



Research Article
Human and Medical Genetics

Combined expression of *JHDM1D/KDM7A* gene and long non-coding RNA *RP11-363E74* as a biomarker for urothelial cancer prognosis

Glenda Nicioli da Silva¹ , Isadora Oliveira Ansaloni Pereira¹ , Ana Paula Braga Lima¹, Tamires Cunha Almeida², André Luiz Ventura Sávio^{3,4}, Renato Prado Costa⁵, Kátia Ramos Moreira Leite⁶ and Daisy Maria Fávero Salvadori⁷

¹Universidade Federal de Ouro Preto, Escola de Farmácia, Departamento de Análises Clínicas, Ouro Preto, MG, Brazil.

²Instituto Butantan, Laboratório de Dor e Sinalização, São Paulo, SP, Brazil.

³Faculdade Centro Oeste Paulista, Departamento de Odontologia, Piratininga, SP, Brazil.

⁴Universidade do Oeste Paulista, Departamento de Ciências Médicas, Jaú, SP, Brazil.

⁵Hospital Amaral Carvalho, Jaú, SP, Brazil.

⁶Universidade de São Paulo, Faculdade de Medicina, Departamento de Cirurgia, São Paulo, SP, Brazil.

⁷Universidade Estadual Paulista, Faculdade de Medicina, Departamento de Patologia, Botucatu, SP, Brazil.

Abstract

Bladder cancer is the tenth most frequently diagnosed cancer globally. Classification of high- or low-grade tumors is based on cytological differentiation and is an important prognostic factor. LncRNAs regulate gene expression and play critical roles in the occurrence and development of cancer, however, there are few reports on their diagnostic value and co-expression levels with genes, which may be useful as specific biomarkers for prognosis and therapy in bladder cancer. Thus, we performed a marker lesion study to investigate whether gene/lncRNA expression in urothelial carcinoma tissues may be useful in differentiating low-grade and high-grade tumors. RT-qPCR was used to evaluate the expression of the *JHDM1D* gene and the lncRNAs *CTD-2132N18.2*, *SBF2-AS1*, *RP11-977B10.2*, *CTD-2510F5.4*, and *RP11-363E74* in 20 histologically diagnosed high-grade and 10 low-grade tumors. A protein-to-protein interaction network between genes associated with *JHDM1D* gene was constructed using STRING website. The results showed a moderate (positive) correlation between *CTD-2510F5.4* and *CTD2132N18.2*. ROC curve analyses showed that combined *JHDM1D* and *RP11-363E74* predicted tumor grade with an AUC of 0.826, showing excellent accuracy. In conclusion, the results indicated that the combined expression of *JHDM1D* and *RP11-363E74* may be a prognostic biomarker and a promising target for urothelial tumor therapy.

Keywords: Biomarker-oriented therapy, cancer progression, tumor biomarker, gene/lncRNA differential expression, urothelial carcinoma.

Received: September 12, 2023; Accepted: June 25, 2024.

Introduction

Bladder cancer is the tenth most frequently diagnosed cancer globally, and the most common occurring cancer of the urological system. In 2020, approximately 0.573 million new cases and 0.213 million deaths were estimated to occur worldwide due to bladder cancer (Sung *et al.*, 2021). Up to 90–95% of urothelial carcinomas are identified as bladder cancers. At the time of diagnosis, bladder cancers can be classified as non-muscle-invasive (75%) or muscle-invasive (25%). Classification of high- or low-grade tumors is based on cytological atypia and cellular architecture and is an important prognostic factor (Martinez Rodriguez *et al.*, 2017; Sanli *et al.*, 2017; Lenis *et al.*, 2020). Low-grade tumors are usually non-invasive, growing as superficial papillary protrusions restricted to the urothelium and lamina propria, with a high

risk of recurrence (Martinez Rodriguez *et al.*, 2017; Lenis *et al.*, 2020). On the other hand, high-grade tumors may become muscle-invasive and progress to metastatic disease. A small percentage of low-grade tumors (10–15%) can progress to high-grade tumors and become invasive (Sanli *et al.*, 2017).

Despite advancements in technology, cystoscopy, and urine cytology remain the gold standard for diagnosing and monitoring bladder cancer. The use of molecular tests may support the earlier detection of disease, risk stratification of patients, improved prediction of oncological outcomes, and optimization of target therapies. However, international guidelines have not yet incorporated new molecular assessments into daily clinical practice due to difficulty in identifying the appropriate scenario of use, as well as the lack of high-quality prospective trials, resulting in a low level of evidence (Giordano and Soria, 2020). In contrast, significant progress has already been made in terms of biomarker-oriented therapy, and newly identified biomarkers have been demonstrated to be essential in providing clinicians with the information needed to significantly expand their therapeutic arsenal (Jones *et al.*, 2016; Scholtes *et al.*, 2021).

