

Effect of environmental factors on the fruit essential oils of *Pistacia terebinthus* L. growing wild in Turkey

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SILVICULTURE

ABSTRACT

Background: The aim of this study was to investigate the effect of environmental factors on the essential oil yields (EOYs) and major essential oil components in the ripe fruits of *Pistacia terebinthus* L. (turpentine tree), which is a medicinal and aromatic plant. Fruit samples were collected from 34 different locations of the turpentine tree growing wild in the Lake District in the Mediterranean Region of Turkey. EOYs (% v/w) and essential oil component ratios (%) were determined in fruit samples by hydrodistillation and GC-MS methods, respectively. The effect of environmental factors on these variables were analyzed using Pearson Correlation and Principal Component Analysis.

Results: The essential oil yield of fruits collected from different localities ranged from 0.05% (v/w) to 0.19% (v/w). In terms of average values, α -pinene with 41.01% and limonene with 14.28% were the major components in the essential oils of ripe fruits. Among the environmental variables, longitude and sand % in 10-30 cm were the variables that made a statistically significant difference ($p < 0.05$) in fruit essential oil yield. According to the results of principal components analysis (PCA), α -pinene and Ocimene components were most affected by silt (% , 10-30 cm) and pH (% , 30-50 cm) ratios. The longitude and total annual precipitation (mm) were the most determining variables in the sabinene and limonene components, respectively. It was seen that the total lime ratios at different depth levels of the soils taken from the localities were the most effective variables for Myrcene and p-Cymene components.

Conclusion: The study findings showed that EOYs and intended volatile component ratios in ripe fruits of turpentine trees grown under appropriate environmental conditions can be increased.

Keywords: Essential oil, limonene, locations, ripe fruits, turpentine tree

HIGHLIGHTS

Essential oil yields of the turpentine tree fruits varied in different locations.

The most abundant component in EOYs of the turpentine tree fruits was α -pinene.

Silt ratio (%) and pH in soil, total annual precipitation (mm) and longitude presented the most affective relations to chemical composition of EOYs of the turpentine tree fruits.

In certain environmental conditions, higher EOYs and targeted chemical compounds can be obtained in plants.

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INTRODUCTION

The Anacardiaceae family includes 81 genera and more than 800 species (Amiri and Mohammadi, 2021). The genus *Pistacia* in this family is classified as 11 species in 4 subsections (Zohary, 1952; Whitehouse, 1957). The *Pistacia* species with the widest distribution in forest areas in Turkey is *P. terebinthus* (Davis, 1967). The *P. terebinthus* species, known as the turpentine tree, has many traditional uses in terms of food and medicine in the Mediterranean basin and its distribution areas in Asia (Baytop, 1999; Topcu *et al.*, 2007). In archaeological studies, it has been determined that the fruits of the species have been used as food by humans since 7000 BC (Ozcan, 2004). Today, in rural areas, the fruits of the species are traditionally consumed as a snack or mixed with products such as walnuts and figs as food. Similarly, turpentine tree fruits are consumed as an additive in some special village breads and as a spice material called 'zahter' (Aydin and Ozcan, 2002). The oils obtained from the fruits and nuts of this species, are widely used in the production of soaps called 'Bittim' as well as for edible use in Turkey (Baytop, 1999; Kafkas *et al.*, 2002). This soap is an important natural product that cleanses the skin from toxins (Tastekin *et al.*, 2014).

Because *P. terebinthus* is rich in tannin and resin contents, it has an aromatic plant quality. In terms of traditional medicine, the fruits of this species have been used directly or externally in the treatment of diseases such as infections, rheumatism, eczema, ulcer, stomach-ache, cough, asthma and sunstroke (Baytop, 1999; Yesilada *et al.*, 1995; Tuzlaci and Eryasar Aymaz, 2001; Flamini *et al.*, 2004; Ozcan, 2004; Ertas *et al.*, 2013). In Turkey, the leaves of the turpentine tree are used in the treatment of burns, and the resin is used in the treatment of antiseptic bronchitis (Topcu *et al.*, 2007). Ripe fruits, crushed by pounding, are used as a mouthwash to prevent toothache after boiling in sugar water or molasses (Senkardes and Tuzlaci, 2014).

Turpentine fruits are rich in protein, fat, fibre, unsaturated fatty acids and minerals, which increases their nutritional value commercially (Ozcan, 2004; Matthäus and Ozcan, 2006; Yilmaz *et al.*, 2010; Senyay-Oncel *et al.*, 2011). Turpentine coffee made of fruits, which had limited use in rural areas in the past, has now become a product of commercial importance. It has been determined that Turpentine coffee, which is completely natural and caffeine-free, is a good source of trace elements and fatty acids (Ciftci *et al.*, 2009; Kizil and Turk, 2010; Ozel *et al.*, 2014). Turpentine coffee has been found to have higher antioxidant

activity compared to direct fruit consumption (Orhan *et al.*, 2012). The resin of this species is valuable as a supporting additive to improve the viability of probiotic bacteria in yogurt, cheese and similar dairy products (Schoina *et al.*, 2015; Schoina *et al.*, 2018). Apart from the specified features, it has been stated that Turpentine oils have good performance as biodiesel fuel and can be an environmentally friendly natural fuel mixture (Ozcanlı *et al.*, 2011; Kar *et al.*, 2012). The films in which the leaf, fruit and shoot extracts of the turpentine tree were used in the mixture offered an alternative food packaging material (Kaya *et al.*, 2018).

Apart from its traditional uses, the turpentine species is also important in terms of modern medicine and pharmacy (Bozorgi *et al.*, 2013). This plant species has various bioactivities, such as antioxidant, antimicrobial and cytotoxic, with its flavonoid, phenolic and alkaloid contents in parts such as flowers, leaves, fruits and shoots (Topcu *et al.*, 2007; Durak and Ucak, 2015; Hacibekiroglu *et al.*, 2015; Çoban *et al.*, 2017). It has been determined that the high antioxidant capacity of the extracts obtained from the leaves of the species has a possible protective role in cancer risks (Lampronti *et al.*, 2005; Kavak *et al.*, 2010). The antioxidant activities of the flowers collected in spring have been found to have preventive or retarding effects against the oxidative stress that causes aging and some chronic diseases (heart, diabetes, cancer, etc.) (Turkoglu *et al.*, 2017). It has been stated that the fruit essential oils of the turpentine tree are promising for the treatment of colorectal cancer cells (Awwad *et al.*, 2020). Turpentine tree oils have phytochemical properties as they are rich in unsaturated fatty acids, tocopherols, polyphenols and carotenoids (Durmaz and Gokmen, 2011). It has been determined that extracts obtained from dried fruits of the turpentine tree have a hypolipidemic effect on rabbits (Bakirel *et al.*, 2003), and their antioxidant properties have a cholesterol reduction (about 1%) effect (Riemersma *et al.*, 1991). This species has shown potential effects for obesity and diabetes prevention and phytotherapy (Foddai *et al.*, 2015).

