



## Genetic characterization and phylogeography of the wild boar *Sus scrofa* introduced into Uruguay

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### Abstract

The European wild boar *Sus scrofa* was first introduced into Uruguay, in southern South America during the early decades of the last century. Subsequently, and starting from founder populations, its range spread throughout the country and into the neighbouring Brazilian state Rio Grande do Sul. Due to the subsequent negative impact, it was officially declared a national pest. The main aim in the present study was to provide a more comprehensive scenario of wild boar differentiation in Uruguay, by using mtDNA markers to access the genetic characterization of populations at present undergoing rapid expansion. A high level of haplotype diversity, intermediate levels of nucleotide diversity and considerable population differentiation, were detected among sampled localities throughout major watercourses and catchment dams countrywide. Phylogenetic analysis revealed the existence of two different phylogroups, thereby reflecting two deliberate introduction events forming distantly genetic lineages in local wild boar populations. Our analysis lends support to the hypothesis that the invasive potential of populations emerge from introgressive hybridization with domestic pigs. On taking into account the appreciable differentiation and reduced migration between locales in wild boar populations, management strategies could be effective if each population were to be considered as a single management unit.

*Key words:* Uruguayan wild boar.

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### Introduction

The European wild boar *Sus scrofa* was first introduced into Uruguay, in southern South America, during the early decades of the last century. During the 1920's, Aaron de Anchorena, an Argentinean landowner, introduced a number of wild boars onto his ranch in the south-western Department of Colonia (one of the administrative divisions in Uruguay) for hunting purposes (Figure 1a). The founder population, on encountering adequate environmental conditions, and through specific dispersal capacity and generalist predator habits, began to increase in numbers and widely expand its range. The present day wild boar population is presumed to comprise a cross-breed with domestic pigs (Figure 1b), thereby giving rise to great variability in phenotypes, albeit with a predominance of wild boar characteristics (Herrero and Fernandez de Luco, 2003). The quick expansion of the animal's range was facilitated by the locally mild climate, dense network of rivers, forest corridors, and an abundant food source of cultivated crops and

vulnerable domestic animals, together with the absence of natural predators. Nowadays, species distribution is widespread, having started in the west, the main agricultural zone, from there later extending to central and eastern parts of the country, and in the neighbouring Brazilian state Rio Grande do Sul.

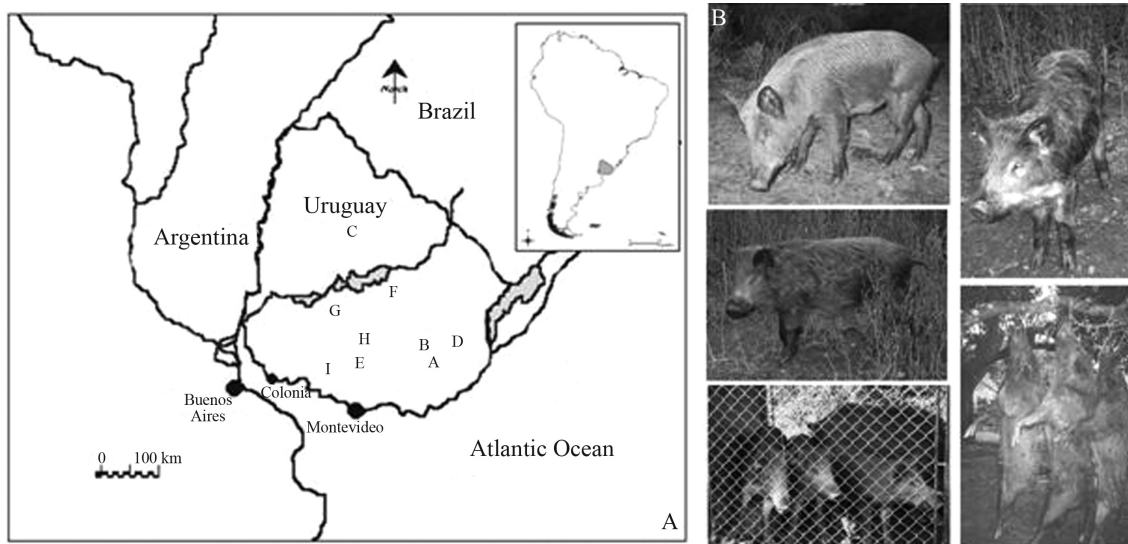
The species has been extensively hunted over recent years. Some landowners have a highly negative perception of wild boars, regarding them as being responsible for direct predation of sheep, even though such was not verified and quantified and only apparent by interviewing farmers (Herrero and Fernandez de Luco, 2003). In 1982, the animal was officially declared a national pest (Decree 463/982).

Feral pigs are also potential reservoirs or vectors for a number of endemic and exotic diseases capable of affecting domestic livestock, wildlife and even humans. In Australia, for example, besides leptospirosis and brucellosis, they are also capable of transmitting exotic diseases, such as foot-and-mouth disease and Japanese encephalitis (Dexter, 2003; Caley and Hone, 2004).

Along with knowledge of spatial genetic structure, that of population dispersal is also essential for reducing and reversing environmental impacts (Hampton *et al.*, 2004), particularly so in the development of effective con-

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**Figure 1** - A. Distribution map of Uruguayan wild boar collecting sites: A- Neighborhood of Velazquez (Rocha Department); B- Rio Cebollatí (Lavalleja Department); C- A° Malo (Tacuarembó Department); D- Neighborhood of Lazzcano (Rocha Department); E- A° Aries (Florida Department); F- A° Saucos (Durazno Department); G- R. Negro (Durazno Department); H- Neighborhood of Sarandí del Yí (Durazno Department); I- Neighborhood of San José (San José Department). B. Various phenotypes of captured Uruguayan wild boars from 9 localities and captives from a farm reserve at Velazquez. Scale bar: 100 km.

control programmes for feral or invasive species, and for obtaining informative and reliable risk analysis (Edwards *et al.*, 2004).