Send correspondence to Glenda Nicioli da Silva. Universidade Federal de Ouro Preto, Escola de Farmácia, Departamento de Análises Clínicas, Campus Morro do Cruzeiro, 35400-000, Ouro Preto, MG, Brazil. E-mail: nicioli@ufop.edu.br

Long noncoding RNAs (lncRNAs), defined as 500+ nucleotides-long non-protein-coding RNAs, have been increasingly shown to regulate gene expression at the epigenetic, transcriptional, and translational levels. Many lncRNAs are often found abnormally expressed in cancer and play critical roles in cancer occurrence and progression, acting as oncogenes as well as tumor suppressors (Mattick *et al.*, 2023). Furthermore, tissue-specific lncRNAs with an ageing-associated expression pattern were observed in human tissues. Thus, lncRNA expression may reflect the tissue-specific fine-tuning of the ageing-associated process. However, no lncRNA has been associated with aging in bladder tissue (Marttila *et al.*, 2020).

lncRNAs *SBF2-AS1* (Fu and Liu, 2021; Zha *et al.*, 2021; Zhang *et al.*, 2021), *RP11-977B10.2* (Liu *et al.*, 2019), *CTD-2510F5.4*, and *RP11-363E7.4* (Wang and Qin, 2019; Chen *et al.*, 2020) are shown to be implicated in carcinogenesis, as well as the histone demethylase *JHDM1D/KDM7A* (Osawa *et al.*, 2011; Meng *et al.*, 2020). However, there are few reports

on their diagnostic value and co-expression levels, which may be useful as biomarkers for prognosis and therapy. Thus, we performed a marker lesion study to investigate whether gene/lncRNA expression in urothelial carcinoma tissues may be useful in differentiating between low- and high-grade tumors.

Subjects and Methods

Patients

Inclusion criteria for patient enrollment in the study were male patients, regardless of age, diagnosed with primary bladder tumors, who attended the University of Sao Paulo Biorepository (São Paulo, Brazil) and Amaral Carvalho Hospital (Jaú, São Paulo, Brazil). A total of 30 fresh bladder cancer tissue samples were collected by consecutive sampling. From those, 20 were histologically diagnosed as high-grade tumors and 10 as low-grade tumors (Table 1). All tumor samples were collected via transurethral resection and were histopathologically classified by a pathologist (K.R.M.L). The

Table 1 – Tumor grade and staging of the tumor tissue samples and patients' ages.

Sample Number	Tumor Grade	Staging	Age
117	High Grade	2	69
113	High Grade	2	77
102	High Grade	2	72
110	High Grade	1	82
130	High Grade	2	46
225	High Grade	2	69
114	High Grade	1	70
104	High Grade	2	63
116	High Grade	1	53
128	High Grade	2	62
125	High Grade	1	70
123	High Grade	1	61
142	High Grade	2	76
169	High Grade	2	56
231	High Grade	2	61
167	High Grade	pTa	73
229	High Grade		54
248	High Grade	3	64
242	High Grade	3	75
163	High Grade	2	68
133	Low Grade	non-invasive	74
124	Low Grade	non-invasive	59
105	Low Grade	non-invasive	70
120	Low Grade	pTa	52
112	Low Grade		67
249	Low Grade	non-invasive	79
247	Low Grade	2a	91
145	Low Grade	pTa	72
151	Low Grade		89
147	Low Grade	pTa	78

pTA: Pappillary non-invasive bladder cancer.

grading and stage were determined according to the World Health Organization (WHO) systems and Tumor-Node-Metastasis (TNM) 2017.

The study was approved by the Ethics Committee of the Sao Paulo State University (protocol 48193715.6.0000.5411), and all methods were performed in accordance with the approved guidelines.

Expression analysis

Tissue biopsies were snap-frozen and stored at -80 °C. Total RNA was isolated using the RNeasy Mini Kit® (Qiagen, Hilden, Germany) according to the manufacturer's protocol. RNA concentration and purity were determined using a NanoDrop spectrophotometer (Thermo Scientific, Waltham, Massachusetts, EUA). RNA quality was analyzed using a 2100 Bioanalyzer (Agilent, Santa Clara, California, USA),

and only samples with an RNA integrity number (RIN) ≥ 6.0 were used. Complementary DNA (cDNA) was synthesized using the High Capacity Kit (Applied Biosystems, Waltham, Massachusetts, USA) with random priming according to the manufacturer's instructions. Expression levels of the *JHDM1D/KDM7A* gene (Pereira *et al.*, 2023) and *CTD-2132N18.2*, *SBF2-AS1*, *RP11-977B10.2*, *CTD-2510F5.4* and *RP11-363E7.4* lncRNAs were analyzed using RT-qPCR. Endogenous reference genes (*HSPCB* and *ACTB*) were selected using the NormFinder software (Andersen *et al.*, 2004).

Each PCR reaction was performed at a final volume of 10 μ L, with the reaction mixture including 0.2 μ g of the cDNA product, 0.75 μ M of each specific primer, Fast Start DNA polymerase, reaction buffer, dNTPs, and SYBR green (Applied Biosystems). The forward and reverse primer sequences are listed in Table 2.

Table 2 – RT-qPCR primers used in this study.

