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Selection of coffee progenies for resistance to nematode *Meloidogyne paranaensis* in infested area

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Abstract – The purpose of this study was to select *Coffea arabica* progenies for resistance to *M. paranaensis* in an infested coffee growing area using Henderson's mixed model methodology. Forty-one genotypes were selected at the Coffee Active Germplasm Bank of Minas Gerais, and evaluated in regard to stem diameter, number of plagiotropic branches, reaction to the nematode, and yield per plant. There was genetic variability among the genotypes studied for all the traits evaluated, and among the populations studied for yield and reaction to the nematode, indicating possibilities for obtaining genetic gains through selection in this population. There was high rate of genotypic association between all the traits studied. Coffee plants of Timor Hybrid UFV408-01 population, and F₃ progenies derived from crossing Catuai Vermelho and Amphillo MR 2161 were the most promising in the area infested by *M. paranaensis*.

Key words: *Coffea arabica*, breeding, additive genetic value, correlation, root knot nematode.

INTRODUCTION

Among the factors that limit the growth and production process of coffee plant, species of *Meloidogyne* stand out. Economic losses due to root knot nematodes vary considerably depending on the species involved and their distribution (Boisseau et al. 2009). *M. paranaensis* is considered more harmful to coffee plant due to high root damage, and may lead to plant death. In face of the occurrence of *M. paranaensis* in coffee crops in the state of Minas Gerais (Castro et al. 2008), and the risks of its spread, the identification of genotypes that are resistant and adapted in infested areas is of great importance in a breeding program.

The resistance mechanism of the plant prevents the nematode from developing and/or leads to low reproduction rates. Consequently, in addition to the parasite not causing damage to the crop, the use of genetic resistance also leads to reduction of nematode population density in the soil, and the possibility of economic maintenance of the crop in infested areas.

Some interspecific hybrids between *C. arabica* and *C. canephora* were resistant to *M. paranaensis*, but still seg-

regate this resistance (Mata et al. 2002, Sera et al. 2004). Advances in research are made difficult by the perennial condition of this crop and the period of time required for evaluating the behavior of plants derived from sources of *Coffea* spp. germplasm in an infested area. In fact, under Brazilian conditions, little research has been carried out to study the performance of resistant plants under field conditions. On the other hand, many studies (Ribeiro et al. 2005, Sera et al. 2007, Sera et al. 2009, Ito et al. 2008, Boisseau et al. 2009) have been carried out under greenhouse conditions. Nevertheless, evaluation of genotypes of coffee plant under the conditions of an infested area in the field allows better knowledge of plant behavior and verifies the stability of plant reaction (Alpizar et al. 2007).

In this case, the use of accurate selection procedures becomes essential. Progeny testing has been used in estimating genetic parameters and selection of individuals when one seeks to evaluate the magnitude and nature of genetic variance available with a view toward quantifying, and maximizing genetic gains using adequate selection procedures (Costa et al. 2010).

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Nevertheless, selection based on inadequate biometric procedures may lead to confounding between genotypic and environmental effects, resulting in low efficiency in selection. The mixed models methodology (Henderson 1984) is a flexible procedure for obtaining estimates of genetic parameters and prediction of breeding values, especially under imbalanced conditions, maximizing genetic gains from selection (Furlani et al. 2005, Chiorato et al. 2008).

The purpose of this study was to select superior *Coffea arabica* progenies in regard to resistance to *M. paranaensis* in an infested area using the Henderson's mixed models methodology.

MATERIAL AND METHODS

Progenies in generation F₃ and F₄, developed in the Coffee Genetic Breeding Program in Minas Gerais, coordinated by EPAMIG (Crop and Livestock Research Company of Minas Gerais), were selected in the Active Germplasm Bank maintained at the EPAMIG Experimental Farm in the municipality of Patrocínio/MG. Forty-four genotypes were evaluated in the experiment (Table 1). Forty-one progenies belonging to 20 populations, and three *Coffea arabica* cultivars checks, (i.e. Mundo Novo 379/19 and

Catuai IAC-64 as susceptible checks and the *C. arabica* cv. IPR 100 as resistant check) were evaluated (Table 1).