A major genetic paradox in invasive species is resolving how bottlenecked populations, with typically low genetic diversity, low evolutionary potential and, possibly, reproductive fitness, can become invasive (Frankham, 2005). The wild boar from Uruguay thus constitutes an interesting species-model for solving the issue, besides representing an unusual challenge for encountering local management strategies.

New approaches, using contemporary molecular techniques, in conjunction with demographic data, can be extremely useful for a better comprehension of the dynamics, population structure and social biology of many invasive species (Taylor *et al.*, 2000).

Among genetic markers, highly polymorphic animal mitochondrial DNA (mtDNA), almost exclusively maternally inherited and without genetic recombination, constitutes a powerful tool for a population genetic approach. The clonal transmission of mtDNA haplotypes facilitates the discrimination of matrilineage within species, the sequence analysis of their most variable regions being useful for investigating the genetic origin of animal populations and breeds, and thus, domestication processes in livestock species themselves (Bradley *et al.*, 1996; Luikart *et al.*, 2001).

The main aim of this investigation is to provide a more comprehensive scenario of wild boar differentiation in Uruguay, using mtDNA population markers and phylogenetic analysis for testing possible hypotheses regarding their rapid expansion. Furthermore, this is a first-time report containing genetic information, with recommenda-

tions for more effective control strategies of feral pigs in Uruguay.

## Material and Methods

### Sampling and DNA extraction

This phylogeographic study of the Uruguayan wild boar included specimens (Appendix 1) mostly from the southern, central, northeastern and eastern regions of the country, covering the geographic range of expansion, starting from the introduction site in the Department of Colonia (Figure 1). The remainder are sequences retrieved from GenBank, and pertaining to Spanish, Italian, central European and Japanese wild boars, as well as commercial pig breeds, viz., Large White, Landrace, Duroc and Pietrain (Supplementary Material, Table S1). Tissues of voucher specimens were deposited in the Sección Genética Evolutiva, Facultad de Ciencias, Uruguay. Outgroup analysis included individuals pertaining to two additional taxa *Sus verrucosus* and *Phacochoerus africanus*.

Genomic DNA was isolated from liver and muscle tissues of freshly sacrificed animals (fixed in ethanol 95%), using an extraction with a sodium chloride protein precipitation, followed by ethanol DNA precipitation (modified from Miller *et al.*, 1988).

### Mitochondrial cytochrome b (cyt b) sequences

A 661-bp fragment of *cyt b* gene between sites 14,695 and 15,355 was amplified using forward and reverse primers, as described by Alves *et al.* (2003), in 30 cycles of 94 °C for 45 s, 53 °C for 45 s, 72 °C for 1 min; 30 cycles. PCR products were cleaned with MARLIGEN Kit (Biosci-

ences inc.) Rapid PCR Purification System, to then undergo sequencing using amplification primers with a Perkin-Elmer ABI Prism 377 automated sequencer. The final sequences for analysis were obtained by reconciling chromatograms for light and heavy DNA strands. Sequence alignment was performed using the CLUSTAL X program (Thompson *et al.*, 1997).

#### Data analysis and DNA polymorphism

Nucleotide composition and substitution patterns were calculated using the MEGA (Kumar *et al.*, 2004) and DNASP4 (Rozas *et al.*, 2003) computer programs. Corrected estimates of pairwise sequence divergence were obtained using the two-parameter algorithm (K2P) of Kimura (1980) implemented into MEGA. Population DNA polymorphism was measured by calculating the proportion of segregating sites (S), haplotype diversity (Nei, 1987; p. 179), and nucleotide diversity  $\pi$  (Nei, 1987; p.257), using ARLEQUIN (Schneider *et al.*, 2000) and DNASP4 (Rozas *et al.*, 2003) software packages.

In order to evaluate neutrality departure in the data, Tajimas D (Tajima, 1989) was calculated using the DNASP4 (Rozas *et al.*, 2003) software package, as a way of testing any significant excess of low-frequency haplotypes.

#### Phylogenetic analyses

Two methods of phylogenetic reconstruction viz., maximum-parsimony (MP) and neighbor-joining (NJ), were employed to define phylogeographic association among mitochondrial sequences, using for the purpose PAUP\*4.0b8 (Swofford, 1998). Equally weighted maximum-parsimony analysis was undertaken by way of heuristic search (MULPARS option, stepwise addition, tree-bisection-reconnection [TBR] branch swapping, 100 replications). Strict consensus between rival trees was computed to reconcile equally parsimonious topologies. Distance trees were generated using a Hasegawa *et al.* (1985) model, taking into consideration differences among transversion and transition substitutions, as well as those among base frequencies. The neighbour-joining method (Saitou and Nei, 1987) was employed for phylogenetic reconstruction. In the case of both methods (MP and NJ), the degree of confidence assigned to nodes in trees was assessed by bootstrapping with 1000 replicates. All the trees were rooted by means of the outgroup criterion.

#### Analysis of Molecular Variance and Nested Clade Analysis (NCA)

In order to examine genetic structuring among Uruguayan wild boar populations, variance components among hierarchical partitions in the data set were assessed through the Analysis of Molecular Variance (AMOVA) developed by Excoffier *et al.* (1992). The Euclidean metric (Excoffier *et al.*, 1992) was used to construct the matrix of pairwise distances. Various grouping hypotheses were proposed for

analyzing the hierarchical partitioning of genetic variation. Three among these were retained, viz., 1) all the haplotypes were gathered into a single group, 2) haplotypes from two neighbouring eastern sites and distributed into the corresponding groups (B and D in Figure 1) vs. all the remaining geographic localities, 3) the haplotypes were assigned to three regions (southern, central-northeastern and eastern Uruguay) representing the diverse Uruguayan basins, as a means of recuperating biogeographic information.