In all the beneficial forms mentioned above, the essential oil characteristics of this aromatic plant are particularly important. Especially, the young shoots, flowers, raw and ripe fruits of the turpentine tree contain essential oil (Couladis *et al.*, 2003). Recently, the *P. terebinthus* species has started to be the focus of attention in sectors such as pharmacology, cosmetics, agriculture and food because of its essential oil properties (Ulukanli *et al.*, 2014). In the literature, it is seen that different results have been obtained in many studies

on the amount, quality and composition of essential oil in certain organs of the turpentine tree (Couladis *et al.*, 2003; Flamini *et al.*, 2004; Ozcan *et al.*, 2009; Pulaj *et al.*, 2016; Piras *et al.*, 2017). In these studies, it has been mentioned that these differences, which cause changes in the aromatic and medicinal properties of the plant, are caused by variables such as sample collection time and method, essential oil analysis method and climatic characteristics of the locations where the samples are harvested. Our hypothesis is that apart from the variables mentioned above and the genetic characteristics of the plant, various environmental factors can cause significant changes in the essential oil characteristics of the Turpentine species. In this study, the effects of environmental factors (elevation, slope, aspect, climate, etc.) on the essential oil yield and component ratios of *P. terebinthus* fruits were investigated.

MATERIAL AND METHODS

Study area

The study area, the Lake District (38°25'–36°06' N, 29°30'–32°34' E), is located in the Mediterranean phytogeographic region of Turkey. The Lake District covering the provinces of Antalya, Isparta and Burdur has coastlands, rugged terrains, plains and mountainous parts up to 3,000 m. Although typical Mediterranean climate prevails near the coastal parts of Antalya in the district, there is a transition to a continental climate in mountainous areas and parts extending towards central Anatolia in Isparta and Burdur. Mostly carbonated rocks, red Mediterranean soils and brown forest soils are dominant in the study area (Sandler *et al.*, 2015). In this area, *P. terebinthus* is located in red pine forests at different heights, in maquis or pseudomaquis vegetation types where the forest structure is degraded and on arid, rocky and stony slopes up to the altitude of 1500–1600 m in the Taurus Mountains (Davis, 1967). Fruit samples of *P. terebinthus* were collected from 34 wild populations in different locations of the district (Figure 1). At each location, elevation (m), slope (%), surface stoniness (%), mean annual temperature (°C) and total annual precipitation (mm) have been recorded as environmental variables. Location aspects were converted into radiation index values (McCune and Keon, 2002). In addition, sand (%), silt (%), clay (%), pH, total lime (%), organic matter (%) and soil skeleton content (%) were determined in soil samples taken from 0–10 cm, 10–30 cm and 30–50 cm depth levels at each location.

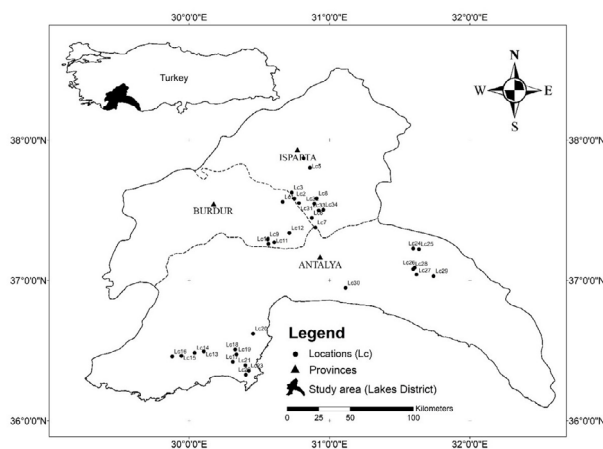


Figure 1. Map of the study area showing the sampling locations in the Lake District, Turkey.

Plant material

The *P. terebinthus* is a deciduous tree that can grow up to 6 metres or a shrub that can grow to 2–3 metres in height. The globose and broadly obovate fruits of the species are drupe. Although the fruits are reddish in raw form, they gradually darken in shades of green during the ripening phase. Ripe fruits are 4–6 mm wide and 5–6 mm long (Davis, 1967). The ripe fruit samples were collected from 34 wild populations from September to the end of October in 2010. The fruit samples taken from the field were brought to the laboratory in polyethylene packages on the same day. The fruits, which were dried until they became air-dried, were first ground in an electronic mixer. Essential oils were obtained by the hydro distillation method for 3 h in a Clevenger-type apparatus by adding 4 litres of water to 350 grams of samples. As a result of this process with 2 repetitions, the average of essential oil yields (EOYs; %, v/w) was recorded for each of the 34 different locations. The samples, which were transferred to Eppendorf by means of micropipettes without water, were stored at –20 °C for use in essential oil component analyses after being covered with parafilm.

Gas chromatography–mass spectrometry (GC/MS) analyses

Analyses were made using a Shimadzu QP 5050 GC/MS spectrometer system in the EI mode at 70 eV ionization energy. The separation was conducted on a CP-Wax 52CB column (50 m x 0.32 mm, 1.2 μm film thickness). For the Carrier gas Helium was used with an inlet pressure of 10 psi and the injection volume of 10

μL . The oven temperature increased from 60 °C to 220 °C at a rate of 2 °C/min, then held isothermal at 220 °C for 20 min with an injector temperature of 250 °C and a detector temperature of 240 °C. Essential oil components were identified by comparing the retention indices with Wiley, NIST, Tutor and private libraries online as well as published articles (Adams, 2007).

Statistical analysis

Principal component analysis (PCA) was applied to determine the relationships between major essential oil components (higher than 5%) and environmental variables in different locations. 'FactoMineR' and 'ggplot2' packages were used in R Studio for PCA analysis (Le et al., 2008; Wickham, 2009). Pearson correlation coefficients (using a 5% significance level) between EOYs and environmental variables was also calculated using 'corrplot' package in RStudio (Wei et al., 2017).

RESULTS AND DISCUSSION

Just like other secondary metabolites (phenolics, flavonoids etc.), essential oils are important indicators in terms of medicinal and aromatic qualities of plants. EOYs from ripe fruits of the turpentine tree and the chemical compositions of these essential oils obtained from 34 different locations (Lc) in the Lake District of Turkey are given in Table 1. EOYs in ripe fruits of the turpentine tree ranged from 0.05% (v/w) to 0.19% (v/w) in the populations of the different locations. The highest EOYs (0.19%) was obtained from the fruits collected from Lc19 at an elevation of 551 m in Antalya province. In total, 21 volatile components, 94.5% and 99.7% of the total essential oils, were identified. The number of components and their ratios vary in different populations. The major components of the turpentine tree essential oil were α -pinene, limonene, Ocimene, *p*-Cymene, myrcene and sabinene. The most abundant component was α -pinene (an average rate of 41.01%), which ranged from 20.87% in Lc4 (Isparta) to 60.98 % in Lc14 (Antalya). Limonene is the second major component, with an average rate of 14.28% in 34 populations. Ocimene, *p*-Cymene, myrcene and sabinene component ratios were determined as 10.68, 7.73, 5.97 and 5.17%, respectively.

In a similar study previously conducted in Turkey, EOYs were found between 0.06% and 0.16% in fruit samples of the turpentine tree collected from different locations, such as Antalya, Izmir and Hatay. α -pinene (9.5-51.3%), limonene (<0.1-39.0%) and caryophyllene oxide (<0.1-51.5) were identified as the most abundant volatile components in essential oils (Ozcan et al., 2009). It has also been stated that EOYs and volatile component ratios vary in fruit samples collected from different localities. EOYs were determined as 0.02% and 0.12% in fruit samples of the turpentine tree harvested from Prizren and Ura e Shejtë in Kosovo, respectively. Although α -pinene ratios were between 12.58–21.43%, limonene ratios were 15.65-23.87% in fruits taken from different locations. The ratios of (Z)- β -Ocimene (2.23-44.85%) and (E)- β -Ocimene (9.59-40.49 %) differed significantly according to the locations (Pulaj et al., 2016). Apart from these studies in the literature, EOYs were found as 0.54% and 0.73% in the raw and ripe fruits, respectively, of the Turpentine tree origins from İçel province in Turkey. In the same study, the major volatile components in the essential oils of ripe fruits were limonene (32.8%), β -pinene (22.5%), α -pinene (5.3%) and germacrene D (4.6%), respectively (Couladis et al., 2003). In the ripe fruits of turpentine trees originating in Jordan, (E) -ocimene (33.8%), sabinene (24.1%), (Z) -ocimene (13.0%) and α pinene (6.5%) were determined as the major components, respectively (Flamini et al., 2004). In another recently completed study in Turkey, EOYs was determined as 0.082% in the ripe fruits of the species, and the major components were α -pinene (37.23%) and limonene (19.89%) (Inan, 2021).