The experiment was carried out in an infested area by *Meloidogyne paranaensis*, located in the municipality of Piumhi (20° 25' 28.7" S, 46° 1' 10.5" W, 812m asl), MG, with annual average temperature of 20.7 °C, annual average rainfall of 1426.3 mm, soil with clayey texture and flat terrain. Identification of the species *M. paranaensis* in the infested area was carried out using the Carneiro and Almeida (2001) technique.

A randomized complete block design with three replications and spacing of 3.00 x 0.80m was used, with plots consisting of seven plants. Crop treatments performed were those recommended for the coffee crop in the region, with the exception of application of chemical products for nematode control.

Soil samples were collected in the rhizosphere of coffee plants at two months after sowing. In a greenhouse, samples of approximately 2000 g from each experimental plot were distributed in three-liter capacity pots where two Santa Clara cv. tomato seedlings were planted for biological indicator test of *M. paranaensis* population in the soil of the plots. Evaluation of the test was carried out by means

Table 1. Numerical identification (NI) and generation (G) of progenies originating from the Coffee Active Germplasm Bank at the EPAMIG Coffee Plant Breeding Program, and check cultivars of *Coffea arabica* evaluated

| NI | G | Accession | NI | G | Accession |
|----|----------------|---|----|----------------|---|
| 1 | F ₄ | H. T. UFV 408-28 pl. 10 | 23 | F ₄ | H. T. UFV 408-28 pl.2 |
| 2 | F ₄ | H.T. UFV 408-29 pl. 2 R1 | 24 | F ₃ | Atypical plant ¹ pl.6 Line 2 |
| 3 | F ₃ | Atypical plant ¹ pl. 34 Line 3 | 25 | - | IPR 100* |
| 4 | F ₄ | Amphillo x H.N. 36-349 pl.2 R2 | 26 | F ₄ | Amphillo x H. N. 36-352 pl.8 R1 |
| 5 | F ₄ | H. T. UFV 408-01 pl.1 R1 | 27 | F ₄ | Amphillo x H. N.36-349 pl.9 R2 |
| 6 | F ₄ | C.V. x Amphillo 2474 pl.10 R2 | 28 | F ₃ | Atypical plant ¹ pl.37 Line 3 |
| 7 | F ₄ | H. T. UFV 408-26 pl.8 R1 | 29 | F ₄ | Durandé Arabica x <i>C. canephora</i> pl.3 R1 |
| 8 | F ₄ | C.V. x Amphillo 2161pl.5 R1 | 30 | F ₄ | Amphillo x H. N. 36-349 pl.10 R2 |
| 9 | F ₄ | H. T. UFV 408-01 pl.2 R1 | 31 | F ₄ | H. T. UFV 408-12 pl.2 R1 |
| 10 | F ₄ | H. T. UFV 408-26pl.4 R1 | 32 | F ₄ | Amphillo x H. N. 36-349 pl.3 R2 |
| 11 | F ₄ | Amphillo x H.N. 36-352 pl.7 R2 | 33 | F ₄ | C. V. x Amphillo 2474 pl.5 R2 |
| 12 | F ₃ | Atypical plant ¹ pl.36 Line 4 | 34 | F ₄ | Sarchimor pl.1 R1 |
| 13 | F ₃ | Atypical plant ¹ pl.10 Bordad. | 35 | F ₄ | H. T. UFV 408-12 pl.1 R1 |
| 14 | F ₃ | Atypical plant ¹ pl.8 R1 | 36 | F ₃ | Atypical plant ¹ pl.8 Line 3 |
| 15 | F ₄ | C.V. x Amphillo 2161 pl.1 R1 | 37 | F ₄ | H. T. UFV 408-11 pl.9 R1 |
| 16 | F ₃ | Atypical plant ¹ pl.10 Line 3 | 38 | F ₄ | Amphillo x H. N. 36-352 pl.1 R1 |
| 17 | F ₄ | Amphillo x H. N. 36-352 pl.1 R2 | 39 | F ₄ | Amphillo x H. N. 36-352 pl.4 R2 |
| 18 | F ₄ | H. T. UFV 408-11 pl.4 R1 | 40 | F ₄ | Icatu x Catimor F5 H 32-11-17-4-2 |
| 19 | F ₄ | H. T. UFV 376-52 pl.7 R1 | 41 | F ₄ | Icatu x Catimor F5 H 29-1-8-5-4 |
| 20 | F ₄ | C. V. x Amphillo 2474 pl.3 R2 | 42 | F ₄ | Icatu x Catimor F5 H 136-1-13-15-3 |
| 21 | F ₄ | C. V. x Amphillo 2161pl.3 R1 | 43 | - | Catuai IAC 64* |
| 22 | F ₄ | C. V. X Amphillo 2474 pl.1 R2 | 44 | - | Mundo Novo 379-19* |

