

Optimization of a New Dissolution Test for Oxcarbazepine Capsules using Mixed-Level Factorial Design

Hudson C. Polonini,^a Marcone A. L. de Oliveira,^b Anderson O. Ferreira,^c
Nádia R. B. Raposo,^a Livia N. Grossi^c and Marcos A. F. Brandão*^a

^aFaculty of Pharmacy and ^bDepartment of Chemistry, Federal University of Juiz de Fora, 36036-900 Juiz de Fora-MG, Brazil

^cOrtofarma – Controle de Qualidade, 36120-000 Matias Barbosa-MG, Brazil

Neste trabalho foi desenvolvido e validado um teste de dissolução para cápsulas de oxcarbazepina. Para o estudo de triagem, foi utilizado um planejamento experimental de níveis mistos composto de fatores com dois e três níveis ($3^2 \times 2$), de modo a selecionar a velocidade de agitação, o meio de dissolução e o aparato. Esta estratégia é útil, pois reduz o tempo de desenvolvimento do método e também gera resultados mais exatos. A melhor porcentagem de dissolução foi obtida utilizando o aparato pá a 80 rpm e com solução aquosa de laurilsulfato de sódio a 1% m/v como meio de dissolução. A quantificação da massa dissolvida foi realizada por análise espectrofotométrica UV-Vis a 304 nm. Os resultados do estudo da validação demonstraram que o método é preciso e linear no intervalo de 83-249 $\mu\text{g mL}^{-1}$ de oxcarbazepina. Os resultados evidenciaram que o método apresenta utilidade e adequabilidade para o estudo da dissolução de cápsulas de oxcarbazepina, uma vez que não existe método oficial com este propósito.

In this work, a dissolution test for oxcarbazepine capsules was developed and validated. For the screening study, a mixed-level factorial design containing factors at three and two levels ($3^2 \times 2$) was used in order to select the stirring speed, the dissolution medium and the dissolution apparatus. This strategy is needful for reducing the time of method development and to provide less ambiguous data. The best *in vitro* percentage dissolution was obtained using apparatus paddle at 80 rpm and sodium lauryl sulfate 1% m/v aqueous solution as the dissolution medium. The quantification of the mass dissolved was obtained by UV-Vis spectrophotometric analysis at 304 nm. The validation results demonstrate that the method was precise and linear over the range of 83-249 $\mu\text{g mL}^{-1}$ of oxcarbazepine. Thus, the method is useful and adequate for oxcarbazepine capsules dissolution test, once there is no official monograph regarding it.

Keywords: oxcarbazepine, dissolution test, mixed-level factorial design

Introduction

Oxcarbazepine (OCBZ) (Figure 1) is a keto analog of carbamazepine¹ indicated as monotherapy or adjunctive therapy for the treatment of partial seizures with or without secondarily generalized tonic-clonic seizures in adults and children aged over six years.^{2,3} The advantages of OCBZ on carbamazepine are: lower frequency and severity of adverse events (particularly endocrine),⁴⁻⁶ better tolerability,^{3,6} fewer drug interactions⁷ and almost complete absorption after oral ingestion.³

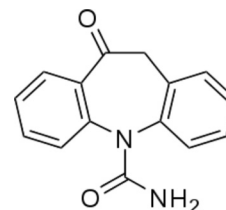


Figure 1. Chemical structure of oxcarbazepine (OCBZ, $\text{C}_{15}\text{H}_{12}\text{N}_2\text{O}_2$).

Another point is that OCBZ is a low therapeutic index, high dosage and low potency drug. Its compounding and marketing, therefore, can only be made if the performance of the formulation is proved by conducting the dissolution test of the final product.⁸

*e-mail: marcosbrand@uol.com.br

Dissolution tests have become a highly valuable tool for quality analysis in the pharmaceutical industry in what concerns the performance of the oral solid dosage form/delivery system,^{9,10} because its results can be directly correlated to and reflect the bioavailability and bioequivalence of an active substance.¹¹ A regulatory approval is a requirement for the product to be marketed, and so the performance of oral solid dosage forms such as soft-gel capsules must be verified through an *in vitro* dissolution test.^{12,13}