When the results obtained from our research and other studies completed in the literature are interpreted together, although there are some partial differences in EOYs and components, the results generally overlap. It was stated that occurring partial differences may depend on variables such as the time and shape of the samples that were collected and the analysis methods (Gogus et al., 2011; Ozel et al., 2014; Inan, 2021). On the other hand, in some studies, it was determined that although standard methods were applied to the samples collected at the same ripening time from different localities, there were variations in EOYs and component ratios (Table 1)(Ozcan et al., 2009; Pulaj et al., 2016).

Table 1. Essential oil yields and the chemical composition of the essential oils obtained from ripe fruits of the turpentine tree in different locations.

	Lc1	Lc2	Lc3	Lc4	Lc5	Lc6	Lc7	Lc8	Lc9	Lc10	Lc11	Lc12	Lc13	Lc14	Lc15	Lc16	Lc17	ID
α -Pinene	54.99	44.16	48.11	20.87	36.09	46.16	59.69	39.26	46.10	41.38	52.99	33.42	51.02	60.98	27.01	52.51	44.02	a, b
Camphene	1.64	2.47	1.03	1.26	0.99	1.65	1.50	1.86	1.63	0.94	1.35	1.07	1.76	2.80	1.21	2.05	1.75	a, b
β -Pinene	1.16	5.42	6.42	6.00	2.42	7.71	2.59	9.73	9.32	2.11	5.75	5.97	3.59	6.69	2.00	3.09	2.87	a, b
Sabinene	3.21	0.74	1.27	7.78	1.40	6.46	1.25	11.05	4.84	9.63	4.95	4.75	6.12	5.03	9.13	2.99	3.03	a
Myrcene	1.94	26.48	2.29	13.46	4.90	10.82	2.80	2.19	3.39	1.37	7.69	2.25	1.78	1.69	12.32	2.92	6.36	a, b
Phellandrene	n	n	n	n	n	n	2.93	1.27	2.31	0.94	2.44	1.53	0.44	n	0.46	n	n	a, b
Limonene	26.95	15.99	12.22	3.23	22.68	8.28	9.43	7.49	16.90	8.07	9.23	8.37	28.45	6.03	14.02	4.08	18.23	a, b
Ocimene	0.46	0.42	0.82	29.39	8.13	5.73	5.35	12.26	3.76	1.01	3.33	23.18	0.63	8.68	9.18	2.16	17.14	a
p-Cymene	0.92	0.47	0.66	11.41	6.49	2.19	1.02	4.27	2.35	26.16	1.42	10.71	0.69	2.60	17.50	0.81	4.19	a, b
β -Caryophyllene	4.48	1.34	11.90	1.32	4.47	1.81	3.38	3.13	3.14	1.47	1.87	2.71	1.46	1.25	1.14	n	0.89	a, b
Terpinene	n	n	n	1.42	4.23	n	3.36	3.53	3.73	n	n	1.09	n	n	0.19	1.11	0.21	a, b
Terpinolene	n	n	n	n	1.86	6.14	n	n	n	3.34	5.77	2.38	0.39	0.62	n	23.95	0.24	a, b
D-carene	n	n	n	n	4.66	n	2.52	n	n	n	n	n	n	n	3.39	2.98	0.16	a
α -Copaene	0.58	0.15	0.16	n	n	n	0.14	n	n	n	n	n	n	n	n	n	n	a
Terpineol-4	n	n	n	n	n	n	n	n	n	n	n	n	0.82	n	n	n	n	a, b
α -Humulene	n	n	1.58	n	n	n	0.41	n	n	n	n	n	n	n	n	n	n	a
Germacrene D	n	n	1.40	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
5-Ethenyl undecane	n	n	2.12	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Ylangene	n	n	1.10	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Naphthalene	1.98	1.75	0.38	n	0.35	n	n	n	n	n	n	n	n	0.36	n	n	n	a
Codinene	n	n	1.77	n	n	n	0.27	n	n	n	n	n	n	n	n	n	n	a
Muurolol	n	n	1.29	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Total (%)	98.29	99.38	94.50	96.12	98.64	96.92	96.62	96.02	97.47	96.41	96.76	97.40	97.14	96.71	97.53	98.63	99.06	
EOYs	0.07	0.09	0.08	0.09	0.05	0.08	0.14	0.08	0.14	0.09	0.13	0.14	0.14	0.09	0.10	0.09	0.12	

EOYs: Yields of Essential Oil (% v/w), ID: Identification, Lc: Locations, a: Identification by retention time and comparison with GC-MS spectra. b: Identification by retention time and comparison with GC-MS spectra and comparison with authentic standards prepared by co – chromatography, n: Not detected

Table 1.(continue) Essential oil yields and the chemical composition of the essential oils obtained from ripe fruits of the turpentine tree in different locations.

	Lc18	Lc19	Lc20	Lc21	Lc22	Lc23	Lc24	Lc25	Lc26	Lc27	Lc28	Lc29	Lc30	Lc31	Lc32	Lc33	Lc34	ID
α -Pinene	30.82	47.61	28.90	27.82	25.90	24.45	43.26	44.10	40.49	40.73	24.43	44.14	29.81	28.42	60.21	49.30	45.38	a, b
Camphene	0.92	1.54	0.94	1.03	0.41	n	1.34	1.38	1.05	1.14	n	0.52	1.79	1.48	2.60	1.50	1.46	a, b
β -Pinene	4.84	3.44	3.46	3.85	4.01	3.50	4.26	3.23	4.37	4.69	1.80	2.98	14.84	2.92	4.42	2.65	5.10	a, b
Sabinene	2.35	0.57	4.84	2.44	2.35	0.88	6.25	3.32	9.40	6.44	3.14	4.31	16.13	12.32	3.49	8.02	6.09	a
Myrcene	3.37	1.06	3.93	4.93	3.37	3.69	6.05	3.27	3.03	6.81	2.29	4.53	1.45	31.80	2.05	11.53	5.10	a, b
Phellandrene	n	n	n	n	n	n	2.18	2.09	3.11	2.51	0.41	4.42	n	2.08	0.76	0.25	1.33	a, b
Limonene	12.50	29.35	10.67	20.10	13.93	18.61	13.50	17.62	12.28	16.38	51.28	13.20	2.51	9.40	14.60	2.91	6.94	a, b
Ocimene	22.27	0.90	27.34	24.42	29.34	29.50	7.91	11.23	7.43	7.04	9.94	13.99	24.51	5.26	4.90	3.50	2.07	a
p-Cymene	15.39	13.85	8.00	7.10	9.86	9.71	10.08	8.78	8.42	7.93	3.61	9.86	6.03	1.65	4.70	19.28	24.70	a, b
β -Caryophyllene	1.12	0.40	3.50	0.46	n	n	2.64	2.98	2.65	2.74	2.64	0.82	0.76	3.64	1.47	0.42	1.17	a, b
Terpinene	0.89	n	1.21	0.80	1.27	0.60	n	n	n	n	n	n	0.40	n	n	n	n	a, b
Terpinolene	4.02	0.55	4.77	3.26	5.59	7.57	2.07	1.73	6.61	0.76	n	0.50	n	n	n	n	n	a, b
D-carene	n	n	n	2.83	3.38	0.99	n	n	n	n	n	n	n	n	n	n	0.33	a
α -Copaene	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Terpineol-4	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a, b
α -Humulene	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Germacrene D	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
5-Ethenyl undecane	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Ylangene	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Naphthalene	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Codinene	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Murolol	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	n	a
Total (%)	98.45	99.25	97.55	99.00	99.40	99.47	99.51	99.70	98.82	97.14	99.52	99.24	98.20	98.95	99.19	99.33	99.65	
EOYS	0.11	0.19	0.09	0.12	0.10	0.12	0.07	0.08	0.09	0.09	0.11	0.13	0.08	0.10	0.07	0.14	0.12	