The existence of geographic association among haplotypes was assessed by NCA (Templeton *et al.*, 1995). The *cyt b* haplotype network in Uruguayan wild boars was estimated by using the statistical parsimony method, with the algorithm described by Templeton *et al.* (1992). Accordingly, the cladogram for finding haplotype connections with probabilities above the 0,95 confidence level was constructed using the TCS 1.06 (Clement *et al.*, 2000) program. Statistics related to data distances, viz., internal clade (Dc), between clades (Dn) and between interior and tip clades (I-T), were generated by exact permutation contingency analysis of clades within nested categories as against their respective geographic locality (Templeton *et al.*, 1995), using in the process 10,000 permutations of nesting clades versus sampling localities, and assuming as recommended statistical significance,  $\alpha = 0.05$ . Results obtained from GEODIS were then interpreted, using the revised Posada and Templeton (2008) inference key to elucidate alternative historical scenarios of wild boar differentiation.

Population subdivision was measured by assuming the infinite mutation model (Kimura and Crow, 1964) and calculating  $F_{ST}$  (Slatkin, 1991) for the whole population. Pairwise estimates of  $F_{ST}$  were calculated using Arlequin (Schneider *et al.*, 2000) to generate pairwise estimates of gene flow levels, as follows:  $N_m \approx 1/2 [(1/F_{ST}) - 1]$  (Wright, 1951).

## Results

### Mitochondrial *cyt b* diversity and population genetic analysis

Mitochondrial cytochrome b sequences pertaining to Uruguayan, European and Japanese wild boars, as well as commercial breeds of *Sus scrofa*, were included for analysis. Most of those from the Uruguayan samples were new for the wild boar *Sus scrofa*, the remainder having been retrieved from GenBank (Table S1). Among the 571 bp analyzed from Uruguayan wild boar samples, 47 variables and 28 phylogenetic informative sites were found in the data set, apart from 39 polymorphic segregating sites and 14 haplotypes in the sample itself. Haplotype diversity was high (0.97 - SD = 0.032), whereas intermediate levels of nucleotide diversity were found (0.014 - SD = 0.021). With the exception of haplotypes 1 (shared by 2 individuals), 2 and 11 (both shared by three individuals), the remainder were carried by a single individual.

The estimated average rate of transitional/transversional substitution ( $r = si/sv$ ) in the wild boar *cyt b* gene was 2:1 (1<sup>st</sup>  $r = 1.7$ ; 2<sup>nd</sup>  $r = 0.5$  and 3<sup>rd</sup>  $r = 2.8$  codon positions). Among the Uruguayan wild boar samples, deduced amino acid sequences showed only 20 of 190 amino acids to be variable and 9 phylogenetically informative sites.

Corrected pairwise K2P sequence divergence among the sampled Uruguayan wild boars is presented in Table 1. The average genetic distance was 1.7% (SD = 0.003), thus remarkably higher than that found among European (0.3% - SD = 0.001), and Japanese and Israeli (1.3% - SD = 0.005) wild boars, as well as the analyzed commercial breeds (0.9% - SD = 0.003). The maximum divergence by including outgroup taxa was 8.4% (SD = 0.009).

No significant excess of low-frequency haplotypes among Uruguayan wild boars ( $D = -1.418$   $p > 0.10$ ) was revealed by Tajima D'test.

### Phylogenetic analyses

The same two evolutionary sister clades that constitute different phylogroups were identified by maximum parsimony and distance analysis. In maximum parsimony analysis, strict consensus (Figure 2) resulted in the 100 most parsimonious trees (156 steps), thereby showing a major and monophyletic clade integrated by 12 Uruguayan wild boar haplotypes. They collapsed into a basal polytomy joining all the haplotypes belonging to European wild boars, together with Landrace and Large White commercial breeds. A minor clade integrated by the Uruguayan wild boar haplotypes 9 and 10 collapsed together with a Japanese wild boar haplotype and the Duroc and Pietrain commercial breeds. All the major clades received high bootstrap values of 80%-100% in both phylogenetic analyses.

### AMOVA analysis and geographic distribution of genetic variation

Statistical parsimony network based on the *cyt b* gene (Figure 3), revealed the strong genetic structuring among haplotypes of the Uruguayan wild boar showing different phylogroups. The total cladogram includes five levels of nested hierarchical clades presenting 11 maximum connection steps at 95%. At a higher level of hierarchy, a major clade (4-3) of haplotypes remains connected by few mutational steps, thereby retaining a central position in the network. This major clade includes those locales in central Uruguay, in the neighborhood of the site in the Department of Colonia (Figure 1, sites I, E, H, G and F), the center of expansion throughout the central region itself, as well as the northeastern (C, Figure 1) and eastern (A, Figure 1). Haplotype 5 relates this clade (4-3) to other more distant haplotypes that integrate clade 4-1. This haplotype corresponds to locale F (Figure 1) in the central region. Furthermore, two other genetically more distant haplotypes (9 and 10) represent a divergent clade (4-2 in Figure 3) in the eastern region, separated by ninety and ten step mutations, respectively, from the remainder. Finally, nested contingency analysis of almost all clade levels revealed no significant association of clades and geographic distances.