This *in vitro* dissolution test is so relevant for the pharmaceutical industries because it can predict the *in vivo* performance of a drug product, once the absorption of an oral drug is mainly dependent on the release of the active substance from its dosage form and also its dissolution and/or solubilization under physiological conditions.¹¹ The capability of releasing the drug substance in aqueous media, making it available for gastrointestinal absorption, is indispensable for quality of the oral dosage forms.¹⁰ Other important applications of the dissolution tests include the assessing of the batch-to-batch quality, the guidance for the development of new formulations and the assurance that changes in the original formulation do not affect the final product.¹¹

After the dissolution testing, the amount of the substance dissolved should be determined by an adequate analytical procedure. This method has to be sufficiently sensitive, selective in presence of excipients, robust and rapid because the dissolution experiments generate a large number of samples.¹³

Many of the official pharmaceutical compendiums such as American,¹⁰ European,¹⁴ British,¹⁵ Japanese,¹⁶ Brazilian¹⁷ pharmacopoeias have general chapters for dissolution harmonized among them, including its standards and policies. However, dissolution test of OCBZ capsule is not included in any of these pharmacopoeias, thus there is no official method description for this purpose. Within this context, a dissolution test using a simple UV-Vis spectrophotometric method to evaluate the dissolution performance of soft-gel capsules containing oxcarbazepine was developed, optimized and validated.

Experimental

Reagents and materials

The following analytical-grade reagents were used: sodium lauryl sulfate (SLS), hydrochloric acid, tribasic sodium phosphate (all from Vetec, Rio de Janeiro, Brazil) and ethanol (Neon, São Paulo, Brazil). Oxcarbazepine (99.1% purity) (Henrifarma, São Paulo, Brazil) was

used as the reference material and as the raw material for the compounding of the capsules. Ultra-pure water (aquaMAX - Ultra 370 Series, Young Lin, Korea) was used for the dissolution medium and throughout analysis. The OCBZ capsules with 150 mg were compounded using: soft-gel white/white n. 1 capsules (Catalent, Moorestown, NJ, USA) and the following preparation of excipients: pharmaceutical talc (Pharmanostra, Rio de Janeiro, RJ, Brazil), 3%; colloidal silica (Deg, São Paulo, SP, Brazil), 1%; SLS (Mapric, São Paulo, SP, Brazil), 1.5%; corn starch (All Chemistry, São Paulo, SP, Brazil), *q.s.p.* (quantity sufficient for preparation) 100%.

Equipment

OCBZ quantification was performed on a dual beam UV-Visible spectrophotometer system from Cary 50 Probe (Varian, Palo Alto, CA, USA) with 190-1100 nm absorption spectrum, 1.5 nm fixed spectral bandwidth, fiber optic reading probe and quartz regular cells of optical path equal to 1.0 cm.

Dissolution tests were conducted in a dissolutor Mod 299 (Nova Ética, São Paulo, SP, Brazil). The equipment consists of (i) six cylindrical glass vessels with hemispherical bottoms, nominal capacity of 1 L, its sides are flanged at the top, and a fitted cover so that the evaporation of the dissolution media is retarded, (ii) a water bath that keeps the temperature inside the vessel at 37 ± 0.5 °C during the test and (iii) a stainless steel drive shaft to which the dissolution apparatus can be attached and “positioned so that its axis is not more than 2 mm at any point from the vertical axis of the vessel and rotates smoothly and without significant wobble that could affect the results”,¹⁰ (iv) a motor to rotate the shaft, and (v) a speed-regulating device to select and maintain, within 4% of variation, the shaft rotation speed. The dissolution apparatus used were: (i) six stainless steel baskets with external diameter of 22.2 ± 1.0 mm, external height of 36.8 ± 3.0 mm and mesh 40 screen with welded seam and (ii) six stainless steel paddles with height of 19.0 ± 0.5 mm, width of 74.5 ± 0.5 mm, and thickness of 4.0 ± 1.0 mm. The distance between the inside bottom of the vessel and the bottom of the apparatuses was maintained at 25 ± 2 mm. Illustrations of the design of the equipment can be found in the previously cited pharmacopoeias.^{10,14-17}

Other equipments used were ultrasonic bath (Cristófoli, Campo Mourão, PR, Brazil), digital pHmeter W3B (Bel Engineering, Monza, Italy), electronic analytical balance AY 220 (Shimadzu, Kyoto, Japan), stainless steel sieve 60 mesh (Bronzinox, São Paulo, SP, Brazil) and porcelain mortar (Chiarotti, São Paulo, SP, Brazil).