¹Atypical plant = natural cross between *Coffea arabica* x diploid species. *Check cultivars. H.N. = Natural Hybrid.

C.V. = Catuai Vermelho. H.T. = Timor Hybrid.

of quantification of the number of galls and eggs of *M. paranaensis* in the roots of tomato plants at 70 days after planting. These data were submitted to analysis of variance with the F test at 5% of significance (Table 2).

At 18 months from planting, the following traits were evaluated: stem diameter, number of plagiotropic branches, reaction of the genotypes to the nematode, and yield in liters of fresh coffee fruits per plant in the first harvest of 2011. Stem diameter was measured at 10 cm above the base of the stem, in millimeters, with the aid of a digital caliper rule. The number of plagiotropic branches was calculated by counting of all the primary lateral branches with a size greater than 5 cm. The reaction of genotypes to the nematode was performed following the scale adapted by Carneiro (1995), considering only the above ground part of the plants. On this scale, extremely debilitated or dead plants were given a score 0. Score 1 was given to plants with severe leaf loss aspect and few leaves; 2 for plants with moderate leaf loss aspect, with small, malformed and stopped leaves, with typical symptoms of nutritional deficiency and accentuated shedding of leaves; 3 for plants of good vigor, with no leaf loss aspect and yellowing, and 4 for plants with great vigor, with no symptoms of parasitic activity. Values less than 3 indicate symptoms of parasite activity by nematodes.

Data were analyzed according to the following mixed linear model in the matrix form (Resende 2002)

$$y = Xr + Za + Wp + Ts + e,$$

where: y : data vector; r : vector of the fixed effects of blocks added to the general average; a : vector of the individual additive genetic effects, $a \sim N(0, A\sigma_a^2)$, being A the additive relationship matrix, and σ_a^2 the additive variance; p : vector of the plot effects, $p \sim N(0, I\sigma_p^2)$, being I the identity matrix, and σ_p^2 the variance among plots; s : vector of the population effects, $s \sim N(0, I\sigma_s^2)$, being σ_s^2 the variance among populations; e : vector of residual errors, $e \sim N(0, I\sigma_e^2)$, being σ_e^2 the residual variance; X , Z , W and T : incidence matrices for r , a , p and s effects, respectively.

Estimates of genetic parameters were obtained by the REML/BLUP (Restricted Maximum Likelihood / Best Linear Unbiased Prediction) procedure with the aid of the

software SELEGEN-REML/BLUP (Resende 2007).

Based on estimates of the components of variance, it was estimated selective accuracies, individual heritabilities and other coefficients of determination associated with the random effects of the model, as well as the genetic, environmental and relative variation coefficients, as described in Resende (2002). Variance components were submitted to likelihood ratio test at 5% probability (Resende 2007). Genotypic correlations between traits were obtained by Genes software (Cruz 2006).

