



The post-transcriptional gene silencing pathway in *Eucalyptus*

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Abstract

Post-transcriptional gene silencing (PTGS) is a conserved surveillance mechanism that identifies and cleaves double-stranded RNA molecules and their cellular cognate transcripts. The RNA silencing response is actually used as a powerful technique (named RNA interference) for potent and specific inhibition of gene expression in several organisms. To identify gene products in *Eucalyptus* sharing similarities with enzymes involved in the PTGS pathway, we queried the expressed sequence tag database of the Brazilian *Eucalyptus* Genome Sequence Project Consortium (FORESTs) with the amino acid sequences of known PTGS-related proteins. Among twenty-six prospected genes, our search detected fifteen assembled sequences encoding products presenting high level of similarity (E value < 10⁻⁴⁰) to proteins involved in PTGS in plants and other organisms. We conclude that most of the genes known to be involved in the PTGS pathway are represented in the FORESTs database.

Key words: gene silencing, PTGS, RNA silencing, EST, *Eucalyptus*.

Received: May 31, 2004; Accepted: March 17, 2005.

Introduction

Post-transcriptional gene silencing (PTGS) is a widely conserved surveillance system that acts at the transcriptome level. It identifies long double stranded RNA molecules (dsRNA) such as those generated during virus replication, transposons mobilization or aberrant RNA synthesis. These molecules act as trigger signals for the PTGS machinery, guiding the cell to an alert state.

Once identified, trigger dsRNAs are cleaved by a cellular dsRNA-specific endonuclease named Dicer (DCL for Dicer-like in plants; Bernstein *et al.*, 2001; Zamore *et al.*, 2000; Schauer *et al.*, 2002) into ~21-23 nucleotide duplexes known as small interfering RNAs (siRNAs; Hamilton and Baulcombe, 1999). Somehow, a Dicer-associated protein (R2D2) "senses" thermodynamic free energy of siRNA 5' ends, binding to its more stable one (Tomari, *et al.*, 2004). This process coordinates Dicer (re-) positioning over siRNA and duplex winding by an RNA helicase (SDE3 in plants; Dalmay *et al.*, 2001) through its more unstable termini, allowing the loading of one strand into a multi-component complex termed RISC (for RNA induced silencing complex; Hammond *et al.*, 2000).

RISC is composed of several proteins of unknown function (Gemin 3 and 4, VIG and dFXR; Hutvagner and Zamore, 2002; Caudy *et al.*, 2002) and by multiple nucleases such as Slicer (or Argonaute 2), the catalytic endonuclease responsible for mRNA target cleavage (Parker *et al.*, 2004; Liu, *et al.*, 2004; Song, *et al.*, 2004; Meister, *et al.*, 2004; Rand, *et al.*, 2004), Tudor-SN, a micrococcal nuclease homologue (Caudy *et al.*, 2003) and probably Wex, a 3'-5' RNase D-like protein (Glazov *et al.*, 2003). Loaded siRNA guides RISC to the target RNA promoting its endonucleolytic cleavage and thus allowing 3'-5' (Wex) and 5'-3' (AtXRN4; Souret *et al.*, 2004) exoribonucleases to act on the generated unprotected ends (Figure 1).

In plants and worms, the system is self-sustained and amplified by the activity of a siRNA-primed RNA-dependent RNA polymerase (RdRP or RDR; Mourrain *et al.*, 2000; Dalmay *et al.*, 2000). In certain cases, such as in silencing induced by dsRNA-replicating viruses, the requirement of an RdRP activity is bypassed (Dalmay *et al.*, 2000). In *Caenorhabditis elegans*, local introduction of dsRNA leads to systemic silencing that is probably mediated by a transmembrane protein named SID-1 (Winston *et al.*, 2002). The activity of a protein of unknown function with coiled-coil domains (SGS3) is also required for gene silencing (Mourrain *et al.*, 2000).