Solutions

All dissolution media used (hydrochloric acid 0.1 mol L⁻¹, pH 6.8 phosphate buffer and SLS 1% m/m aqueous solution) were degassed in ultrasonic bath for 30 min prior to use.

A pH 6.8 buffer potassium phosphate was prepared by adding 76 g of tribasic sodium phosphate in water to obtain 1000 mL of solution. An aliquot of 250 mL of this solution was mixed with 750 mL of hydrochloric acid 0.1 mol L⁻¹. The pH value was checked with the pHmeter and if needed adjusted with hydrochloric acid 2 mol L⁻¹ or sodium hydroxide 2 mol L⁻¹, if necessary.

The standard solutions of OCBZ were prepared in a 1000 mL volumetric flask by dissolving an accurately weighed amount (0.166 g) of OCBZ standard in 10 mL of ethanol and diluting to volume with 1% m/v SLS aqueous solution, hydrochloric acid or sodium phosphate buffer, depending on the dissolution medium used. It was used ultrasonic bath for solubilization of the OCBZ reference substance in the dissolution media. This solution was filtered through a 0.45 µm porosity regenerated cellulose membrane (Macherey-Nagel, Duren, Germany) and used for the spectrophotometric analysis (final concentration *ca.* 166 µg mL⁻¹).

Sample solutions were prepared in six replicates by putting one capsule in each vessel containing the dissolution medium (900 mL) at the temperature of 37 ± 0.5 °C and using the adequate apparatus (final concentration *ca.* 166 µg mL⁻¹). A sample aliquot of 10 mL was withdrawn from each vessel at the end of 60 min and filtered in a quantitative filter paper (Vetec, Rio de Janeiro, RJ, Brazil). This solution was assayed by spectrophotometry at 304 nm, using the respective dissolution medium as blank. Masses of drug dissolved from the capsules were calculated by comparison with the standard solution measured by spectrophotometric analysis.¹⁸

Factorial design with mixed levels

In order to establish a method of analyzing dissolution of OCBZ, the following parameters were evaluated accordingly to criteria of The United States Pharmacopoeia (USP),¹⁰ The U. S. Food and Drug Administration (FDA),¹¹ The International Conference on Harmonization (ICH)¹⁹ and the Brazilian Legislation:²⁰ (i) dissolution medium at pH 6.8 phosphate buffer, HCl 0.1 mol L⁻¹ and 1% m/v SLS aqueous solution, (ii) 1 (basket) and 2 (paddle) concerning apparatus and (iii) stirring speed of 50, 75 and 100 rpm.

To evaluate the significance that each parameter/factor has on the dissolution of OCBZ and to determine the

optimal conditions for inclusion of the method in routine laboratorial tests, a 3² × 2 factorial design with mixed levels containing factors at three (-1, 0, +1 for the factors stirring speed and dissolution medium) and two (-1 and +1 for the factor dissolution apparatus) levels was randomly conducted in a total of 18 experiments with six replicates in each experimental level.²¹ The factors, levels and matrix of contrast coefficients are listed in Table 1.

Each experiment was conducted by adding one OCBZ capsule in a dissolution vessel (n = 6) containing 900 mL of dissolution medium stabilized at 37 ± 0.5 °C (variables kept constant). After 60 min of testing (usual time for immediate release solid pharmaceutical forms),²¹ aliquots of 10 mL were collected, filtered and measured directly in spectrophotometer at 304 nm using the dissolution medium as blank.

After the experiments, statistical analysis was conducted.^{22,23} From the results of each experiment, the coefficients for determining the statistical model prediction were calculated accordingly to equation 1:

$$b = (X^t X)^{-1} X^t y_b \quad (1)$$

where vector *b* represents model parameter estimators, *X* and *y* are matrix and vector respectively, according to Table 1. To achieve the equation of the fitted model, it was calculated the standard errors of the coefficients using equation 2:

$$\varepsilon(b) = \sqrt{(X^t X)^{-1} \sigma^2} \quad (2)$$

where $\varepsilon(b)$ is the matrix of standard errors of the elements of vector *b* and σ^2 is the populational variance of the experiments, which can be estimated as *s*² using equation 3:

$$s_{clustered}^2 = \frac{\sum_{i=1}^{18} v_i s_i^2}{\sum_{i=1}^{18} v_i} \quad (3)$$

where *v_i* and *s_i*² are the number of degree of freedom and the variance within each experimental level, respectively.

