic-ischemic encephalopathy.

	≤1000 g	1001-1499 g	1500-2499 g	≥2500 g	Total	χ^2	P
MRI grade 2	4 (10%)	8 (21%)	12 (32%)	14 (37%)	38 (100%)	4.15	0.246
MRI grade 3	10 (14%)	20 (28%)	30 (42%)	12 (16%)	72 (100%)		
Total	14 (13%)	28 (25%)	42 (38%)	26 (24%)	110 (100%)		

Data are reported as number with percent in parentheses. MRI: magnetic resonance imaging. The χ^2 test was used for analyses.

(1.55:1). Another epidemiological study showed up to 30% higher incidence of cerebral palsy among boys than girls. The risk of developing cerebral palsy increased 4-fold for infants in the ninety-seventh percentile of birth weight and 16 times for male infants in the third percentile (25,27). Recent studies in mice suggest that the differences between males and females in signaling pathways that regulate apoptosis may at least partly explain this epidemiological observation. Males may be more vulnerable to glutamate-mediated excitotoxicity leading to poly [ADP-ribose] polymerase (PARP)-1 activation, which causes apoptosis. One possible clue to the mechanism was the finding that levels of the energy intermediate NAD⁺, which is consumed by PARP-1 activation, were reduced more in males than in females (28). Finally, we compared the relationship of brain damage (defined by MRI and clinical contributors), and found that children of lower gestational age and lower birth weight had a 90% greater chance of developing severe brain damage.

References

1. Perlman JM. Summary proceedings from the neurology group on hypoxic-ischemic encephalopathy. *Pediatrics* 2006; 117: S28-S33, doi: 10.1542/peds.2005-2425.
2. Graham EM, Ruis KA, Hartman AL, Northington FJ, Fox HE. A systematic review of the role of intrapartum hypoxia-ischemia in the causation of neonatal encephalopathy. *Am J Obstet Gynecol* 2008; 199: 587-595, doi: 10.1016/j.ajog.2008.06.094.
3. Jensen FE. Developmental factors regulating susceptibility to perinatal brain injury and seizures. *Curr Opin Pediatr* 2006; 18: 628-633, doi: 10.1097/MOP.0b013e328010c536.
4. Reed JC. Mechanisms of apoptosis. *Am J Pathol* 2000; 157: 1415-1430, doi: 10.1016/S0002-9440(10)64779-7.
5. Blomgren K, Hagberg H. Free radicals, mitochondria, and hypoxia-ischemia in the developing brain. *Free Radic Biol Med* 2006; 40: 388-397, doi: 10.1016/j.freeradbiomed.2005.08.040.
6. Haynes RL, Baud O, Li J, Kinney HC, Volpe JJ, Folkert DR. Oxidative and nitrative injury in periventricular leukomalacia: a review. *Brain Pathol* 2005; 15: 225-233, doi: 10.1111/j.1750-3639.2005.tb00525.x.
7. Khwaja O, Volpe JJ. Pathogenesis of cerebral white matter injury of prematurity. *Arch Dis Child Fetal Neonatal Ed* 2008; 93: F153-F161, doi: 10.1136/adc.2006.108837.
8. Albrecht EW, Stegeman CA, Heeringa P, Henning RH, van Goor H. Protective role of endothelial nitric oxide synthase. *J Pathol* 2003; 199: 8-17, doi: 10.1002/path.1250.
9. Baier RJ. Genetics of perinatal brain injury in the preterm infant. *Front Biosci* 2006; 11: 1371-1387, doi: 10.2741/1890.
10. Casas JP, Cavalleri GL, Bautista LE, Smeeth L, Humphries SE, Hingorani AD. Endothelial nitric oxide synthase gene polymorphisms and cardiovascular disease: a HuGE review. *Am J Epidemiol* 2006; 164: 921-935, doi: 10.1093/aje/kwj302.
11. Howard TD, Giles WH, Xu J, Wozniak MA, Malarcher AM, Lange LA, et al. Promoter polymorphisms in the nitric oxide synthase 3 gene are associated with ischemic stroke susceptibility in young black women. *Stroke* 2005; 36: 1848-1851, doi: 10.1161/01.STR.0000177978.97428.53.
12. Kuzmanić Samija R, Primorac D, Resić B, Lozić B, Krzelj V, Tomasović M, et al. Association of NOS3 tag polymorphisms with hypoxic-ischemic encephalopathy. *Croat Med J* 2011; 52: 396-402, doi: 10.3325/cmj.2011.52.396.
13. Sarnat HB, Sarnat MS. Neonatal encephalopathy following fetal distress. A clinical and electroencephalographic study. *Arch Neurol* 1976; 33: 696-705, doi: 10.1001/archneur.1976.00500100030012.
14. Weisglas-Kuperus N, Baerts W, Fetter WP, Sauer PJ. Neonatal cerebral ultrasound, neonatal neurology and perinatal conditions as predictors of neurodevelopmental outcome in very low birthweight infants. *Early Hum Dev* 1992; 31: 131-148, doi: 10.1016/0378-3782(92)90040-N.
15. Benjak V, Culjat M, Pavlovic M, Kostovic-Szrentic M.