PTGS can also affect the genome promoting alterations such as DNA methylation (through MET1; Finnegan

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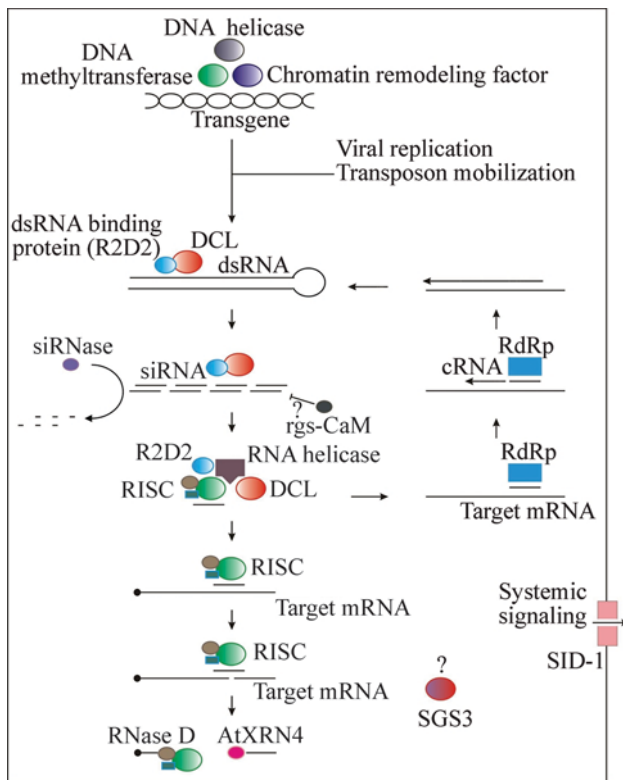


Figure 1 - Schematic representation of the molecular steps in PTGS pathway. Nuclear events at the loci of the target gene or transgene are promoted by MET1, DDM1 and QDE3 (a RecQ DNA helicase). PTGS is triggered by long dsRNA molecules that are converted by the ribonuclease III Dicer (or Dicer-like enzymes) into small interfering dsRNAs of 21-23 nucleotides (siRNA). One strand of siRNA is transferred from DCL to RISC, thus directing target RNA cleavage. Unprotected generated ends are targets for exoribonucleases (Rnase D and AtXRN4). A siRNA-primed RNA-dependent RNA polymerase (RdRp) synthesizes new dsRNA molecules from target RNA thus perpetuating the process. Sid-1 is a transmembrane protein required for systemic signaling in *C. elegans*.

et al., 1996) and chromatin remodeling (through DDM1; Jeddeloh *et al.*, 1999) at loci homologous to the target RNA (Morel *et al.*, 2000).

PTGS phenomenon, also called RNA silencing, is a potent means to counteract foreign sequences and its natural role in host defenses against transposable elements and viruses has been demonstrated. Consistent with this defense role, a great number of viral suppressors of RNA silencing have been identified so far (Voinnet *et al.*, 1999). A gene encoding an endogenous RNA silencing suppressor was discovered in tobacco (named *rgs-CaM*; Anandalakshmi *et al.*, 2000) and was proposed to be involved in endogenous gene regulation.

RNA silencing has been observed in plants, protozoans, insects, fungi (named quelling) and mammals (named RNA interference; RNAi), indicating common aspects and functional analogy among pathways. RNAi is actually used as an experimental approach to suppress the expression of specific transcripts, thus becoming a powerful tool to dissect gene function and a promising therapeutic technology.

In this paper we provide an inventory of the putative genes involved in the PTGS pathway of *Eucalyptus* by exploring the expressed sequence tag (EST) information generated in the Brazilian *Eucalyptus* Genome Sequence Project Consortium (FORESTs). Enhanced understanding of this pathway will contribute to the establishment of RNAi-based protocols intending functional genomic studies in this important tree.

Materials and Methods

To identify gene products sharing similarities with enzymes involved in the PTGS pathway (Figure 1), similarity searches against cluster consensus sequences (contigs)

Table 1 - PTGS-related proteins and homologues identified in *Eucalyptus*.