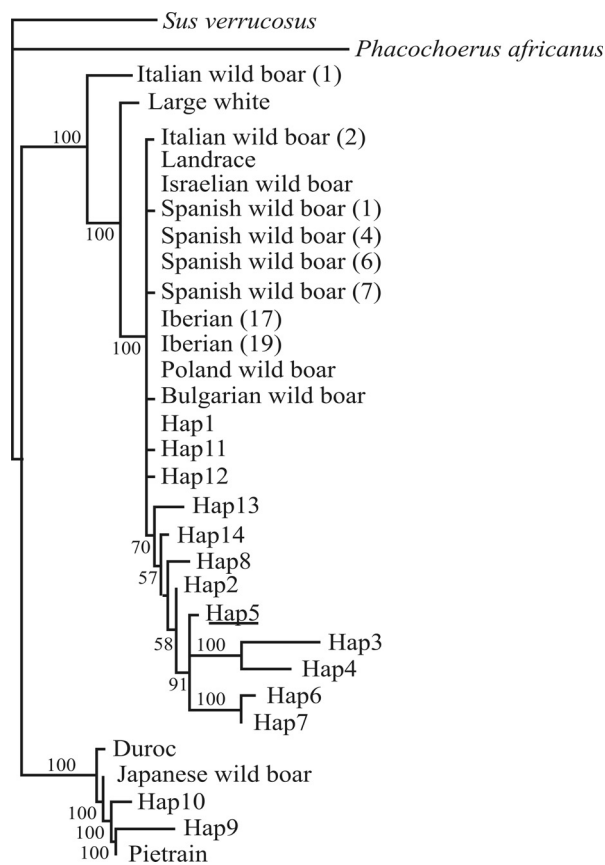
AMOVA results compiling the three retained hypotheses are shown in Table 2. Under the two-group hypothesis (2), most genetic variation among *cyt b* haplotypes was distributed among-groups ( $\Phi_{CT} = 0.673$ ). All the other tested structuring hypotheses failed to provide a more reasonable explanation for maximization of among-group hierarchical molecular variation.

Indirect estimates of pairwise  $F_{ST}$  values revealed almost complete genetic isolation among locales, except for

**Table 1** - Corrected genetic distances among 14 haplotypes of the Uruguayan wild boar *Sus scrofa*, according to the Kimura 2-P model (below the diagonal), and Standard Deviation (SD) estimated by the bootstrap method (above the diagonal).

	hap1	hap2	hap3	hap4	hap5	hap6	hap7	hap8	hap9	hap10	hap11	hap12	hap13	hap14
hap1	-	0.002	0.027	0.029	0.005	0.011	0.007	0.005	0.022	0.016	0.002	0.002	0.002	0.004
hap2	0.002	-	0.029	0.027	0.004	0.013	0.009	0.004	0.024	0.018	0.004	0.004	0.004	0.002
hap3	0.007	0.008	-	0.027	0.029	0.027	0.024	0.033	0.042	0.044	0.029	0.029	0.029	0.031
hap4	0.007	0.007	0.007	-	0.024	0.033	0.029	0.027	0.052	0.046	0.031	0.031	0.031	0.029
hap5	0.003	0.002	0.008	0.007	-	0.009	0.005	0.007	0.027	0.022	0.007	0.007	0.007	0.005
hap6	0.004	0.005	0.007	0.008	0.004	-	0.004	0.016	0.022	0.027	0.013	0.013	0.013	0.014
hap7	0.003	0.004	0.007	0.008	0.003	0.002	-	0.013	0.022	0.024	0.009	0.009	0.009	0.011
hap8	0.003	0.002	0.008	0.007	0.003	0.005	0.005	-	0.027	0.022	0.007	0.007	0.007	0.005
hap9	0.006	0.007	0.010	0.011	0.007	0.006	0.006	0.007	-	0.009	0.024	0.024	0.024	0.025
hap10	0.005	0.006	0.010	0.010	0.006	0.007	0.007	0.007	0.004	-	0.018	0.018	0.018	0.020
hap11	0.002	0.002	0.008	0.008	0.003	0.005	0.004	0.003	0.007	0.006	-	0.004	0.004	0.005
hap12	0.002	0.002	0.008	0.008	0.003	0.005	0.004	0.003	0.007	0.006	0.002	-	0.004	0.005
hap13	0.002	0.002	0.008	0.008	0.003	0.005	0.004	0.003	0.007	0.006	0.002	0.002	-	0.005
hap14	0.002	0.002	0.008	0.008	0.003	0.005	0.004	0.003	0.007	0.006	0.003	0.003	0.003	-

site A, with considerable genetic exchange with the remainder. On the other hand, all were genetically isolated from B and D.



**Figure 2** - Maximum parsimony phylogenetic relationships based on the cytochrome b dataset of haplotypes of Uruguayan, European and Japanese wild boars, as well as sequences from commercial breeds of *Sus scrofa*. The strict consensus resulted in 100 of the shortest most parsimonious trees (156 steps). All the trees were rooted by using *Sus verrucosus* and *Phacochoerus africanus* as outgroups. Bootstrap values above 50% are shown on the relevant nodes.

**Table 2** - Analysis of molecular variance (AMOVA) in the Uruguayan wild boar *Sus scrofa*. Hierarchical partition of genetic variation into three components: among groups ( $\Phi_{CT}$ ), among populations within groups ( $\Phi_{CS}$ ), and among individuals within populations ( $\Phi_{ST}$ ), disregarding either their original populations or groups. Among the tested hypotheses, three were selected: a) including all the collecting sites into just one group; b) forming two groups of populations, consisting of all the collection sites vs. Rio Cebollatí and Lascano; c) the separation into three groups of samples pertaining to the various river basins, viz., the eastern, central-northeastern and southern. The highlighted line corresponds to values which maximal among-group differentiation.