RESULTS AND DISCUSSION

All the plots showed a high and statistically homogeneous dispersion of *M. paranaensis*, which was expressed by the number of galls and eggs per root system, and number of eggs/g of tomato root (Table 2). This fact is essential for evaluating progenies in field experiments when the aim is centered in measuring the resistance to nematodes.

The likelihood ratio test revealed the existence of genetic variability among the genotypes studied for all the traits evaluated. In the same way, variability was detected among the populations studied for the fresh fruit yield and reaction of genotypes to the nematode traits (Table 3). The estimates of genetic and phenotypic parameters for the traits evaluated were compatible with other studies in coffee progenies (Freitas et al. 2007, Petek et al. 2008).

All the traits studied were highly influenced by the environment. The individual heritability values in the narrow sense were low, ranging from 4.84 to 14.12% (Table 3). The observed values of the coefficient of determination ranged from 7.14% to 21.28% due to the common environment of the plot that quantifies the environmental variability within the plot, which are compatible with experiments considered as precise (Resende et al. 2001, Freitas et al. 2007).

The relative coefficient of variation presented median values for the number of plagiotropic branches and fresh fruit yield (0.81 and 0.92, respectively) showing that selection of the best progenies may increase the genetic value of the population possibly in regard to these traits (Vencovsky 1987). The other traits showed low values for the relative

Table 2. Summary of analysis of variance of the biological indicator test in the area infested by *Meloidogyne paranaensis*

| Source of Variation | df | Mean Square | | |
|---------------------|----|------------------------|-----------------------|------------------------|
| | | Number of eggs | Number of galls | Number of eggs/g root |
| Blocks | 2 | - | - | - |
| Genotypes | 43 | 39204622 ^{ns} | 28920.4 ^{ns} | 302598.5 ^{ns} |
| Error | 86 | 36521472 | 29124.778 | 291323.89 |
| Mean | | 6033 | 227 | 553 |

^{ns} not significant by the F test (5%).

Table 3. Estimates of genetic and phenotypic parameters related to reaction of the genotypes to the nematode (NEM, grade), stem diameter (SD, mm), number of plagiotropic branches (NPB), and fresh coffee fruits yield in the first harvest (YIELD, liter plant⁻¹), through evaluation of coffee plant progenies from different populations at 18 months after planting

| Parameter | NEM | SD | NPB | YIELD |
|-----------------|----------|----------|---------|----------|
| σ_a^2 | 0.0552** | 3.0195** | 6.7091* | 0.3014** |
| σ_p^2 | 0.2427** | 5.9118** | 5.4737* | 0.1523* |
| σ_s^2 | 0.1121** | 3.4105 | 3.3218 | 0.2708* |
| h_a^2 | 0.0484 | 0.0854 | 0.1371 | 0.1412 |
| c_p^2 | 0.2128 | 0.1672 | 0.1118 | 0.0714 |
| CV _r | 0.3991 | 0.5729 | 0.8090 | 0.9232 |
| General mean | 2.2206 | 13.7014 | 16.1179 | 0.6892 |

** Significant at 1% and at 5% respectively by the likelihood ratio test. Additive genetic variance (σ_a^2), environmental variance among plots (σ_p^2), genetic variance among populations (σ_s^2), individual heritability in the narrow sense (h_a^2), coefficient of determination of the plot effects (c_p^2), and relative coefficient of variation (CV_r).

coefficient of variation (0.40 and 0.57, respectively) manifesting the difficulty of selecting superior plants based on the reaction of genotypes to the nematode trait and stem diameter at 18 months after planting (Table 3).

Number of plagiotropic branches and yield were chosen as major characteristics for selection of individual plants, once these characteristics have shown higher coefficient of relative variation. Also productivity is a major criterion for coffee plants selection (Gichimu and Omondi 2010, Cilas et al. 2011). Another selection criterion was the reaction to nematode, which was determined in plants at 18 months after planting, aiming the identification of nematode resistant plants.