Method validation

The dissolution test was validated after establishing the optimal conditions. The validation of a dissolution test can be divided into two parts. The first one concerns equipment validation, once all equipment and glassware used has to be calibrated and qualified.²⁴ Thus, the dissolutor was calibrated and so the glassware, by the Brazilian Calibration

Table 1. Factors, levels, matrix of contrast coefficients and responses for $3^2 \times 2$ mixed experimental design for dissolution of OCBZ capsules

Issues	Mean	Contrast coefficients									Responses								
		X_1	X_2	X_3	X_1^2	X_2^2	X_1X_2	X_1X_3	X_2X_3	$X_1X_2X_3$	% dissolved (n = 6)						Average	Variance	DF
1	1	-1	-1	-1	1	1	1	1	1	-1	50.46	51.02	51.57	49.16	51.14	49.97	50.55	0.78	5
2	1	0	-1	-1	0	1	0	0	1	0	62.02	64.57	60.78	61.15	63.57	60.90	62.17	2.46	5
3	1	1	-1	-1	1	1	-1	-1	1	1	63.32	65.87	65.43	64.25	67.17	67.29	65.56	2.49	5
4	1	-1	0	-1	1	0	0	1	0	0	49.39	49.58	51.31	48.71	50.20	50.07	49.88	0.77	5
5	1	0	0	-1	0	0	0	0	0	0	58.89	57.90	57.52	56.84	59.14	59.01	58.22	0.88	5
6	1	1	0	-1	1	0	0	-1	0	0	61.21	61.15	62.14	63.63	60.96	60.65	61.62	1.22	5
7	1	-1	1	-1	1	1	-1	1	-1	1	85.96	85.83	84.91	82.43	85.09	83.17	84.56	2.08	5
8	1	0	1	-1	0	1	0	0	-1	0	87.97	88.34	91.18	92.05	87.47	84.38	88.57	7.62	5
9	1	1	1	-1	1	1	1	-1	-1	-1	87.69	86.51	85.33	88.93	89.11	86.63	87.37	2.21	5
10	1	-1	-1	1	1	1	1	-1	-1	1	52.62	51.63	53.67	52.68	52.19	54.10	52.81	0.85	5
11	1	0	-1	1	0	1	0	0	-1	0	66.09	64.93	65.00	64.39	66.29	66.77	65.58	0.87	5
12	1	1	-1	1	1	1	-1	1	-1	-1	69.97	66.92	67.86	65.19	66.29	66.77	67.17	2.64	5
13	1	-1	0	1	1	0	0	-1	0	0	50.07	51.06	50.01	50.50	49.57	50.44	50.28	0.26	5
14	1	0	0	1	0	0	0	0	0	0	60.30	59.24	59.31	58.75	60.49	60.92	59.83	0.73	5
15	1	1	0	1	1	0	0	1	0	0	62.33	61.78	62.21	63.70	62.71	64.26	62.83	0.91	5
16	1	-1	1	1	1	1	-1	-1	1	-1	82.90	83.65	79.37	84.14	78.68	80.98	81.62	5.25	5
17	1	0	1	1	0	1	0	0	1	0	94.16	93.10	92.91	95.59	91.92	97.40	94.18	4.05	5
18	1	1	1	1	1	1	1	1	1	1	89.95	90.57	90.20	90.07	89.70	90.44	90.15	0.10	5

X_1 : stirring speed (rpm): (-1):50; (0):75; (+1):100; X_2 : dissolution medium: (-1): phosphate buffer pH 6.8; (0): HCl 0.1 mol L⁻¹; (+1): SLS 1% m/v; X_3 : apparatus: (-1): basket; (+1): paddle; DF = degrees of freedom; % dissolved = $\frac{C_{\text{standard}}}{A_{\text{standard}}} \times \frac{A_{\text{sample}}}{C_{\text{sample}}} \times P$, where C is the concentration, A is the absorbance and P is the purity/potency of the standard.