Gene/lncRNA	Primer	Sequence	Reference
<i>CTD-2132N18.2</i>	CTD-2132N18.2 F	5'-GGCGTCAAGGTGGAGTTAGA-3'	Xia <i>et al.</i> , 2021
	CTD-2132N18.2 R	5'-ATCCTCCTTTGCCATGCAGT-3'	
<i>CTD-2510F5.4</i>	CTD-2510F5.4 F	5'-GGTCTCTTGCTCTGTCACCC-3'	Wang and Qin, 2019
	CTD-2510F5.4 R	5'-GCACACCTGTAGTCCCAGTT-3'	
<i>RP11-363E7.4</i>	RP11-363E7.4 F	5'-CGACCACCTATTCCAATT-3'	Wang <i>et al.</i> , 2018
	RP11-363E7.4 R	5'-GCCAGGAAGGCTCAAATC-3'	
<i>RP11-977B10.2</i>	RP11-977B10.2-F	5'-GGTCTTGAGTGGGGCAATCAGC-3'	Liu <i>et al.</i> , 2019
	RP11-977B10.2-R	5'-GAGGTCTTTCAGGAGCCGATG-3'	
<i>HSPCB</i>	HSPCB F	5'-AAGAGAGCAAGGCAAAGTTTGAG-3'	Andersen <i>et al.</i> , 2004
	HSPCB R	5'-TGGTCACAATGCAGCAAGGT-3'	
<i>ACTB</i>	ACTB-S	5'-AGAAGGAGATCACTGCCCTGGCACC-3'	Kondo <i>et al.</i> , 2017
	ACTB-A	5'-CCTGCTTGCTGATCCACATCTGCTG-3'	
<i>JHDM1D</i>	JHDM1D-S	5'-TATTCAAGGGCATGCTGTCTATG-3'	Kondo <i>et al.</i> , 2017
	JHDM1D-AS	5'-GGGATCCTGGAGAGAGTTTCTT-3'	
<i>SBF2-AS1</i>	SBF2AS1S	5'-CACGACCCAGAAGGAGTCTAC-3'	Chen <i>et al.</i> , 2019
	SBF2AS1AS	5'-CCCGTACCTTCTGTGCATA-3'	

The thermal cycling conditions for all genes comprised a temperature profile at 95 °C for 10 min, for initial denaturation, followed by 40 cycles at 95 °C for 15 s and 60 °C for 60 min. RT-qPCR specificity was assessed through melting curves analyses of the amplification products, and the reaction efficiency was estimated using the slope of the standard curve. Standard curves were designated based on the Ct values of cDNA serial dilutions. The relative expression of gene transcripts was calculated using Δ Ct and $2^{(-\Delta\text{Ct})}$ formulas concerning multiple genes (Vandesompele *et al.*, 2002).

“The Atlas of ncRNAs in cancer” (TANRIC – <https://www.tanric.org>) database, which comprises data from expression profiles of lncRNAs and patient survival outcomes, was used to access the survival analysis of groups with high or low expression of the investigated lncRNAs (Li *et al.*, 2015). Survival analyses were accessed by Cox Regression analysis, and the significant differences between the survival curves of the two groups were analyzed by Log-Rank P-value. The R2: Genomics Analysis and Visualization Platform (<https://hgserver1.amc.nl/cgi-bin/r2/main.cgi>), was

used to access the Cox Regression analysis for *JHDM1D/KDM7A* gene.

The STRING 11.5 website (<https://string-db.org/>) and MCL clustering algorithms were used to create a protein-to-protein interaction network (PPI) between genes associated with *JHDM1D* gene.

Statistical analysis

The nonparametric Mann-Whitney test was used for differential gene expression analyses, and the values are expressed as the mean \pm SD. The correlation between the differentiated values was examined using Spearman's rank correlation test (r = correlation coefficient), according to Akoglu (2018). A receiver operating characteristic (ROC) curve was constructed, and the area under the curve (AUC) was calculated to assess the specificity and sensitivity of the predicted gene/lncRNA in differentiating high- and low-grade tumors. Statistical significance was set at $p < 0.05$. Data analysis was performed using GraphPad Prism version 6 and IBM SPSS 17.0.

Results

No significant differential expression of the lncRNAs *CTD-2132N18.2*, *SBF2-AS1*, *RP11-977B10.2*, *CTD-2510F5.4*, and *RP11-363E7.4* was detected between low- and high-grade tumors (Figure 1). Although not statistically significant, lower

expression of *RP11-363E7.4* ($p = 0.1041$) was observed in high-grade tumors compared to low-grade tumors.