Genetic gain is inversely proportional to the intensity of selection, which quantifies the number of selected individuals (Cruz and Carneiro 2003). Therefore, in this study, the need for working with a greater number of individuals (selection intensity of 5%) was considered to ensure an effective size that, according to Rocha et al. (2009), allows greater efficiency in the following steps of selection.

Predicted mean with the selection of the 20 individuals (Table 4) based on fresh fruit yield was 2.21, being this value 125% greater than the general mean value in the yield of all the plants evaluated in the experiment. Plants with greater yield also proved to have greater reaction to nematode grades. Therefore, the reaction of these plants may be resistance or tolerance to *M. paranaensis* since, according to Roberts (2002), resistant plants affect diverse phases of the life cycle and of parasitism of the nematode, restricting or preventing its multiplication, while plants considered to be tolerant allow multiplication of the nematode in their roots without harming yield. It is worth highlighting that in taking into account the initial development of plants, root samples were not collected for quantification of *M. paranaensis* population.

It was observed that the individuals of progeny 15 (Catuaí Vermelho x Amphillo 2161 pl.1 R1) were predominant among the most promising, with expressive predicted additive genetic values ranging from 2.22 to 2.60 for the reaction to nematode, from 0.68 to 2.21 L plant⁻¹ for yield, and from 16.11 to 19.92 for number of plagiotropic branches, and constitute 13 of the 20 best classified genetic materials, suggesting good possibilities for genetic progress in the sequence of evaluations in more advanced generations.

Progeny 15, as well as progenies 21 and 22, from which were selected two and one plants, respectively, refers to the F₄ generation of the population derived from the cross between Catuaí Vermelho and Amphillo MR 2161. In the same way, Gonçalves et al. (1996) observed partial resistance of the variety Amphillo to race 2 of *M. incognita*; however, for *M. paranaensis*, this genotype had not yet been studied. Plant six, from the third block of progeny 15 stood out with the yield of 7.00 L plant⁻¹, a nematode grade of 4.00, and 28 plagiotropic branches with predicted additive genetic values of 2.63, 2.61 and 19.98, respectively.

Another promising progeny, of which four of the 20 best classified plants were selected, was the progeny 5, which refers to the Timor Hybrid UFV 408-01. This interspecific hybrid derived from a possible spontaneous crossing of *Coffea arabica* L. and *Coffea canephora* Pierre ex A. Froehner constitutes a source of genetic diversity for the development of new cultivars, which have presented promising yields, together with resistance to the agent which causes rust and to the nematode *M. exigua* (Ribeiro et al. 2005, Carneiro et al. 2008).

With the selection of the 20 best individuals, the initial yield of the population of 0.68 L plant⁻¹ (Table 3) would increase after one selection cycle to 2.21 L plant⁻¹ (Table 4). Considering an average yield of 480 liters of fresh coffee fruit for each 60 kg bag of processed coffee, yield would

increase from 4.05 to 13.15 bags ha⁻¹. These results denote that the selection of plants in this population in an area infested by *M. paranaensis* is quite promising.

In this study, resistance or tolerance reaction to the *M. paranaensis* nematode was observed in *C. arabica* coffee plants derived from Catuaí Vermelho x Amphillo MR 2161 and Timor Hybrid UFV 408-01. In other studies, sources of resistance were also identified, including some cultivars being identified as resistant to *M. paranaensis*. Arabica coffee cultivars IPR 100 (“Catindú”) and IPR 106 (“Icatu”),

both developed by IAPAR and susceptible to rust (Sera et al. 2010), show simultaneous resistance to nematodes *M. paranaensis* and some races of *M. incognita* (Sera et al. 2007, Sera et al. 2009, Ito et al. 2008, Kanayama et al. 2009). ‘Icatu Vermelho IAC 3888’ (Gonçalves and Silvarolla 2007) and other “Icatu” progenies (Mata et al. 2002, Sera et al. 2004) were also resistant to *M. paranaensis*. Selections from “Icatu”, such as line 925, have shown good resistance to *M. paranaensis* (Matiello et al. 2010). In some accessions of *C. arabica* from Ethiopia, resistance to *M. paranaensis* was observed (Anthony et al. 2003, Boisseau et al. 2009).