- Changes of the corpus callosum in children who suffered perinatal injury of the periventricular crossroads of pathways. *Coll Antropol* 2008; 32 (Suppl 1): 25-29.
16. Jovanov-Milosevic N, Culjat M, Kostovic I. Growth of the human corpus callosum: modular and laminar morphogenetic zones. *Front Neuroanat* 2009; 3: 6.
 17. Gauderman WJ. Sample size requirements for matched case-control studies of gene-environment interaction. *Stat Med* 2002; 21: 35-50, doi: 10.1002/sim.973.
 18. Kapitanovic Vidak H, Catela Ivkovic T, Jokic M, Spaventi R, Kapitanovic S. The association between proinflammatory cytokine polymorphisms and cerebral palsy in very preterm infants. *Cytokine* 2012; 58: 57-64, doi: 10.1016/j.cyto.2011.12.018.
 19. Nelson KB, Dambrosia JM, Iovannisci DM, Cheng S, Grether JK, Lammer E. Genetic polymorphisms and cerebral palsy in very preterm infants. *Pediatr Res* 2005; 57: 494-499, doi: 10.1203/01.PDR.0000156477.00386.E7.
 20. Wang J, Dudley D, Wang XL. Haplotype-specific effects on endothelial NO synthase promoter efficiency: modifiable by cigarette smoking. *Arterioscler Thromb Vasc Biol* 2002; 22: e1-e4, doi: 10.1161/01.ATV.0000016248.51577.1F.
 21. Chorley BN, Wang X, Campbell MR, Pittman GS, Noureddine MA, Bell DA. Discovery and verification of functional single nucleotide polymorphisms in regulatory genomic regions: current and developing technologies. *Mutat Res* 2008; 659: 147-157, doi: 10.1016/j.mrrev.2008.05.001.
 22. Yoon Y, Song J, Hong SH, Kim JQ. Plasma nitric oxide concentrations and nitric oxide synthase gene polymorphisms in coronary artery disease. *Clin Chem* 2000; 46: 1626-1630.
 23. Chen W, Srinivasan SR, Li S, Boerwinkle E, Berenson GS. Gender-specific influence of NO synthase gene on blood pressure since childhood: the Bogalusa Heart Study. *Hypertension* 2004; 44: 668-673, doi: 10.1161/01.HYP.0000145474.23750.2b.
 24. American Academy of Pediatrics. The Apgar score. *Pediatrics* 2006; 117: 1444-1447, doi: 10.1542/peds.2006-0325.
 25. Lie KK, Groholt EK, Eskild A. Association of cerebral palsy with Apgar score in low and normal birthweight infants: population based cohort study. *BMJ* 2010; 341: c4990, doi: 10.1136/bmj.c4990.
 26. Hintz SR, Kendrick DE, Vohr BR, Kennedy PW, Higgins RD. Gender differences in neurodevelopmental outcomes among extremely preterm, extremely-low-birthweight infants. *Acta Paediatr* 2006; 95: 1239-1248, doi: 10.1080/08035250600599727.
 27. Thorngren-Jerneck K, Herbst A. Perinatal factors associated with cerebral palsy in children born in Sweden. *Obstet Gynecol* 2006; 108: 1499-1505, doi: 10.1097/01.AOG.0000247174.27979.6b.
 28. Johnston MV, Hagberg H. Sex and the pathogenesis of cerebral palsy. *Dev Med Child Neurol* 2007; 49: 74-78, doi: 10.1017/S0012162207000199.x.