Hypothesis	Source of variation	df	Sum of squares	Variance components	Percentage of variation	$\Phi$ statistics
a	Among groups	8	26.140	0.35864Va	11.71	-
	Among populations within groups	-	-	-	-	-
	Within populations	8	21.625	2.70312 Vb	88.29	$\Phi_{ST} = 0.11714$
b	Among groups	1	15.731	0.01813 Va	67.39	$\Phi_{CT} = 0.67394$
	Among populations within groups	7	10.408	-0.76469Vb	-12.86	$\Phi_{SC} = -0.39449$
	Within populations	8	21.625	2.70312 Vc	45.47	$\Phi_{ST} = 0.54531$
c	Among groups	2	7.248	0.01698Va	0.55	$\Phi_{CT} = 0.00554$
	Among populations within groups	6	18.892	0.34564 Vb	11.27	$\Phi_{CS} = 0.11337$
	Within populations	8	21.625	2.70312 Vc	88.17	$\Phi_{ST} = 0.00554$

## Discussion

### Mitochondrial cytochrome b variation in Uruguayan wild boar populations

Present results represent the first population genetic characterization of Uruguayan wide-ranging wild boars, when using the mitochondrial cytochrome b gene. High levels of haplotype diversity, intermediate levels of nucleotide diversity, and considerable population differentiation among sampled localities throughout major watercourses and catchment dams, were detected (Figure 1).

Intermediate nucleotide diversity was similar to that reported for Artiodactyla, taxa, when using mtDNA cytochrome b sequences in a comparative analysis of various mammalian orders and families (Nabholz *et al.*, 2008).

According to the present study, levels of corrected sequence divergence among Uruguayan wild boar haplotypes, although higher than in the European wild type and commercial breeds, was similar to the Japanese and Israeli. In previous studies, when considering both *cyt b* and the mtDNA control region, an appreciable genetic distance ( $1.2 \pm 0.09\%$ ) between European wild boars (Italy and Poland) and Asian (Israel), was found (Giuffra *et al.*, 2000), therefore consistent with the divergence found in the Uruguayan wild boar data set. In contrast, on analyzing mitochondrial control region data from domestic pigs, Fang and Andersson (2006) reported low genetic divergence in all the Chinese mtDNA haplotypes (mean  $\pm$  SD,  $0.006 \pm 0.001$ ), as well as all the European (mean  $\pm$  SD,  $0.005 \pm 0.001$ ).

All accumulated data, plus the average si/sv ratio detected in the *cyt b* fragment, indicate that this molecular marker, besides revealing no evidence of among-site saturation, represents a useful tool for genetically characterizing the recently introduced feral pig in Uruguay.

### Population structuring in Uruguayan wild boar populations

Phylogenetic analysis (Figure 2) revealed the existence of two different phylogroups among the Uruguayan wild boar haplotypes. The major of the two comprises all those, with the exception of 9 and 10, pertaining to the different localities in southern, central and northeastern Uruguay. This clade collapsed into a basal polytomy with all the sequences from European wild boars, and Landrace and Large White domestic pigs. The minor phylogroup, comprised of haplotypes 9 and 10, joins the sequences from Pietrain and Duroc domestic pigs, as well as the Japanese wild boar. Generally speaking, the statistical parsimony network (Figure 3) showed the same genealogical history, although here the major phylogroup is divided into two clades, 4-1 (haplotypes 3 and 4) and 4-3 (this including the remaining haplotypes of the group). Consequently, clade 4-1 corresponds to a minor derivative monophyletic clade (integrated by haplotypes 3 and 4) in the phylogenetic tree (Figure 2). On the other hand, the minor phylogroup in phylogenetic reconstruction (Figure 2) was consistent with clade 4-2 in the statistical parsimony network (Figure 3).

The present analysis supports the hypothesis of two different deliberate introduction events of distantly related genetic lineages in the Uruguayan wild boar populations. The distribution of the major clade is consistent with the historical introduction of the European wild boar into the Colonia Department, and its dispersal throughout localities in southern, central and northeastern Uruguay, by way of the principal basins and watercourse of the Rio Santa Lucía and the Río Negro. The second introduction, occurred in southeastern Uruguay near the Cebollatí river in the Merin lagoon basin, the border between Uruguay and the Brazilian state Rio Grande do Sul, may have been a deliberate introduction since southern Brazil. Further analysis, including samples from these neighbouring localities, could possibly clarify the issue.

Moreover, two major hypotheses would explain the association of the Uruguayan wild boar with others populations of different origins, as well as with different strains of domestic pigs, *i.e.*, the existence of ancestral polymorphisms and/or multiple introgressive hybridization events, as possible, although not mutually exclusive, scenarios. These scenarios are consistent with the level of genetic variation accumulated in the Uruguayan wild boar populations. The evidence of ancestral polymorphism has been a plausible hypothesis since 1920, when the first introduction of wild boars of unknown origin was reported (Herrero and Fernández de Luco, 2003). In this case, various types of European wild boars would be involved, as can be inferred on examining taxa association in the major clade (Figure 2). The presence of lung parasites of the genus *Metastrongylus* sp., quite common in European wild boars, is notable in the Uruguayan wild boar and could give support to this possible origin. Nevertheless, it is difficult to explain the associ-

ation with the Japanese wild boar, Duroc and Pietrain sequences in the minor clade, through the lack of additional information regarding neighbouring south Brazilian wild boar populations.