Table 4. Estimates of mean components: Individual phenotypic value (f); predicted additive genetic value ($\hat{u}+\hat{a}$); predicted additive genetic gain and predicted bred mean value of the 20 best individuals of the experiment in 41 progenies of *Coffea arabica*, in an area infested by *M. paranaensis*, selected for the fresh fruit yield of the first harvest (YIELD, liter/plant⁻¹); reaction of the genotypes to the nematode (NEM, grade); and number of plagiotropic branches (NPB)

| Progeny | Block | Plant | NPB | | NEM | | YIELD | |
|---------------------------------|-------|-------|-------|-------------------|------|-------------------|-------|-------------------|
| | | | f | $\hat{u}+\hat{a}$ | f | $\hat{u}+\hat{a}$ | f | $\hat{u}+\hat{a}$ |
| 15 | 3 | 6 | 28 | 19.98 | 4 | 2.61 | 7.0 | 2.63 |
| 15 | 1 | 6 | 27 | 19.97 | 4 | 2.61 | 5.9 | 2.54 |
| 15 | 3 | 5 | 24 | 19.60 | 4 | 2.61 | 6.0 | 2.53 |
| 15 | 1 | 7 | 24 | 19.68 | 4 | 2.61 | 4.5 | 2.39 |
| 15 | 1 | 5 | 23 | 19.59 | 4 | 2.61 | 4.4 | 2.38 |
| 15 | 2 | 6 | 20 | 19.42 | 3 | 2.61 | 3.0 | 2.28 |
| 5 | 3 | 7 | 28 | 22.22 | 4 | 2.86 | 6.0 | 2.25 |
| 5 | 2 | 6 | 28 | 22.23 | 4 | 2.87 | 6.0 | 2.22 |
| 21 | 2 | 6 | 27 | 18.56 | 4 | 2.61 | 7.0 | 2.18 |
| 15 | 2 | 4 | 23 | 19.71 | 3 | 2.61 | 2.0 | 2.17 |
| 15 | 1 | 3 | 20 | 19.3 | 3 | 2.59 | 2.25 | 2.15 |
| 5 | 3 | 2 | 27 | 22.12 | 4 | 2.86 | 5.0 | 2.14 |
| 15 | 3 | 4 | 26 | 19.79 | 3 | 2.59 | 2.4 | 2.14 |
| 15 | 1 | 4 | 22 | 19.49 | 4 | 2.61 | 2.1 | 2.14 |
| 5 | 1 | 7 | 33 | 22.53 | 4 | 2.87 | 4.0 | 2.06 |
| 15 | 3 | 3 | 19 | 19.13 | 3 | 2.59 | 1.5 | 2.05 |
| 15 | 2 | 5 | 19 | 19.33 | 1 | 2.55 | 0.5 | 2.02 |
| 15 | 2 | 3 | 18 | 19.24 | 2 | 2.58 | 0.4 | 2.01 |
| 21 | 1 | 5 | 23 | 18.24 | 4 | 2.61 | 5.0 | 1.99 |
| 22 | 1 | 2 | 26 | 18.20 | 4 | 2.60 | 5.5 | 1.94 |
| ¹ New mean value | | | 24.25 | 19.92 | 3.50 | 2.60 | 4.02 | 2.21 |
| ² Predicted gain (%) | | | | 23.64 | | 17.11 | | 125 |
| Mean value of the progenies | | | | 16.11 | | 2.22 | | 0.68 |
| Mean value of IPR 100 | | | | 23.25 | | 2.89 | | 3.04 |
| Mean value of ‘Catuaí 64’ | | | | 15.1 | | 2.11 | | 0.41 |
| Mean value of ‘Mundo Novo’ | | | | 14.89 | | 2.05 | | 0.41 |

¹ Mean value of selected plants, ² Predicted additive genetic gain (%).