As formerly presented by Pereira *et al.* (2023), *JHDMID/KDM7A* gene expression was 2.01 times greater in high-grade tumors than in low-grade tumors. Figure 2 shows the *JHDMID/KDM7A*

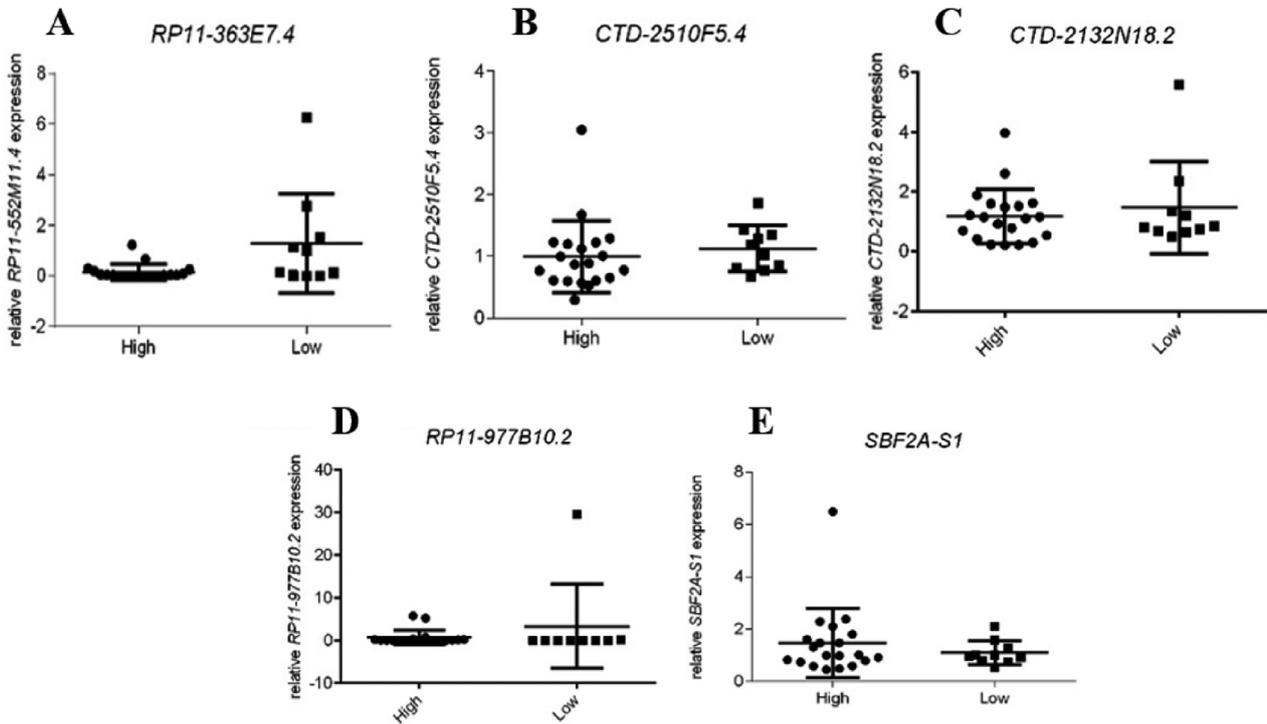


Figure 1 – Relative expression levels of lncRNAs *RP11-363E7.4* (A), *CTD-2510F5.4* (B), *CTD-2132N18.2* (C), *RP11-977B10.2* (D), and *SBF2-AS1* (E) in patients with low- and high-grade bladder tumors. Values are expressed as the mean \pm SD. * $p < 0.05$.

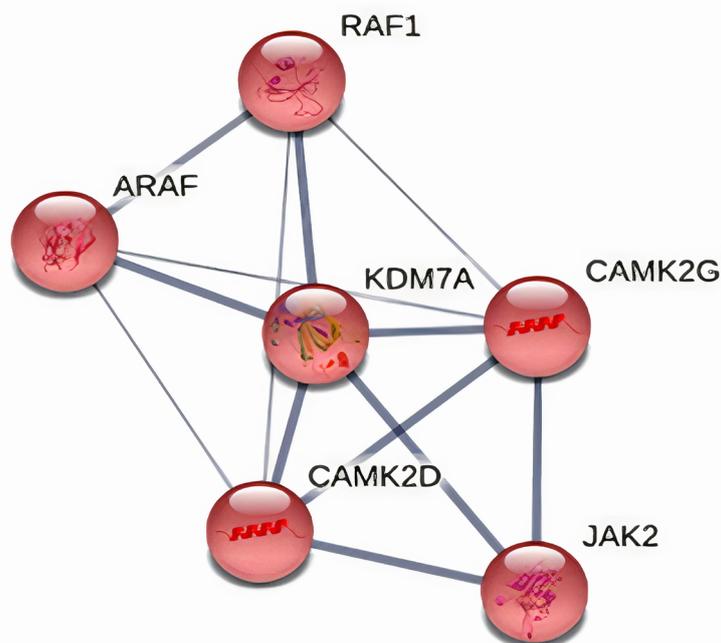


Figure 2 – *JHDMID/KDM7A* gene interaction network, created using String 11.5 software and MCL clustering algorithms. Line thickness indicates data support strength.

KDM7A interaction network. The analysis highlights the relationship between *JHDM1D/KDM7A* and *RAF1*, *JAK2*, *ARAF*, and *CAMK2*.

A correlation analysis was performed to investigate the relationship between *JHDM1D/KDM7A* and the lncRNAs *CTD-2132N18.2*, *RP11-977B10.2*, *CTD-2510F5.4*, *SBF2-AS1*, and *RP11-363E7.4* in both low- and high-grade tumors. This analysis using low- and high-grade samples revealed a moderate (positive) correlation between *CTD-2510F5.4* and *CTD-2132N18.2* expression ($r = 0.6488$, $p < 0.0011$) (Figure 3).