EOYS: Yields of Essential Oil (% v/w), ID: Identification, Lc: Locations, a: Identification by retention time and comparison with GC-MS spectra. b: Identification by retention time and comparison with GC-MS spectra and comparison with authentic standards prepared by co – chromatography, n: Not detected

Only longitude ($r:-0.344$) and sand % in 10-30 cm ($r:0.346$) from the environmental variables indicated statistically significant correlations with EOYs at the level $p < 0.05$ (Figure 2). Similar to the results of this study, the increase in the amount of sand (%) in the soil was also positively correlated with the EOYs of *Juniper* cones collected from the Lake District (Gulsoy *et al.*, 2019). In Mediterranean-type arid or semi-arid ecosystems, the increase in the amount of sand in the soil is one of the most important factors that cause a decrease in plant nutrients and an imbalance in the water economy of the soil. It has been determined that the turpentine tree, which spreads by closing its stomata in arid environments, is a water-spender plant, and accordingly, the relative humidity content of the plant changes in different environments (Sakcali and Ozturk, 2004). It is possible that the changes in the nutrient and water economy in the soil depending on the sand content increased the drought stress in the plant and changed the amounts of the secondary metabolites, such as essential oil and phenolics.

PCA is a suitable technique for associating the major essential oil components from fruits in different geographical origins with numerous environmental variables. The eigenvalues and percentages of variance explanation for the first 3 PCA axes were 1.98 and 32.9 for PC1, 1.48 and 24.7 for PC2 and 1.09 and 18.3 for PC3, respectively. The first 3 PCA axes explained 75.9% of the total variance in the essential oil components data set (Table 2).

Table 2. Eigenvalues and variances (%) of the first 3 PCA axes according to the Essential Oil Components dataset.

PCA Axes	Eigenvalue	Variance (%)	Cumulative Variance (%)
PC1	1.98	32.9	32.9
PC2	1.48	24.7	57.6
PC3	1.10	18.3	75.9

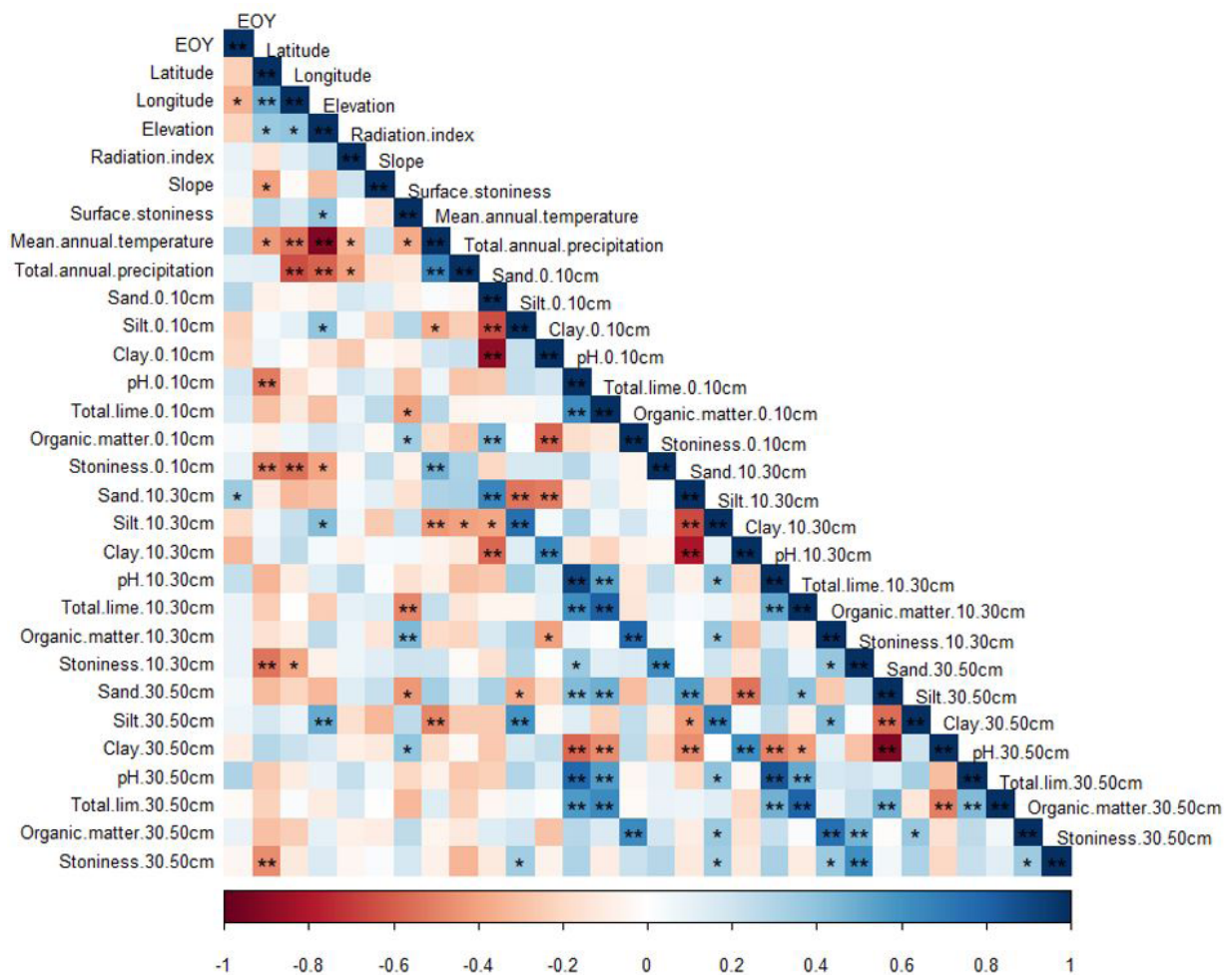


Figure 2. Correlation coefficients (* $p < 0.05$, ** $p < 0.01$) between essential oil yield and environmental variables.

The correlation coefficients of the first 3 PCA axes with essential oil components and environmental variables are listed in Table 3. Although PC1 axis (with a 32.9% variance) had the highest negative correlation with α -Pinene (r : -0.816) among 3 PCA axes, this axis provided the highest positive contribution to Ocimene (r : 0.719). PC2 (with a 24.7% variance) was the axis with the highest positive contribution to Sabinene (r : 0.593) and the highest negative contribution to Limonene (r : -0.685). PC3 (with

a 18.3% variance), the 3rd axis, had the highest negative correlation with p-Cymene (r : -0.591) and the highest positive correlation with Myrcene (r : 0.776).