Table 5. Genotypic correlation among the traits fresh fruit yield of the first harvest (liter plant⁻¹), stem diameter (mm) (SD), number of plagiotropic branches (NPB) and reaction of the genotypes to the nematode (grade) (NEM) evaluated in coffee plant progenies at 18 months after planting

| Traits | YIELD | SD | NPB | NEM |
|--------|-------|--------|--------|--------|
| YIELD | 1.00 | 0.82** | 0.78** | 0.78** |
| SD | | 1.00 | 0.87** | 0.94** |
| NPB | | | 1.00 | 0.79** |
| NEM | | | | 1.00 |

** Significant at 1% of probability by bootstrap sampling.

Some authors emphasize the need for use of special methods of prediction of genetic values in relation to all the candidate individuals as the best strategy for increasing the efficiency of coffee breeding (Resende et al. 2001, Petek et al. 2008). In methodological terms, the great utility and flexibility of mixed models method should be stressed, which has made a complete estimate and prediction possible, in a situation of great imbalance of data since 57.25% of the plants were not able to survive in an infested area by *M. paranaensis*. Four hundred seedlings were initially planted, and only 229 plants were evaluated. This leads to maximization of genetic gain through selection, allowing the genetic breeding program to be carried out with a view toward resistance to *M. paranaensis*.

For the purpose of knowing the changes which occurred in the traits studied, involving associations of an inherited nature so that they could be used in guiding the breeding program with a view toward resistance to *M. paranaensis*, the genotypic correlation among the traits studied was estimated (Table 5). It is observed that the correlations among all the traits studied were positive, significant and of high magnitude, ranging from 0.78 to 0.94. The significant positive correlation between the first yield, stem diameter and number of plagiotropic branches in coffee plants were

reported by other authors (Bonomo et al. 2004, Martinez et al. 2007, Carvalho et al. 2010).

Characterization of the reaction of the genotypes to the nematode by grades also showed positive correlation with yield. This indicates the possibility of obtaining genetic gains through selection based on symptomatological characterization of plants in field conditions, and valid for application of the grading scale, which was used by Carneiro (1995) to evaluate coffee plants at 36 months from planting in an area infested by *M. incognita*. Thus, the results confirm the usefulness of this scale also for coffee plants in initial development, in an area infested by *M. paranaensis*, constituting a tool for early evaluation of plant reaction to the nematode.

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Seleção de progênies de café para a resistência ao nematoide *Meloidogyne paranaensis* em área infestada

Resumo – *Objetivou-se neste trabalho selecionar progênies de Coffea arabica para resistência à M. paranaensis em área cafeeira infestada usando a abordagem de modelos mistos de Henderson. Quarenta e um genótipos foram selecionados no Banco Ativo de Germoplasma de Café de Minas Gerais e avaliados individualmente quanto ao diâmetro de caule, número de ramos plagiotrópicos, caracterização da reação dos genótipos ao nematoide e produção por planta. Houve variabilidade genética entre os genótipos estudados para todas as características avaliadas, e entre as populações estudadas para as características produção e caracterização da reação dos genótipos ao nematoide, indicando possibilidades de obtenção de ganhos genéticos pela seleção nesta população. As correlações genóticas entre todas as características estudadas foram significativas. Cafeeiros da população de Híbrido de Timor UFV 408-01 e progênies F₃ derivadas do cruzamento entre Catuaí Vermelho e Amphillo MR 2161 foram os mais promissores em área infestada por M. paranaensis.*

Palavras-chave: Coffea arabica, melhoramento, valor genético aditivo, correlação, nematoide das galhas.

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