The hypothesis of hybridization and introgression is well supported in phenotypical characterization among current Uruguayan wild boar populations (Herrero and Fernández de Luco, 2003). These authors proposed their emergence from cross-breeding between wild boars and domestic pigs, hence the wide diversity in phenotypes, with a predominance of characteristics from the former. Consequently, some individuals have white hair on the feet, the underside of the neck, tarsus and carpus, drooping ears and jet black or red tails, whereas others present outstanding and fast growth, great intrapopulation morphological variability and considerable accumulation of subcutaneous fat (Herrero and Fernández de Luco, 2003).

According to Lee (2002), the inter- or intraspecific hybridization of an invasive population with native or non-native populations would alleviate the loss of additive genetic variance during founder events, and thus generate novel genotypes. Numerous studies have documented the positive effects of hybridization on invisibility, such as faster growth, greater size and increased aggression. All these characteristics were encountered in Uruguayan wild boar populations (Herrero and Fernández de Luco, 2003). Lee (2002) also proposed successful invasion to be a probable result of advantageous selection among numerous hybrid combinations.

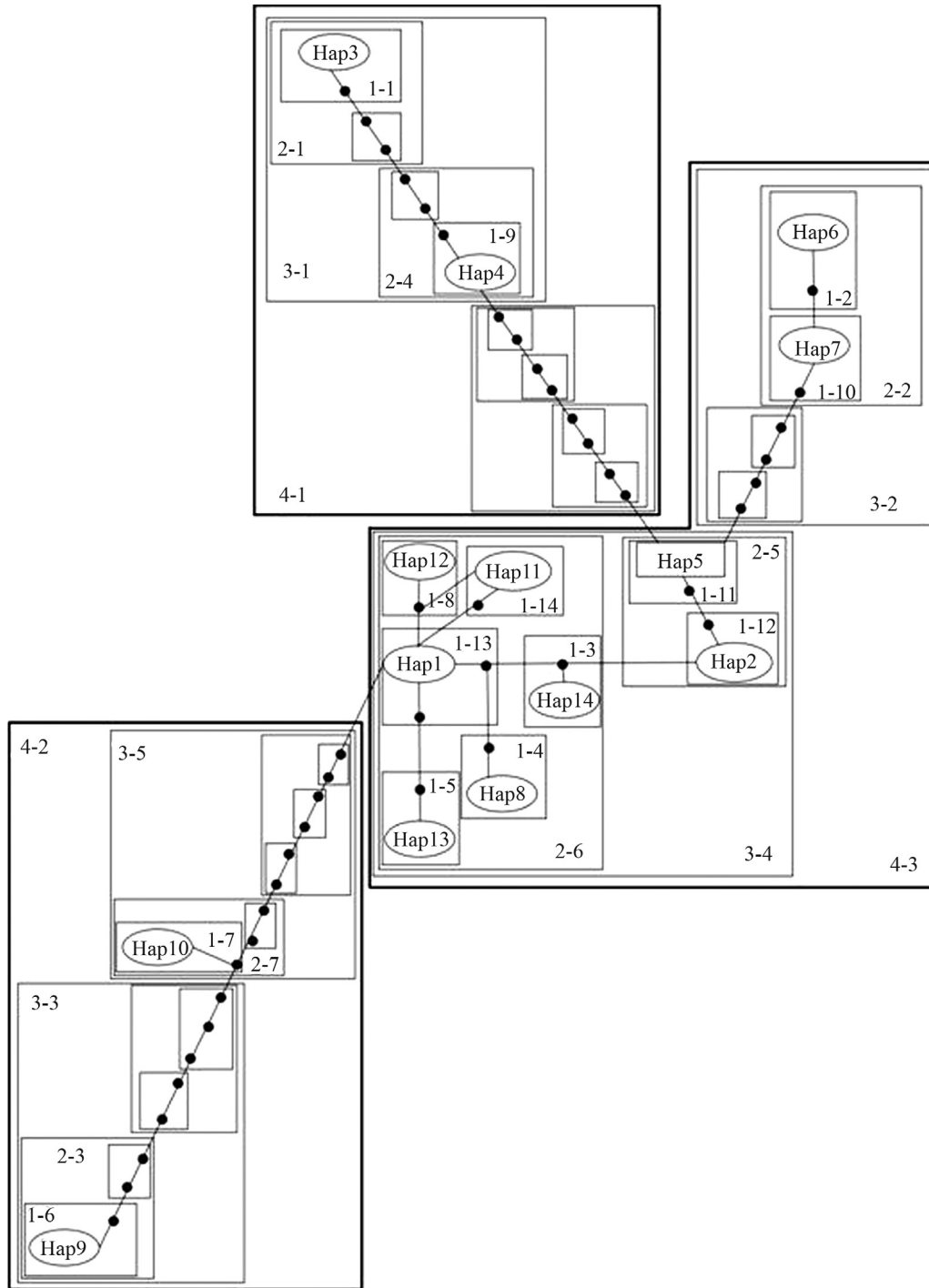
According to Avise (2000), high haplotype and low nucleotide diversities infer rapid population growth from an ancestral population with small  $N_e$ . Current results concerning mtDNA polymorphism parameters could conform to this interpretation. Moreover, Frankham (2005) postulated that certain invasive species possess elevated genetic diversity and the enhanced ability to evolve when invading novel localities. All present population genetic data pointed out that hybridization between introduced wild boars and domestic pigs could be a plausible explanation of the invasive potential of these cross-bred populations.

AMOVA analysis (Table 2) confirmed the considerable population differentiation among sampled localities, as well as the existence of two unexpected, different and highly structured evolutionary lineages among Uruguayan wild boars. No genetic exchange was detected among individuals belonging to these different phylogroups. Moreover, indirect estimates of gene flow revealed no homogeneity among all the sampled localities. In fact, with the exception of the neighboring B and D, only locale A (Figure 1) showed any considerable genetic exchange with the remainder. Nevertheless, this is an expected population scenario, seeing that this locality included a mixed group captured from other distant sites in the country, thereby representing a semi-captive wild boar stock.