The ability of each isolated lncRNA and the interactions between gene/lncRNA and lncRNA/lncRNA to predict the different tumor subgroups was also tested. Receiver operating characteristics (ROC) curve analyses showed that combined *JHDM1D/KDM7A* gene and *RP11-363E7.4* lncRNA predicted tumor grade with an AUC of 0.826 ($p = 0.004$), showing excellent discrimination capacity (Mandrekar, 2010). Combined expression of *CTD-2510F5.4* and *RP11-363E7.4* also showed an acceptable potential diagnostic value, with an AUC of 0.779 ($p = 0.015$), demonstrating their ability to distinguish between low- and high-grade tumors (Figure 4) (Mandrekar, 2010).

Discussion

Several studies have revealed that lncRNA show highly specific expression patterns in different biological contexts, and their abnormal expression is associated with the progression and prognosis of human malignancies (Fu and Liu, 2021; Gao *et al.*, 2021; Pereira *et al.*, 2023). Thus, a better understanding of the interaction between genes and lncRNAs may be useful in cancer diagnosis, prognosis, and treatment. Based on this rationale, we conducted a marker lesion study using gene/lncRNA expression to distinguish, with high specificity, between low- and high-grade bladder tumors.

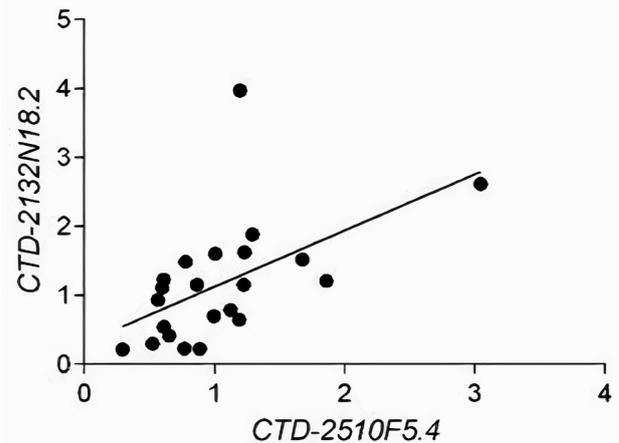


Figure 3 – Correlation analysis showing a moderate (positive) correlation between the lncRNAs *CTD-2132N18.2* and *CTD-2510F5.4* expression in both low- and high-grade tumor samples. Spearman correlation analysis. $r = 0.6488$, $p < 0.0011$.

The histone demethylase *JHDM1D* gene (also known as *KDM7A*) is a member of the plant homeodomain (PHD) finger protein (PHF) family of PHD and JmjC domain-containing histone demethylases and participates in epigenetic regulation (Klose *et al.*, 2006). This gene also regulates many biological processes, including differentiation, development, and growth of several cancer cells (Yang *et al.*, 2019; Meng *et al.*, 2020). In bladder cancer cell lines, *JHDM1D* knockdown led to impaired cell growth, increased cell death, and reduced rates of cell migration (Lee *et al.*, 2018). Indeed, a study from Pereira *et al.* (2023) showed different expression levels of *JHDM1D* in low- and high-grade tumors, suggesting a possible role of this gene in bladder tumor progression. In addition, we found