When environmental variables and essential oil components are linearly correlated according to the highest correlation rankings on the same PCA axis, silt (%; 10-30 cm) and pH (30-50 cm) were the variables with the highest positive contribution to α -Pinene and the highest negative relation to Ocimene according to PC1. In

Table 3. The correlation coefficients (r) of the first 3 PCA axes with essential oil components and environmental variables.

Major Essential Oil Components				
PCA Axes	PC1	PC2	PC3	
Alfa Pinene	-0.816	0.440	-0.288	
Sabinene	0.553	0.593	-0.024	
Myrcene	0.238	0.439	0.776	
Limonene	-0.407	-0.685	0.251	
Ocimene	0.719	-0.523	0.000	
pCymene	0.514	0.047	-0.591	
Environmental Factors				
PCA Axes	PC1	PC2	PC3	
Latitude	0.040	0.029	0.023	
Longitude	-0.098	0.432	0.161	
Elevation (m)	-0.061	0.018	-0.289	
Radiation index	-0.235	0.051	-0.045	
Slope (%)	-0.337	-0.142	0.148	
Surface stoniness (%)	-0.112	-0.162	-0.348	
Mean annual temperature (°C)	0.130	-0.132	0.198	
Total annual precipitation (mm)	0.243	-0.388	0.034	
Sand (%; 0-10cm)	0.193	-0.127	-0.073	
Silt (%; 0-10cm)	-0.252	0.175	-0.153	
Clay (%; 0-10cm)	-0.094	0.056	0.186	
pH (0-10cm)	-0.147	0.332	0.011	
Total lime (%; 0-10cm)	-0.018	0.276	0.413	
Organic matter (%; 0-10cm)	0.069	-0.152	-0.263	
Stoniness (%; 0-10cm)	-0.115	-0.052	-0.162	
Sand (%; 10-30cm)	0.239	-0.176	0.012	
Silt (%; 10-30cm)	-0.341	0.218	-0.122	
Clay (%; 10-30cm)	-0.054	0.066	0.078	
pH (10-30cm)	-0.287	0.191	-0.009	
Total lime (%; 10-30cm)	-0.067	0.315	0.590	
Organic matter (%; 10-30cm)	-0.024	-0.210	-0.291	
Stoniness (%; 10-30cm)	-0.121	0.148	-0.358	
Sand (%; 30-50cm)	0.116	0.175	0.007	
Silt (%; 30-50cm)	-0.153	0.040	-0.137	
Clay (%; 30-50cm)	-0.068	-0.224	0.054	
pH (30-50cm)	-0.388	0.210	0.046	
Total lime (%; 30-50cm)	0.040	0.311	0.257	
Organic matter (%; 30-50cm)	0.047	0.012	-0.361	
Stoniness (%; 30-50cm)	-0.206	0.138	-0.232	

relation to PC2, longitude is the most important variable that contributes positively to the Sabinene component, whereas the Limonene component appears to be positively associated with locations where total annual precipitation (mm) increases. When the relationships were interpreted according to PC3, the increases in total lime

(%, 0-10 cm) and the total lime amounts of soils taken from different localities (% , 10-30 cm) had a positive effect on the myrcene component, whereas it had a negative effect on p-Cymene. In addition, the positions of all these variables according to their correlations with the PCA axes are showed in Figure 3 and Figure 4.

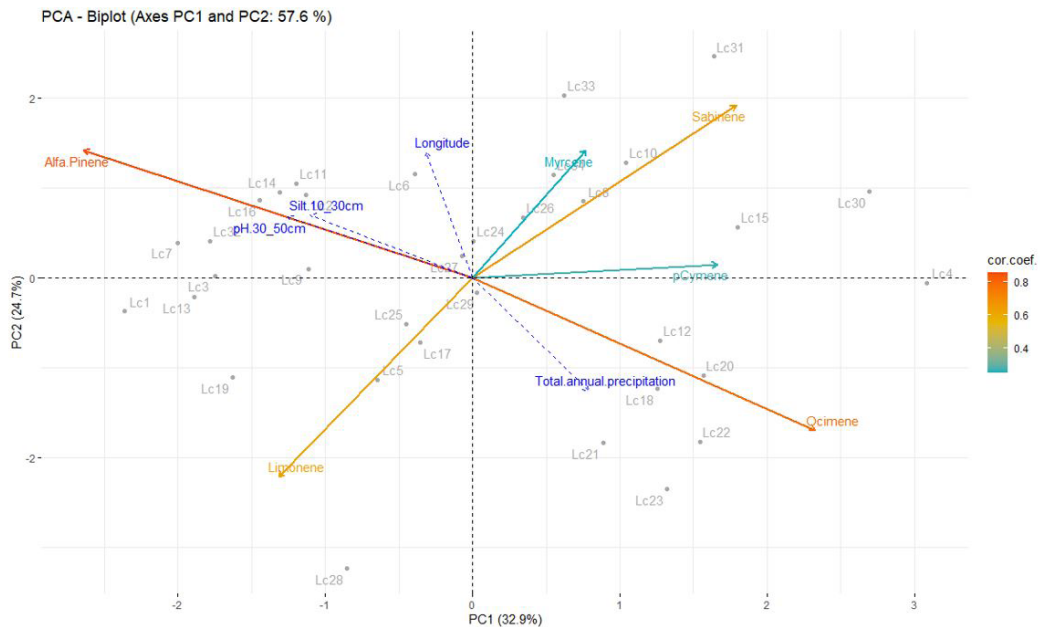


Figure 3. PCA graph of major components (higher than 5%) of *P. terbinthus* L. fruits from 34 different locations and relations with environmental variables in PC1 and PC2.

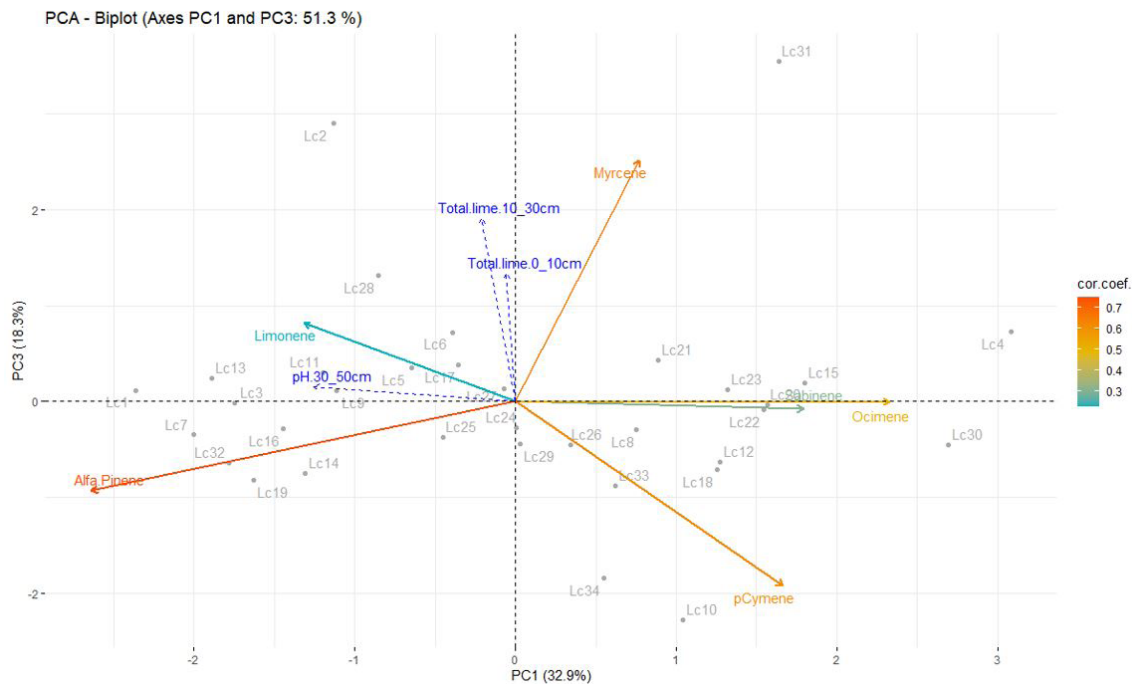


Figure 4. PCA graph of major components (higher than 5%) of *P. terbinthus* L. fruits from 34 different locations and relations with environmental variables in PC1 and PC3.