Number	Name	Organism	Function	Access	Clusters	e-value	Identity	Coverage
1	MET1	<i>Zea mays</i>	DNA methyltransferase	AAG15406	EGSBSL4219H01.g	2e-11	55%	8%
2	DDM1	<i>Arabidopsis thaliana</i>	Chromatin remodeling factor	At5g66750	EGMCST7267G07.g	e-150	72%	44%
3	QDE-3	<i>Neurospora crassa</i>	RecQ DNA Helicase	AAF31695	EGCCSL4031H03.g	2e-54	50%	11%
4	Argonaute family	AGO.1	Involved in siRNA and miRNA pathways	At1g48410	EGEQST6003A06.g	0.0	86%	43%
		EGEQRT3002E10.g			0.0	76%	78%	
5		AGO.2	RISC component	NP_730054	EGEQST2200C06.g	0.0	61%	50%
					EGEQSL5001E09.g	4e-89	63%	25%
					EGJEST2008F12.g	8e-53	44%	23%
6	AGO.4	<i>Arabidopsis thaliana</i>	Control of siRNA accumulation and DNA methylation	At2g27040	EGBMRT6278A01.g	e-130	67%	35%
					EGJFFB1207F01.g	3e-70	83%	16%
					EGMCRT6006C01.g	e-153	73%	38%
					EGEQFB1034D02.g	1e-75		23%
7	SGS3	<i>Arabidopsis thaliana</i>	Unknown	At5g23570	EGQHST6221A03.g	1e-49	41%	46%
					EGJFLV2207H03.g	3e-42	41%	33%
8	SDE3	<i>Arabidopsis thaliana</i>		At1g05460	EGEZWD2227F09.g	2e-77	58%	23%
	MUT6	<i>Chlamydomonas reinhardtii</i>	RNA helicase	AF305070	EGJFRT3163C04.g	1e-86	71%	16%
					EGACLV3290A02.g	2e-64	61%	10%

Table 1 (cont).

Number	Name	Organism	Function	Access	Clusters	e-value	Identity	Coverage
9	DCL1			At1g01040	EGEQWD2256B05.g	e-139	72%	18%
10	DCL2	<i>Arabidopsis thaliana</i>	dsRNA-specific endonuclease (RNase III)	At3g03300	EGEZWD2267C04.g	2e-71	56%	18%
11	DCL3			At3g43920	EGEPST6161G12.g	e-139	60%	27%
					EGACSL5244C02.g	2e-50	55%	12%
12	DCL4			At5g20320	EGSBCL1312G08.g	2e-63	57%	13%
13	rgs-CaM	<i>Nicotiana tabacum</i>	Suppressor of PTGS	AAK11255	EGEZRT5203G11.g	2e-41	62%	72%
14	RDR6	<i>Arabidopsis thaliana</i>	RNA-dependent RNA polymerase	At3g49500	EGRFST6265B04.g	2e-92	61%	22%
					EGEZST2003F10.g	2e-77	37%	39%
					EGQHCL1342B05.g	3e-56	54%	18%
15	AtXRN4	<i>Arabidopsis thaliana</i>	5'-3' exoribonuclease	At1g54490	EGBGRT6263B05.g	1e-80	60%	26%
					EGEZCL1262C10.g	2e-72		
					EGBGWD2290C10.g	7e-42		

in the FORESTs database (<https://forests.esalq.usp.br>) were performed using BLAST search programs (Altschul *et al.*, 1990). The database was first prospected using query sequences from known PTGS-related proteins (Table 1) and the TBLASTN algorithm. Positive hits (e-values $< 10^{-40}$) were validated against existing homologous sequences in the GenBank nonredundant protein database by using BLASTX. To further validate annotation, a protein domain analysis was performed using the Reverse Position Specific-BLAST (RPS-BLAST) algorithm at www.ncbi.nlm.nih.gov/structure/cdd/wrpsb.cgi, Pfam (Bateman *et al.*, 2000) and SMART (Schultz *et al.*, 2000) databases. An estimate of the relative abundance of the identified putative genes was generated based on EST counts per corresponding contig.

Results and Discussion

The FORESTs database was mined for *Eucalyptus* gene products potentially involved in the PTGS pathway (Figure 1). We have searched for genes encoding 26 PTGS-related proteins and have found matches for 15 of them (Table 1). Among the identified EST clusters, the one annotated as MET1 homologue exhibited a low e-value ($2e-11$) but contained a highly conserved methyltransferase domain, indicating that it probably encodes a DNA methyltransferase. Searches also revealed several clusters with significant sequence and domain similarities to the four *Arabidopsis* homologues of Dicer (DCL1 to 4) and four clusters encoding RDR6 (also known as SDE1/SGS2).