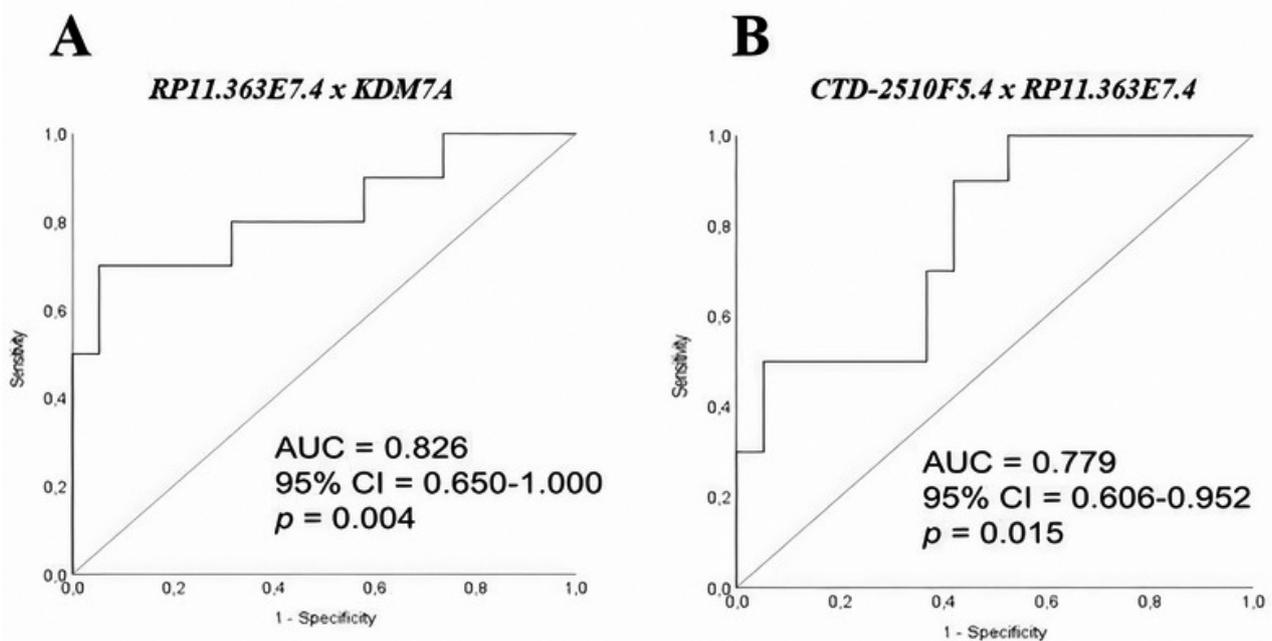


Figure 4 – Receiver operating characteristics (ROC) curves using the combined expression of gene/lncRNA *KDM7A/ RP11-363E7.4* (A) and lncRNAs *CTD-2510F5.4/ RP11-363E7.4* (B) to assess accuracy to distinguish between low- and high-grade tumors.

that the interaction network involving *JHDMID/KDM7A* was associated with *RAF1*, *ARAF*, *JAK2*, and *CAMK2* activation, reinforcing the involvement of *JHDMID/KDM7A* in tumor aggressiveness and progression. As per the literature, these four genes also play a role in carcinogenesis. *RAF* kinases normally function as activators of the mitogen-activated protein kinase (MAPK) signaling pathway, which indirectly regulates cell proliferation and survival (Montagut and Settleman, 2009). *ARAF*, also required for MAPK activation in a variety of cancer types (e.g., colorectal, pancreatic, and breast cancers), is associated with the migration and invasiveness of tumor cells (Mooz *et al.*, 2014). The exact role of *JAK2* signaling in solid cancers is unclear, but *JAK2* inhibition may prevent disease progression through restriction of malignant cell phenotypes (Harry *et al.*, 2012). The emerging role of the *CAMK2* gene in the regulation of cancer progression, especially proliferation, cell cycle, and metastasis, and in therapy response has also been reported (Wang *et al.*, 2015) Therefore, *JHDMID* gene expression levels were chosen to be correlated with lncRNAs expression to access our prognostic marker lesion study.

Combined gene/lncRNA or lncRNA/lncRNA expression can be used to achieve more accurate diagnosis and prognosis when compared to the analysis of the gene/lncRNA alone. Indeed, Pereira *et al.* (2023) found a moderate positive correlation between *JHDMID* gene and lncRNA *JHDMID-ASI* (an antisense transcript from *JHDMID*) expression in high-grade tumors. In addition, the combination of *JHDMID* and *JHDMID-ASI* showed potential prognostic value in distinguishing between low- and high-grade bladder tumors. In our study, no correlation was found between *JHDMID* gene and the analyzed lncRNAs. Nevertheless, although a positive correlation between the lncRNAs *CTD-2132N18.2* and *CTD-2510F5.4* was observed, no diagnostic predictive value was detected. Similarly, *SBF2-ASI* and *RP11-977B10.2* levels also failed to differentiate between low- and high-grade tumors. Therefore, the lncRNAs *CTD-2132N18.2*, *CTD-2510F5.4*, *SBF2-ASI*, and *RP11-977B10.2* were not associated with bladder tumor progression.

The best potential diagnostic values of combined detection occurred with lncRNA *RP11-363E7.4*, a recently discovered novel lncRNA. *JHDMID/RP11-363E7.4* combination appeared to have excellent diagnostic value, as predicted by the ROC curve. Furthermore, the combination of lncRNAs *CTD-2510F5.4* and *RP11-363E7.4* also showed an acceptable potential diagnostic value. Despite the lack of statistical significance ($p = 0.1041$), decreased expression of *RP11-363E7.4* in high-grade tumors compared to low-grade tumors may have some clinical significance. This finding suggests that downregulation of this lncRNA is associated with tumor aggressiveness. Similarly, *RP11-363E7.4* downregulation was observed in gastric cancer, with its higher expression correlated with better overall survival in cancer patients (Wang *et al.*, 2018). Although the functional role and molecular mechanisms are still unclear, a recent study by (Chen *et al.*, 2020) showed that *RP11-363E7.4* can function as a tumor suppressor by inhibiting proliferation, migration, and invasion, and inducing apoptosis.

The prognostic biomarkers proposed aimed at predicting the tumor grade, however, the lncRNA and/or gene expression levels may also be valuable when evaluating clinical patient outcomes. Although our study has not revealed a significant difference in the expression of the mentioned lncRNAs between high and low-grade bladder tumors, the Log-Rank Test applied to Cox regression analysis showed that higher expression of lncRNAs *CTD-2132N18.2* (Log-Rank P-value = 0.04908) and *RP11-977B10.2* (Log-Rank P-value = 0.0060563) were associated with lower survival probability when compared to the group with lower expression of these lncRNAs. No significant differences in the outcome were found between groups with lower and higher expression of lncRNAs *SBF2-ASI* and *RP11-363E7.4*, and no data about lncRNA *CTD-2510F5.4* was found in TANRIC database (Li *et al.*, 2015). Moreover, a significantly lower survival probability of groups with high expression of *JHDMID/KDM7A* gene, which showed potential prognostic value in predicting tumor grade in our study, was also observed (Cox p-value = 0.00217).