It is an important process to obtain high EOYs and desired essential oil component ratios from plants with medicinal and aromatic properties to meet the requirements of the sector. For this, it is necessary to determine the effect factors that cause changes in EOYs and essential oil component ratios. In general, besides genetic, ontogenic and morphogenetic factors, it is stated that another important group of factors that cause changes on various secondary metabolites such as essential oil in plants are environmental variables (Verma and Shukla, 2015). During their growing periods, plants are constantly interacting with biotic and abiotic environmental factors. Especially changing abiotic environmental factors (climatic, edaphic, and physiographic) can be responsible for the variations in secondary metabolites (phenolics, flavonoids, essential oils etc.,) as an important stress factor in the medicinal plants (Li *et al.*, 2020). As a result of the PCA analysis in this study, it was observed that various abiotic environmental variables caused changes in the essential oil components of the turpentine fruits. In the study, the intensity of annual precipitation, which is one of the climate parameters that vary depending on the location of the sample areas, caused an increase in the Limonene component and a decrease in the Sabinene component in the essential oils of the turpentine fruits. Figueiredo *et al.*, (2008) mentioned that the weather conditions can be crucial determinants for the essential oil properties of various medicinal plant species. In a study, the amount of some essential oil components of the *Flourensia cernua* plant have been detected to increase in shaded environments as a locally climatic factor (Estell *et al.*, 2016). Turtola *et al.*, (2003) found that various monoterpene concentrations in the stem wood of two different conifer species (*Pinus sylvestris* L. and *Picea abies* (L.) Karst.) were significantly affected by drought stress. Therefore, it would be correct to interpret the changes in essential oil components due to annual precipitation as drought stress.

On the other hand, in this study, it was observed that some soil properties such as total lime (%), sand (%), pH caused changes in the concentration of essential oil components such as α -pinene, ocimene, myrcene and p-Cymene. In literature, some researchers considered that soil variables may be determinants in the essential oil properties of plants, too. For instance, Hermino and Antonio (1998) reported that p-cymene, which is the major essential oil component of the *Thymus piperella* L. plant, is positively affected by the increasing organic matter in the soil. Also, Pereira *et al.* (2000) stated that chemical variation in essential oils of *Thymus caespitius* plant may be affected by genetics and edaphic factors in an environment where many other environmental conditions (slope, altitude etc.) are constant. As a result, although the relationships between essential oil properties and all the factors are very complex, interactions with environmental variables should definitely be considered.

CONCLUSION

Medicinal and aromatic plants containing essential oils provide important ecosystem services to industries such as pharmacology, cosmetics, agriculture and food. It has been stated that the trace element levels of the turpentine tree, which has essential oil properties in different organs, are within the tolerance range prescribed by the World Health Organization and can be evaluated in the category of medicinal and aromatic plants (Hamlat *et al.*, 2019). It is important to determine the factors affecting the EOYs and component ratios, which are decisive for the phytochemical and pharmacological use of the turpentine tree. Many studies mentioned that the EOYs and volatile components of turpentine trees vary depending on the different plant organs, ripening period of plant material, harvest time, locations, climatic conditions and analysis techniques in the laboratory. In this study, it was determined that the EOYs and components extracted from the fruits of the turpentine tree grown in different regions differ according to environmental variables. Soil properties such as sand (%), total lime (%) and pH, annual precipitation from climatic properties and longitude as a location parameter caused changes in the EOYs and component ratios of the turpentine tree. Our findings supported the idea that environmental factors could be a limiting factor for the demands of the essential oil industry. On the other hand, plant essential oil and environmental factor relations are still very complex. Consequently, more research on the bioactivity of the turpentine tree and other similar species is needed to clarify the findings of this study. It is important to strengthen the findings obtained by comparing this study with future research.

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 Review: SG, KÖ, GÖ

REFERENCES

- ADAMS, R.P. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry. (4th ed.). Allured Publishing Corporation, Carol Stream, IL, US. 2007. 804 p.
- AMIRI, S.; MOHAMMADI, R. Establishment of an efficient in vitro propagation protocol for Sumac (*Rhus coriaria* L.) and confirmation of the genetic homogeneity. Scientific Reports, v.11, n.1, p. 1-9, 2021.