The levels of differentiation found are consistent with existing knowledge of the regional distribution and biology of the Italian wild boar (Vernesi *et al.*, 2003). There was also a certain similarity with the population structure of feral pigs in southwestern Australia (Hampton *et al.*, 2004), whereby dispersal rates between, but not within, the inferred feral pig

populations were relatively low. According to the relatively small home range of feral pigs in this region (Choquenot *et al.*, 1996), a high level of genetic structuring was not unexpected even between populations that were only 25 km apart.

A similar dispersal pattern was encountered in neighbouring wild boar localities (A vs. B and D) which, al-



**Figure 3** - Statistical parsimony network and the corresponding nested design for cytochrome b haplotypes in Uruguayan wild boars. The cladogram was estimated under 95% statistical limits of parsimony using the algorithm by Templeton *et al.* (1992). The oval includes haplotype numbers (0-step clades). Solid circles represent hypothetical haplotypes. Thin-lined polygons enclose 1-step to 3-step clades and thick-lined polygons enclose 4-step clades, all within the total cladogram. Specimens and the respective haplotype are listed in Table S1, Supplementary Material.

though only around 60 km apart, remained genetically isolated. Further population genetics analysis, using nuclear markers (*i.e.* microsatellites, SNPs), could be a means of clarifying aspects of social organization, dispersal and possible asymmetric gene flow among populations in the Uruguayan wild boar. Even so, recent studies on wild boar from Tuscany (Italy) did not reveal that the predicted matrilinearity in wild boar social units. In this study, aggregations of unrelated adult females were detected, thereby indicating a low degree of within-group relatedness (Iacolina *et al.*, 2009).

### Population genetics analysis in Uruguayan wild boars and management strategies

Present population genetics has contributed to the strategic management of Uruguayan wild boar populations. Considering both the high levels of differentiation and the normally low migration rates among localities in these populations, management strategies would be more effective if each population were to be considered as a single management unit. A similar *modus operandi* has already been proposed for other feral pig populations (Hampton *et al.*, 2004). The failure to recognize these units could result in the inevitable recurrent invasion of controlled areas. On the other hand, recurrence from neighbouring pig populations would be relatively slow, due to the low migration rate between discrete or adjacent localities.

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### References

- Alves E, Ovilo C, Rodríguez MC and Siló L (2003) Mitochondrial DNA sequence variation and phylogenetic relationships among Iberian pigs and other domestic and wild pigs populations. *Anim Genet* 34:319-324.
- Avice JC (2000) *Phylogeography: The History and Formation of Species*. Harvard University Press, Cambridge, 447 pp.
- Bradley DG, MacHugh DE, Cunningham P and Loftus RT (1996) Mitochondrial diversity and the origins of African and European cattle. *Proc Natl Acad Sci USA* 93:5131-5.
- Caley P and Hone J (2004) Disease transmission between and within species, and the implications for disease control. *J Appl Ecol* 41:94-104.
- Choquenot D, McIlroy J and Korn T (1996) *Managing Vertebrate Pests: Feral Pigs*. Australian Publishing Service, Canberra, 163 pp.
- Clement M, Posada D and Crandall KA (2000) TCS: A computer program to estimate gene genealogies. *Mol Ecol* 9:1657-1659.
- Decreto 463/1982 del Poder Ejecutivo del 15/12/1982. Declara plaga nacional al jabalí europeo y autoriza su libre caza en todo el territorio nacional. *Diario Oficial* 21388-500-A.
- Dexter N (2003) Stochastic models of foot and mouth disease in feral pigs in the Australian semi-arid rangelands. *J Appl Ecol* 40:293-306.
- Edwards GP, Pople AR, Saalfeld K and Caley P (2004) Introduced mammals in Australian rangelands: Future threats and the role of monitoring programmes in management strategies. *Austral Ecol* 29:40-50.
- Excoffier L, Smouse PE and Quattro JM (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: Application to human mitochondrial DNA restriction data. *Genetics* 131:479-491.
- Fang M and Andersson L (2006) Mitochondrial diversity in European and Chinese pigs is consistent with population expansions that occurred prior to domestication. *Proc R Soc Lond B Biol Sci* 273:1803-1810.
- Frankham R (2005) Resolving the genetic paradox in invasive species. *Heredity* 94:385.
- Giuffra E, Kijas JMH, Amarger LV, Carlborg O, Jeon JT and Andersson L (2000) The origin of the domestic pig: Independent domestication and subsequent introgression. *Genetics* 154:1785-1791.
- Hampton JO, Spencer PBS, Alpers DL, Twigg LE, Woolnough AP, Doust J, Higgs T and Pluske J (2004) Molecular techniques, wildlife management and the importance of genetic population structure and dispersal: A case study with feral pigs. *J Appl Ecol* 41:735-743.
- Hasegawa M, Kishino H and Yano T (1985) Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J Mol Evol* 22:160-174.
- Herrero J and Fernández de Luco D (2003) Wild boars (*Sus scrofa* L.) in Uruguay: Scavengers or predators? *Mammalia* 67:485-491.
- Iacolina L, Scandura M, Bongi P and Apollonio M (2009) Non kin associations in wild boar social units. *J Mammal* 90:666-674.
- Kimura M (1980) A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 16:111-120.
- Kimura M and Crow JF (1964) The number of alleles that can be maintained in a finite population. *Genetics* 49:725-738.
- Kumar S, Tamura K and Nei M (2004) MEGA3: Integrated software for molecular evolutionary genetics analysis and sequence alignment. *Brief Bioinform* 5:150-163.
- Lee CE (2002) Evolutionary genetics of invasive species. *Trends Ecol Evol* 17:386-391.
- Luikart G, Gielly L, Excoffier L, Vigne JD, Bouvet J and Taberlet P (2001) Multiple maternal origins and weak phylogeographic structure in domestic goats. *Proc Natl Acad Sci USA* 98:5927-5932.
- Miller SA, Dikes DD and Polesky HF (1988) A simple salting out procedure for extracting DNA for human nucleated cells. *Nucleic Acids Res* 16:215.
- Nabholz B, Mauffrey JF, Bazin E, Galtier N and Glemin S (2008) Determination of mitochondrial genetic diversity in mammals. *Genetics* 178:351-361.
- Nei M (1987) *Molecular Evolutionary Genetics*. Columbia University Press, New York, 512 pp.