It is important to highlight that the number of tissue samples analyzed may have been a limitation of the current study. A greater number of bladder tumor specimens may be able to more clearly demonstrate the link between certain gene/lncRNA combinations and tumor progression. lncRNA *RP11-363E7.4* and *JHDMID* silencing in low- and high-grade cell lines, and subsequently resulting changes in biological behavior should also be considered.

In conclusion, this study revealed that the combined expression of *JHDMID/RP11-363E7.4* may predict tumor progression, and this combination may serve as an attractive prognostic biomarker and a promising target for urothelial carcinoma treatment.

Acknowledgements

This study was supported by the National Council for Scientific and Technological Development (CNPq) [grant numbers 303435/2016-0, 305277/2023-5 and 406334/2018-8]. GNS, KRML, and DMFS thank CNPq for their fellowships.

Conflict of Interest

All the authors declare there are no conflicts of interest.

Author Contributions

GNS, ALVS, RPC, KRML and DMFS conceived and designed the study; APBL and GNS conducted the experiments; TCA and IOAP analyzed the data; DMFS and GNS wrote the manuscript. All authors read and approved the final version.

References

- Akoglu H. User's guide to correlation coefficients (2018) Turkish J Emerg Med 18:91–93.
- Andersen CL, Jensen JL and Ørntoft TF (2004) Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance estimation approach to identify genes suited for normalization, applied to bladder and colon cancer data sets. Cancer Res 64:5245–5250.