- AWWAD, O.; ALABBASSI, R.; ABAZA, I.F.; COPERCHINI, F.; ROTONDI, M.; CHIOVATO, L.; AFIFI, F.U. Effect of *Pistacia palaestina* Boiss. essential oil on colorectal cancer cells: inhibition of proliferation and migration. *Journal of Essential Oil Bearing Plants*, v. 23, n.1, p. 26-37, 2020.
- AYDIN, C.; OZCAN, M. Some physico-mechanic properties of terebinth (*Pistacia terebinthus* L.) fruits. *Journal of Food Engineering*, v.53, n.1, p. 97-101, 2002.
- BAKIREL, T.; SENER, S.; BAKIREL, U.; KELES, O.Y.A.; SENNAZLI, G.; GUREL, A. The investigation of the effects of *Pistacia terebinthus* L. upon experimentally induced hypercholesterolemia and atherosclerosis in rabbits. *Turkish Journal of Veterinary and Animal Sciences*, v.27, n.6, p. 1283-1292, 2003.
- BAYTOP, T. Therapy with medicinal plants in Turkey (past and present) (2nd ed.). Nobel Medicine Publication, 1999.
- BOZORGI, M.; MEMARIANI, Z.; MOBLI, M.; SALEHI SURMAGHI, M.H.; SHAMS-ARDEKANI, M.R.; RAHIMI, R. Five *Pistacia* species (*P. vera*, *P. atlantica*, *P. terebinthus*, *P. khinjuk*, and *P. lentiscus*): A review of their traditional uses, phytochemistry, and pharmacology. *The Scientific World Journal*, p. 1-33, 2013.
- CIFTCI, H.; OZKAYA, A.; KARIPTAS, E. Determination of fatty acids, vitamins and trace elements in *Pistacia terebinthus* coffee. *Journal of Food, Agriculture and Environment*, v.7, n.3-4, p. 72-74, 2009.
- COBAN, E.P.; BIYIK, H.; TORUN, B.; YAMAN, F. Evaluation the antimicrobial effects of *Pistacia terebinthus* L. and *Papaver rhoeas* L. extracts against some pathogen microorganisms. *Indian J Pharm Education Research*, v.51, n.3, p. 377-80, 2017.
- COULADIS, M.; OZCAN, M.; TZAKOU, O.; AKGUL, A. Comparative essential oil composition of various parts of the turpentine tree (*Pistacia terebinthus* L.) growing wild in Turkey. *Journal of the Science of Food and Agriculture*, v.83, n.2, p. 136-138, 2003.
- DAVIS, P.H. Flora of Turkey and the east Aegean Islands, Vol. 2. Edinburgh University Press, 1967. p. 546-547.
- DURAK, M.Z.; UCAK, G. Solvent optimization and characterization of fatty acid profile and antimicrobial and antioxidant activities of Turkish *Pistacia terebinthus* L. extracts. *Turkish Journal of Agriculture and Forestry*, v.39, n.1, p. 10-19, 2015.
- DURMAZ, G.; GOKMEN, V. Changes in oxidative stability, antioxidant capacity and phytochemical composition of *Pistacia terebinthus* oil with roasting. *Food Chemistry*, v.128, n.2, p. 410-414, 2011.
- ERTAS, E.; BEKIROGLU, S.; OZDEMIR, I.; DEMIRTAS, I. Comparison of fatty acid, sterol, and tocol compositions in skin and kernel of turpentine (*Pistacia terebinthus* L.) fruits. *Journal of the American Oil Chemists' Society*, v.90, n.2, p. 253-258, 2013.
- ESTELL, R.E.; FREDRICKSON, E.L.; JAMES, D.K. Effect of light intensity and wavelength on concentration of plant secondary metabolites in the leaves of *Flourensia cernua*. *Biochemical Systematics and Ecology*, v.65, p. 108-114, 2016.
- FLAMINI, G.; BADER, A.; CIONI, P.L.; KATBEH-BADER, A.; MORELLI, I. Composition of the essential oil of leaves, galls, and ripe and unripe fruits of Jordanian *Pistacia palaestina* Boiss. *Journal of Agricultural and Food Chemistry*, v.52, n.3, p. 572-576, 2004.
- FODDAI, M.; KASABRI, V.; AFIFI, F.U.; AZARA, E.; PETRETTO, G.L.; PINTORE, G. In vitro inhibitory effects of Sardinian *Pistacia lentiscus* L. and *Pistacia terebinthus* L. on metabolic enzymes: Pancreatic lipase, α -amylase, and α -glucosidase. *Starch-Stärke*, v.67, n.1-2, p. 204-212, 2015.
- FIGUEIREDO, A.C.; BARROSO, J.G.; PEDRO, L.G.; SCHEFFER, J.J. Factors affecting secondary metabolite production in plants: volatile components and essential oils. *Flavour and Fragrance journal*, v.23, n.4, p. 213-226, 2008.
- GOGUS, F.; OZEL, M. Z.; KOCAK, D.; HAMILTON, J. F.; LEWIS, A. C. Analysis of roasted and unroasted *Pistacia terebinthus* volatiles using direct thermal desorption-GC/GC-TOF/MS. *Food Chemistry*, v.129, n.3, p. 1258-1264, 2011.
- GULSOY, S.; OZKAN, G.; SENOL, H.; MERT, A. Assessment of essential oil properties in *Juniperus excelsa* subsp. *excelsa* cones depending on site factors. *Fresenius Environmental Bulletin*, v.28, n.4, p. 2380-2389, 2019.
- HACIBEKIROGLU, I.; YILMAZ, P.K.; HASIMI, N.; KILINC, E.; TOLAN, V.; KOLAK, U. In vitro biological activities and fatty acid profiles of *Pistacia terebinthus* fruits and *Pistacia khinjuk* seeds. *Natural Product Research*, v.29, n.5, p. 444-446, 2015.
- HAMLAT, N.; BENARFA, A.; BELADEL, B.; BEGAA, S.; MESSAOUDI, M.; HASSANI, A. Assessment of the contents of essential and potentially toxic elements in *Pistacia terebinthus* L. and *Pistacia lentiscus* L. by INAA technique. *Journal of Radioanalytical and Nuclear Chemistry*, v.322, n.2, p. 1127-1131, 2019.
- HERMINO, B.; ANTONIO, A. Environmental factors affecting chemical variability of essential oils in *Thymus piperella* L. *Biochemistry Systematic and Ecology*, v.26, p. 811-822, 1998.
- INAN, M. Seasonal variation of fatty and essential oil in terebinth (*Pistacia terebinthus* L.) fruit. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, v.49, n.1, p. 12171-12171, 2021.
- KAFKAS, S.; KAFKAS, E.; PERL-TREVES, R. Morphological diversity and a germplasm survey of three wild *Pistacia* species in Turkey. *Genetic Resources and Crop Evolution*, v.49, n.3, p. 261-270, 2002.
- KAR, Y.; SEN, N.; DEVECI, H. Usability of terebinth (*Pistacia terebinthus* L.) fruits as an energy source for diesel-like fuels production. *Energy Conversion and Management*, v.64, p. 433-440, 2012.
- KAVAK, D.D.; ALTIOK, E.; BAYRAKTAR, O.; ULKU, S. *Pistacia terebinthus* extract: As a potential antioxidant, antimicrobial and possible β -glucuronidase inhibitor. *Journal of Molecular Catalysis B: Enzymatic*, v.64, n.3-4, p. 167-171, 2010.
- KAYA, M.; KHADEM, S.; CAKMAK, Y.S.; MUJTABA, M.; ILK, S.; AKYUZ, L.; SALABERRIA, A.M.; LABIDI, J.; ABDULQADIRA, A.H.; DELIGOZ, E. Antioxidative and antimicrobial edible chitosan films blended with stem, leaf and seed extracts of *Pistacia terebinthus* for active food packaging. *RSC Advances*, v.8, n.8, 3941-3950, 2018.
- KIZIL, S.; TURK, M. Microelement contents and fatty acid compositions of *Rhus coriaria* L. and *Pistacia terebinthus* L. fruits spread commonly in the south eastern Anatolia region of Turkey. *Natural Product Research*, v.24, n.1, p. 92-98, 2010.
- LAMPRONTI, I.; SAAB, A.; GAMBARI, R. Medicinal plants from Lebanon: Effects of essential oils from *Pistacia palaestina* on proliferation and erythroid differentiation of human leukemic K562 cells. *Minerva Biotechnologica*, v.17, n.3, p. 153-158, 2005.
- LE, S.; JOSSE, J.; HUSSON, F. *FactoMineR*: An R package for multivariate analysis. *Journal of Statistical Software*, v.25, n.1, p. 1-18, 2008.
- LI, Y.; KONG, D.; FU, Y.; SUSSMAN, M.R.; WU, H. The effect of developmental and environmental factors on secondary metabolites in medicinal plants. *Plant Physiology and Biochemistry*, v.148, p. 80-89, 2020.
- MATTHÄUS, B.; OZCAN, M. M. Quantitation of fatty acids, sterols, and tocopherols in turpentine (*Pistacia terebinthus* Chia) growing wild in Turkey. *Journal of Agricultural and Food Chemistry*, v.54, n.20, p. 7667-7671, 2006.
- MCCUNE, B.; KEON, D. Equations for potential annual direct incident radiation and heat load. *Journal of Vegetation Science*, v.13, p. 603-606, 2002.
- ORHAN, I.E.; SENOL, F.S.; GULPINAR, A.R.; SEKEROGLU, N.; KARTAL, M.; SENER, B. Neuroprotective potential of some terebinth coffee brands and the unprocessed fruits of *Pistacia terebinthus* L. and their fatty and essential oil analyses. *Food Chemistry*, v.130, n.4, p. 882-888, 2012.
- OZCAN, M. Characteristics of fruit and oil of terebinth (*Pistacia terebinthus* L.) growing wild in Turkey. *Journal of the Science of Food and Agriculture*, v.84, n.6, p. 517-520, 2004.
- OZCAN, M. M.; TZAKOU, O.; COULADIS, M. Essential oil composition of the turpentine tree (*Pistacia terebinthus* L.) fruits growing wild in Turkey. *Food Chemistry*, v.114, n.1, p. 282-285, 2009.
- OZCANLI, M.; KESKIN, A.; AYDIN, K. Biodiesel production from terebinth (*Pistacia terebinthus*) oil and its usage in diesel engine. *International Journal of Green Energy*, v.8, n.5, p. 518-528, 2011.
- OZEL, M.Z.; YANIK, D.K.; GOGUS, F.; HAMILTON, J.F.; LEWIS, A.C. Effect of roasting method and oil reduction on volatiles of roasted *Pistacia terebinthus* using direct thermal desorption-GC/GC-TOF/MS. *LWT-Food Science and Technology*, v.59, n.1, p. 283-288, 2014.
- PEREIRA, S. I.; SANTOS, P.A.G.; BARROSO, J.G.; FIGUEIREDO, A.C.; PEDRO, L.G.; SALGUEIRO, L.R.; DEANS, S.G.; SCHEFFER, J.J.C. Chemical polymorphism of the essential oils from populations of *Thymus caespitius* grown on the island S. Jorge (Azores). *Phytochemistry*, v.55, n.3, p. 241-246, 2000.
- PIRAS, A.; MARZOUKI, H.; MAXIA, A.; MARENCO, A.; PORCEDDA, S.; FALCONIERI, D.; GONCALVES, M.J.; CAVALEIRO, C.; SALGUEIRO, L. Chemical characterisation and biological activity of leaf essential oils obtained from *Pistacia terebinthus* growing wild in Tunisia and Sardinia Island. *Natural Product Research*, v.31, n.22, p. 2684-2689, 2017.
- PULAJ, B.; MUSTAFA, B.; NELSON, K.; QUAVE, C. L.; HAJDARI, A. Chemical composition and in vitro antibacterial activity of *Pistacia terebinthus* essential oils derived from wild populations in Kosovo. *BMC Complementary and Alternative Medicine*, v.16, n.1, p. 1-9, 2016.
- RIEMERSMA, R.A.; WOOD, D.A.; OLIVER, M.F.; ELTON, R.A.; MACINTYRE, C.C.A.; GEY, K.F. Risk of angina pectoris and plasma concentrations of vitamins A, C, and E and carotene. *The Lancet*, v.337, n.8732, p. 1-5, 1991.