Network. The second part regards the test validation itself. In this case, the dissolution test is categorized by USP¹⁰ and Brazilian legislation²⁵ as “category III - performance test”, being the parameter precision required. It were also evaluated the parameters linearity and limits of detection and quantification.²⁶

Linearity

The test was performed from three analytical curves, plotted from the concentrations of 50-150% of the OCBZ work concentration (real concentrations: 83, 124, 166, 208 and 249 $\mu\text{g mL}^{-1}$), to assess a linear relationship between analyte concentration and absorbance. For this purpose, data from each concentration level were statistically evaluated using analysis of variance (ANOVA) and the value of the correlation coefficient of the analytical curve.

Limits of Detection and Quantification

The limit of detection (LOD) and limit of quantification (LOQ) of the proposed method were determined from three standard analytical curves and they were calculated using equations 4 and 5:

$$LOD = S \frac{3}{a} \quad (4)$$

$$LOQ = S \frac{10}{a} \quad (5)$$

where S is the standard deviation of the intercept with the y axis of the three analytical curves and a is their slope.

Precision

The precision was assessed by the coefficient of variation, determined for the repeatability and the intermediate precision. The repeatability (intra-assay precision) was assessed by analyzing seven replicates at the concentration of 166.6 $\mu\text{g mL}^{-1}$ of OCBZ (100%). The intermediate precision consisted of the same test, but performed for two consecutive days with two different analysts. A coefficient of variation < 5% was considered appropriate.²⁵

Software

All statistical analysis was performed in Microsoft Office® Excel 2007 software, which is easy to use and therefore can be widely applied in teaching the topics elaborated upon here.

Results and Discussion

Factorial design is a very useful tool to obtain deeper information about the relationships among the studied variables and to establish the optimum experimental conditions. Its application to dissolution studies is needful for reducing the time of method development and to provide

less ambiguous data.²⁷ The application of experimental design for optimizing the dissolution test is a valuable tool for reducing the amount of work needed for formulation and test development. In fact, the factorial design makes possible the reduction of the number of single experiments and the improvement of the method reliability.²⁸

In the present study, each experiment was conducted in a number of 6 replicates because of the design of the equipment, which contains 6 dissolution vessels and because the pharmacopoeic acceptance criterion states that is needed to have 6 units being tested in order to achieve an adequate overview of the dosage form dissolution quality. The pharmacopoeias have harmonized general chapters for dissolution test, and they recommend that the first 6 units must give results fallen within the specification “ $Q + 5\%$ ”, being Q the amount of dissolved active ingredient specified in the individual monograph, expressed as a percentage of the labeled content of the dosage unit. This value is generally 75-80%.²⁹ However, it is acceptable that the dosage form does not meet this first requirement. In this case, another test must be conducted therefore with additional 6 units (criterion: the average of the 12 units, the first 6 and the additional ones, must be equal to or greater than Q , and no unit is less than “ $Q - 15\%$ ”). Once the results do not show adequate values, the test is repeated with more 12 capsules (specification: the average of the all 24 units must be equal to or greater than Q , not more than 2 units are less than “ $Q - 15\%$ ” and no unit is less than “ $Q - 25\%$ ”).^{10,14,16,17}

When there is no official monograph for a given medicine, the method has to be developed and validated, and the referred Q value can be selected by the authors, provided it has a plausible justification. In this case, it was chosen to work with $Q = 80\%$, because of its widespread use by the pharmacists. The other test parameters had also to be selected. According to the Brazilian legislation,²⁰ one has to investigate three different dissolution media (all within the physiological pH 1.2-6.8). Oxcarbazepine is well-characterized as a water-insoluble drug,¹⁰ thus it is required to use dissolution media dissimilar from the normally employed in this type of test. In this sense, the utilization of surfactants such as the SLS is largely widespread, once it mimics, up to some extent, the human body physiological conditions, such as the presence of natural surfactants (bile salts and lecithin, for instance) in the gastrointestinal tract. Surfactants as the SLS are wetting, micellar solubilizing agents, so they have the capacity of enhancing the solubility of such drugs. This probably happens via the reduction of the interfacial tension and/or their association with the excipients and the active ingredient itself. Nevertheless, it is recommended that these substances are used only in small concentrations, so that the *in vitro* and *in vivo* results are better

correlated,²⁹ so that is why it was chosen to work with the concentration of 1% m/v. The other media was chosen to verify whether the substance is better solubilized in acidic or neutral pH.