- Chen C, Wang X, Liu T, Tang X, Liu Y, Liu T and Zhu J (2020) Overexpression of long non-coding RNA RP11-363E7.4 inhibits proliferation and invasion in gastric cancer. *Cell Biochem Funct* 38:921–931.
- Chen R, Xia W, Wang S, Xu Y, Ma Z, Xu W, Zhang E, Wang J, Fang T, Zhang Q *et al* (2019) Long Noncoding RNA SBF2-AS1 is critical for tumorigenesis of early-stage lung adenocarcinoma. *Mol Ther Nucleic Acids* 16:543–553.
- Fu DW and Liu AC (2021) LncRNA SBF2-AS1 Promotes diffuse large B-Cell lymphoma growth by regulating FGFR2 via Sponging miR-494-3p. *Cancer Manag Res* 13:571–578.
- Gao X, Liu Q, Chen X, Chen S, Yang J, Liu Q and Cheng Y (2021) Screening of tumor grade-related mRNAs and lncRNAs for esophagus squamous cell carcinoma. *J Clin Lab Anal* 35:e23797.
- Giordano A and Soria F (2020) Role and efficacy of current biomarkers in bladder cancer. *AME Med J* 5:6.
- Harry BL, Eckhardt SG and Jimeno A (2012) JAK2 inhibition for the treatment of hematologic and solid malignancies. *Expert Opin Investig Drugs* 21:637–655
- Jones RT, Felsenstein KM and Theodorescu D (2016) Pharmacogenomics: Biomarker-directed therapy for bladder cancer. *Urol Clin North Am* 43:77–86.
- Klose RJ, Kallin EM and Zhang Y (2006) JmjC-domain-containing proteins and histone demethylation. *Nat Rev Genet* 7:715–727.
- Kondo A, Nonaka A, Shimamura T, Yamamoto S, Yoshida T, Kodama T, Aburatani H and Osawa T (2017) Long Noncoding RNA JHDM1D-AS1 promotes tumor growth by regulating angiogenesis in response to nutrient starvation. *Mol Cell Biol* 37:e00125-17.
- Lee K-H, Hong S, Kang M, Jeong CW, Ku JH, Kim HH and Kwak C (2018) Histone demethylase KDM7A controls androgen receptor activity and tumor growth in prostate cancer. *Int J Cancer* 143:2849–2861.
- Lenis AT, Lec PM and Chamie K (2020) Bladder cancer: A review. *JAMA* 324:1980–1991.
- Li J, Han L, Roebuck P, Diao L, Liu L, Yuan Y, Weinstein JN and Liang H (2015) TANRIC: An interactive open platform to explore the function of lncRNAs in cancer. *Cancer Res* 75:3728–3737.
- Liu H, Sun Y, Tian H, Xiao X, Zhang J, Wang Y and Yu F (2019) Characterization of long non-coding RNA and messenger RNA profiles in laryngeal cancer by weighted gene co-expression network analysis. *Aging* 18:10074–10099.
- Mandrekar JN (2010) Receiver operating characteristic curve in diagnostic test assessment. *J Thorac Oncol* 5:1315–1316.
- Marttila S, Chatsirisupachai K, Palmer D and Magalhães JP (2020) Ageing-associated changes in the expression of lncRNAs in human tissues reflect a transcriptional modulation in ageing pathways. *Mech Ageing Dev* 185:111177.
- Martinez Rodriguez RH, Buisan Rueda O and Ibarz L (2017) Bladder cancer: Present and future. *Med Clin* 149:449–455.
- Mattick JS, Amaral PP, Carninci P, Carpenter S, Chang HY, Chen L-L, Chen R, Dean C, Dinger ME, Fitzgerald KA *et al* (2023) Long non-coding RNAs: Definitions, functions, challenges and recommendations. *Nat Rev Mol Cell Biol* 24:430–447.
- Meng Z, Liu Y, Wang J, Fan H, Fang H, Li S, Yuan L, Liu C, Peng Y, Zhao W *et al* (2020) Histone demethylase KDM7A is required for stem cell maintenance and apoptosis inhibition in breast cancer. *J Cell Physiol* 235:932–943.
- Montagut C and Settleman J (2009) Targeting the RAF–MEK–ERK pathway in cancer therapy. *Cancer Lett* 283:125–134.
- Mooz J, Oberoi-Khanuja TK, Harms GS, Wang W, Jaiswal BS, Seshagiri S, Tikkanen R and Rajalingam K (2014) Dimerization of the kinase ARAF promotes MAPK pathway activation and cell migration. *Sci Signal* 7:ra73.
- Osawa T, Muramatsu M, Wang F, Tsuchida R, Kodama T, Minami T, Kodama T, Minami T and Shibuya M (2011) Increased expression of histone demethylase JHDM1D under nutrient starvation suppresses tumor growth via down-regulating angiogenesis. *Proc Natl Acad Sci U S A* 108:20725–20729.
- Pereira IOA, da Silva GN, Almeida TC, Lima APB, Sávio ALV, Leite KRM and Salvadori DMF (2023) LncRNA JHDM1D-AS1 is a key biomarker for progression and modulation of gemcitabine sensitivity in bladder cancer cells. *Molecules* 28:2412.
- Sanli O, Dobruch J, Knowles MA, Burger M, Alemozaffar M, Nielsen ME and Lotan Y (2017) Bladder cancer. *Nat Rev Dis Prim* 13:17022.
- Scholtes MP, Alberts AR, Iffé IG, Verhagen PCMS, Van der Veldt AAM and Zuiverloon TCM (2021) Biomarker-Oriented therapy in bladder and renal cancer. *Int J Mol Sci* 22:2832.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A and Bray F (2021) Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 71:209–249
- Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paep A and Speleman F (2002) Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biol* 3:research0034.1.
- Wang P, Li J, Zhao W, Shang C, Jiang X, Wang Y, Zhou B, Bao F and Qiao H (2018) A Novel LncRNA-miRNA-mRNA triple network identifies LncRNA RP11-363E7.4 as an important regulator of miRNA and gene expression in gastric cancer. *Cell Physiol Biochem* 47:1025–1041.
- Wang Y, Zhao R and Zhe H (2015) The emerging role of CaMKII in cancer. *Oncotarget* 6:11725–11734.
- Wang Z and Qin B (2019) Prognostic and clinicopathological significance of long noncoding RNA CTD-2510F5.4 in gastric cancer. *Gastric Cancer* 22:692–704.
- Xia L, Chen X, Yang J, Zhu S, Zhang L, Yin Q, Hong Y, Chen H, Chen G and Li H (2021) Long Non-Coding RNA-PAICC promotes the tumorigenesis of human intrahepatic cholangiocarcinoma by increasing YAP1 transcription. *Front Oncol* 10:595533.
- Yang X, Wang G, Wang Y, Zhou J, Yuan H, Li X, Liu Y and Wang B (2019) Histone demethylase KDM7A reciprocally regulates adipogenic and osteogenic differentiation via regulation of C/EBP α and canonical Wnt signalling. *J Cell Mol Med* 23:2149–2162.
- Zha W, Li X, Tie X, Xing Y, Li H, Gao F, Ye T, Du W, Chen R and Liu Y (2021) The molecular mechanisms of the long noncoding RNA SBF2-AS1 in regulating the proliferation of oesophageal squamous cell carcinoma. *Sci Rep* 11:805.
- Zhang Q, Liu X-J, Li Y, Ying X-W and Chen L (2021) Prognostic Value of Immune-Related lncRNA SBF2-AS1 in Diffuse Lower-Grade Glioma. *Technol Cancer Res Treat* 20:15330338211011966.

Internet Resources

- STRING 11.5 Software, <https://string-db.org/> (accessed 01 February 2022).
- TANRIC – The Atlas of ncRNAs in cancer – database, <https://www.tanric.org> (accessed 14 March 2024).
- The R2: Genomics Analysis and Visualization Platform, <https://hgserver1.amc.nl/cgi-bin/r2/main.cgi> (accessed 14 March 2024).

Associate Editor: Carlos R. Machado

License information: This is an open-access article distributed under the terms of the Creative Commons Attribution License (type CC-BY), which permits unrestricted use, distribution and reproduction in any medium, provided the original article is properly cited.