- SAKCALI, M.S.; OZTURK, M. Eco-physiological behaviour of some Mediterranean plants as suitable candidates for reclamation of degraded areas. *Journal of Arid Environments*, v.57, n.2, p. 141-153, 2004.
- SANDLER, A.; MEUNIER, A.; VELDE, B. Mineralogical and chemical variability of mountain red/brown Mediterranean soils. *Geoderma*, v. 239, p. 156-167, 2015.
- SCHOINA, V.; TERPOU, A.; ANGELIKA-IOANNA, G.; KOUTINAS, A.; KANELLAKI, M.; BOSNEA, L. Use of *Pistacia terebinthus* resin as immobilization support for *Lactobacillus casei* cells and application in selected dairy products. *Journal of Food Science and Technology*, v.52, n.9, p. 5700-5708, 2015.
- SCHOINA, V.; TERPOU, A.; BOSNEA, L.; KANELLAKI, M.; NIGAM, P.S. Entrapment of *Lactobacillus casei* ATCC393 in the viscous matrix of *Pistacia terebinthus* resin for functional myzithra cheese manufacture. *LWT*, v.89, p. 441-448, 2018.
- SENKARDES, I.; TUZLACI, E. Some ethnobotanical notes from Gundogmus District (Antalya/Turkey). *Clinical and Experimental Health Sciences*, v.4, n.2, p.63-75, 2014.
- SENYAY-ONCEL, D.; ERTAS, H.; YESIL-CELIK, O. Effects of supercritical fluid extraction parameters on unsaturated fatty acid yields of *Pistacia terebinthus* berries. *Journal of the American Oil Chemists' Society*, v.88, n.7, p. 1061-1069, 2011.
- TASTEKIN, D.; TAMBAS, M.; KILIC, K.; ERTURK, K.; ARSLAN, D. The efficacy of *Pistacia terebinthus* soap in the treatment of cetuximab-induced skin toxicity. *Investigational New Drugs*, v.32, n.6, p. 1295-1300, 2014.
- TOPCU, G.; Ay, M.; BILICI, A.; SARIKURKCU, C.; OZTURK, M.; ULUBELEN, A. A new flavone from antioxidant extracts of *Pistacia terebinthus*. *Food Chemistry*, v.103, n.3, p. 816-822, 2007.
- TURTOLA, S.; MANNINEN, A. M.; RIKALA, R.; KAINULAINEN, P. Drought stress alters the concentration of wood terpenoids in Scots pine and Norway spruce seedlings. *Journal of Chemical Ecology*, v.29, n.9, p. 1981-1995, 2003.
- TURKOGLU, S.; CELIK, S.; KESER, S.; TURKOGLU, I.; YILMAZ, O. The effect of *Pistacia terebinthus* extract on lipid peroxidation, glutathione, protein, and some enzyme activities in tissues of rats undergoing oxidative stress. *Turkish Journal of Zoology*, v.41, n.1, p. 82-88, 2017.
- TUZLACI, E.; ERYASAR AYMAZ, E. Turkish folk medicinal plants, Part IV: Gönen (Balıkesir). *Fitoterapia*, v.72, p. 323-343, 2001.
- ULUKANLI, Z.; KARABORKLU, S.; OZTURK, B.; CENET, M.; BALCILAR, M. Chemical composition, antibacterial and insecticidal activities of the essential oil from the *Pistacia terebinthus* L. spp. *palaestina* (Boiss.) (Anacardiaceae). *Journal of Food Processing and Preservation*, v.38, n.3, p. 815-822, 2014.
- VERMA, N.; SHUKLA, S. Impact of various factors responsible for fluctuation in plant secondary metabolites. *Journal of Applied Research on Medicinal and Aromatic Plants*, v.2, n.4, p. 105-113, 2015.
- WEI, T.; SIMKO, V.; LEVY, M.; XIE, Y.; JIN, Y.; ZEMLA, J. Package 'corrplot'. *Statistician*, v.56, n.316, p. e24, 2017.
- WHITEHOUSE W.E. The pistachio nut – a new crop for the Western United States. *Econ. Bot.*, v.11, p. 281-321, 1957.
- WICKHAM, H. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2009.
- YESILADA, E.; HONDA, G.; SEZIK, E.; TABATA, M.; FUJITA, T.; TANAKA, T.; TAKEDA, Y.; TAKAISHI, Y. Traditional medicine in Turkey. V. Folk medicine in the inner Taurus Mountains. *J. Ethnopharmacol*, v.46, p. 133-152, 1995.
- YILMAZ, O.; OZSAHIN, A.D.; BIRCAN, B.; ERDEN, Y.; KARABOGA, Z. Radical Scavenging activity of the *Pistacia terebinthus* in fenton reagent environment and its protective effects on the unsaturated fatty acids. *Asian Journal of Chemistry*, v.22, n.10, p.7949-7958, 2010.
- ZOHARY, M. A monographical study of the genus *Pistacia*. *Palestine Journal of Botany (Jerusalem Series)*, v.5, n.4, p. 187-228, 1952.