- Rozas J, Sánchez-Delbarrio JC, Messeguer X and Rozas R (2003) DnaSP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* 19:2496-2497.
- Saitou N and Nei M (1987) The neighbour-joining method: A new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4:406-425.
- Schneider S, Roessli D and Excoffier L (2000) Arlequin: A software for population genetics data analysis. University of Geneva, Switzerland.
- Slatkin M (1991) Inbreeding coefficients and coalescence times. *Genet Res* 58:167-175.
- Swofford DL (1998) PAUP\* (Phylogenetic Analysis Using Parsimony). v. 4. Sinauer, Sunderland.
- Tajima F (1989) Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* 123:585-595.
- Taylor AC, Cowan PE, Fricke BL and Cooper DW (2000) Genetic analysis of the mating system of the brushtail possum (*Trichosurus vulpecular*) in New Zealand farmland. *Mol Ecol* 9:869-879.
- Templeton AR, Crandall KA and Sing CF (1992) A cladistic analysis of phenotypic associations with haplotypes inferred from restriction endonuclease mapping and DNA sequence data. III. Cladogram estimation. *Genetics* 132:619-633.
- Templeton AR, Routman E and Phillips CA (1995) Separating population structure from population history: A cladistic analysis of the geographical distribution of mitochondrial DNA haplotypes in the Tiger Salamander, *Ambystoma tigrinum*. *Genetics* 140:767-782.
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F and Higgins DG (1997) The CLUSTAL\_X windows interface: Flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* 25:4876.
- Vernes C, Crestanello B, Pecchioli E, Tartari D, Carnelli D, Hauffe H and Bertorelle G (2003) The genetic impact of demographic decline and reintroduction in the wild boar (*Sus scrofa*): A microsatellite analysis. *Mol Ecol* 12:585-595.
- Wright S (1951) The genetical structure of populations. *Ann Eugenics* 15:323-354.

## Internet Resources

- Posada D and Templeton AR (2008) Inference Key for the Nested Haplotype Tree Analysis of Geographical Distance, <http://darwin.uvigo.es/software/geodis.html> (December, 2008).

## Supplementary Material

The following online material is available for this article:

Table S1 - List of specimen and haplotypes from cytochrome b sequences.

This material is available as part of the online article from <http://scielo.br/gmb>.

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**Table S1** - List of specimen, the corresponding number of haplotypes from cytochrome b sequences; Collection Institutional Code of specimens sampled in the present study from Uruguay, and the corresponding GenBank accession numbers. Additional sequences from Spanish, Italian, central European and Japanese wild boars, as well as commercial breeds (Large White, Landrace, Duroc and Pietrain), besides two outgroup taxa, were retrieved from the GenBank.

Species	Cytochrome b sequence	Locality	Collection institutional code	GenBank access number
<i>Sus scrofa</i> Uruguayan wild boar	hap1	Velazquez (Rocha)	S2,S3	GU937806
	hap2	A° Malo (Tacuarembó), R.Negro (Durazno), San José	S14, S18, S21	GU937807
	hap3	Velazquez (Rocha Department)	S6	GU937808
	hap4	Velazquez (Rocha Department)	S7	GU937809
	hap5	A° Las Cañas (Durazno)	S17	GU937810
	hap6	Velazquez (Rocha Department)	S9	GU937811
	hap7	Velazquez (Rocha Department)	S10	GU937812
	hap 8	A° Arias (Florida)	S16	GU937813
	hap9	R. Cebollatí (Lavalleja)	S13	GU937814
	hap10	Lazcano (Rocha)	S15	GU937815
	hap11	Velazquez (Rocha Department)	S1,S8,S11	GU937816
	hap12	Velazquez (Rocha Department)	S5	GU937817
	hap13	Sarandí del Yí (Durazno)	S19	GU937818
	hap14	San José	S23	GU937819
Iberian <i>Sus scrofa</i>			IB17 IB19	AY237500 AY237502
<i>Sus scrofa</i> Spanish wild boar			SWB1 SWB4 SWB6 SWB7	AY237510 AY237513 AY237515 AY237516

<i>Sus scrofa</i> Italian wild boar	EWB3		Ssc3	AF136549 AJ314558
<i>Sus scrofa</i> Polish wild boar	EWB2			AF136542
<i>Sus scrofa</i> Bulgarian wild boar			Ssc1	AJ314556
<i>Sus scrofa</i> Israeli wild boar	E1		IWB1	AF163100
<i>Sus scrofa</i> Japanese wild boar	AWB10			AF136550
<i>Sus scrofa</i> Large White	LW2			AF136548
<i>Sus scrofa</i> Landrace			LR1	AY237526
<i>Sus scrofa</i> Duroc				AF136554
<i>Sus scrofa</i> Pietrain			P2	AY237529
<i>Sus verrucosus</i>			Sve1	AJ314552
<i>Phacochoerus africanus</i>			Pafr1	AJ314547