Using the amino acid sequences of AGO-1, AGO-2 and AGO-4 as queries, nine clusters encoding Argonaute proteins were identified. AGO proteins are a common component of all RISC-related complexes and are defined by the presence of two conserved domains: a ~20 kDa N-terminal PAZ domain and a ~40 kDa C-terminal PIWI domain, which is in fact a (cryptic) RNase H domain responsible for RNA target cleavage (Song *et al.*, 2004). At least ten AGO homologues have been identified in the *Arabidopsis*

genome. Among them, AGO-1 is involved in the siRNA and microRNA (miRNA) pathways (Vaucheret *et al.*, 2004) while AGO-4 plays a role in locus-specific siRNA accumulation and RNA-directed DNA methylation (Zilberman, *et al.*, 2003; Zilberman, *et al.*, 2004). In *Drosophila*, *Homo sapiens* and *Mus musculus*, AGO-2 mediates target mRNA cleavage (Okamura *et al.*, 2004, Meister, *et al.*, 2004; Liu, *et al.*, 2004). Since these proteins are closely related to each other, and biochemical characterization of AGO-2 in plants is still missing, the correct assignment of the annotated *Eucalyptus* clusters proved to be difficult without further analyses.

We were also able to assign three clusters to AtXRN4, two clusters to SGS3, one cluster to SDE3 and a cluster to rgs-CaM ($2e-41$), a calmodulin-related protein that acts as a cellular suppressor of PTGS (Anandalakshmi *et al.*, 2000).

In contrast, BLAST searches revealed no gene product with significant similarity to RDE-4 (R2D2 in *Drosophila*), a dsRNA binding protein that interacts with RNA molecules identical to the trigger dsRNA during RNAi in *C. elegans* (Tabara *et al.*, 2002). Likewise no HEN1-related clusters could be found in the FORESTs database. HEN1 is a protein of 942 amino acids playing a role

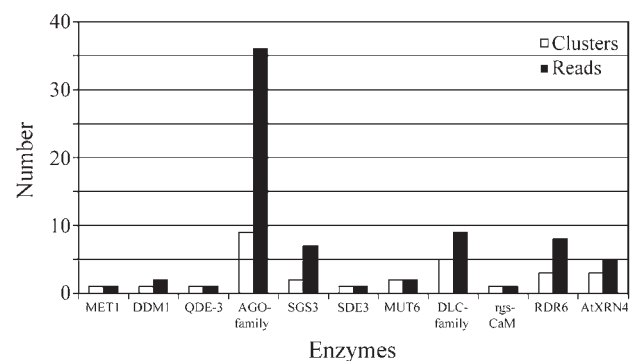


Figure 2 - Cluster distribution and total number of ESTs (in each cluster) encoding enzymes involved in the PTGS pathway.

in siRNA and miRNA accumulation in plants (Boutet *et al.*, 2003). We do believe, however, that this protein is present in *Eucalyptus* but is not represented in the FORESTs database. Moreover, we did not find any EST clusters similar to proteins that have been genetically or biochemically linked to the PTGS pathway in humans (Gemin 3 and 4 of unknown function), *Drosophila* (RNA-binding proteins VIG and dFXR associated to RISC) and *C. elegans* (MUT-7 RNase D also known as WEX in *Arabidopsis*; ERI-1, a siRNA-degrading RNase; Kennedy *et al.*, 2004).

An estimate of the relative abundance of the identified genes was obtained by comparing the number of times ESTs were assigned to a particular contig. An overall view of the obtained results shows that the enzymes involved in PTGS are poorly represented in the FORESTs database (Figure 2). In general, transcripts encoding proteins of the AGO family, DCL family and RDR6 figured as the most extensively represented.

In summary, several EST clusters could be assigned to proteins of known functions in PTGS in plants and other organisms, covering almost the entire pathway (Figure 1 and Table 1). The identification of components of the PTGS pathway within the generated set of *Eucalyptus* ESTs provides good evidence for the conservation of the RNA silencing mechanism among different plant species and organisms. Recent findings suggest that RNA silencing and related pathways are involved in several cellular processes such as defense, RNA surveillance and development. Elucidation of the molecular mechanisms underlying these processes is an important step towards the full understanding of the PTGS phenomenon. From our analyses, we can conclude that *Eucalyptus* encodes a functional RNA silencing pathway. This collection of PTGS-related ESTs provides an interesting resource for molecular and functional genomic studies in this important tree.

Acknowledgements

The authors are grateful to the *Eucalyptus* Consortium for the computer facilities and to FAPESP for financial support. T.C-P. is supported by FAPESP and I.G.M. is a recipient of a research fellowship from CNPq.

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Associate Editor: Carlos F. M. Menck