In addition to the dissolution medium, it were also measured the influence of two stirring apparatus (basket and paddle, the most commonly used) and three different stirring speeds (50, 75 and 100 rpm, also the most commonly used in the in the pharmaceutical field). The dissolution experiment was therefore conducted using a $3^2 \times 2$ mixed-level factorial design, in a total of 18 experiments, whose parameters are described in Table 1, as well as the contrast coefficients for the interaction effects of the factors and the results of each experiment.

In Table 1, can be seen that keeping X_3 (apparatus) fixed at level -1 , it appears that the increased level of X_1 (stirring speed), when X_2 (dissolution medium) is kept constant, leads to higher response, except when X_2 is at level $+1$. In this case, the highest response occurs when X_1 is at level 0. The same analysis is true when one considers the level X_3 fixed at $+1$, which suggests that the answer related to X_1 is likely to forecast, with the two other factors held constant. Similarly, when X_3 remains fixed at level -1 , the response as a function of X_2 , with X_1 fixed, also follows predictable behavior, but differently from what happens in the previous situation. With X_2 , the lower response is observed at level 0 and the highest at level $+1$. When X_1 and X_2 are set constant to investigate the effect of X_3 on the response, it is observed that changing the level -1 to $+1$ always leads to an increased response. Thus, from this initial screening, it appears that higher responses are observed when the agitation speed is at levels $+1$ (100 rpm, when X_2 is at -1 and 0) and 0 (75 rpm, when X_2 is $+1$), the dissolution medium at level $+1$ (1% m/v SLS) and the apparatus at level $+1$ (paddle). However, it was performed a study involving factorial surface response for the experiments in order to achieve and verify the optimal conditions for the dissolution test.

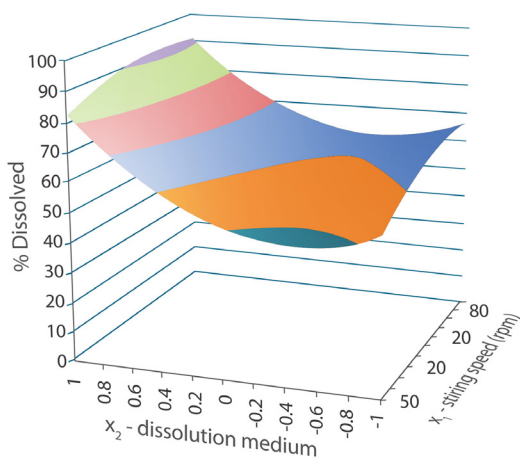
Calculating the coefficients of the statistical model using the equations 1, 2 and 3 (Tables S1 and S2), it was obtained $\hat{y} = 60.03 (\pm 0.30) + 5.41 x_1 (\pm 0.17) + 13.55 x_2 (\pm 0.17) + 0.88 x_3 (\pm 0.14) - 4.38 x_1^2 (\pm 0.29) + 17.08 x_2^2 (\pm 0.29) - 2.25 x_1 x_2 (\pm 0.20) + 0.49 x_1 x_3 (\pm 0.17) - 0.15 x_2 x_3 (\pm 0.17) + 0.79 x_1 x_2 x_3 (\pm 0.20)$. The significance evaluation for each error using t value (*ca.* 2.0) with 90 degrees of freedom was performed and only coefficient $x_2 x_3$ was considered no significant for 95% confidence, because contain zero within the interval. In order to verify the model lack of fit, it was performed the analysis of variance according to Table 2. The percentage of variation explained by regression and maximum percentage of variation explained were equal to 98.70 and 99.23%, respectively.

Table 2. Analysis of variance for fitting the model of prediction for the screening study

Source	Sum of square (SS)	Number of degrees of freedom	Mean of square (MS)
Model	23175.86	9	2675.09
Residual	305.64	98	3.12
Lack of fit	124.77	8	15.57
Pure error	180.87	90	2.01
Total	23481.50	107	

% variation explained = 98.70; % maximum of variation explained = 99.23.

The regression significance test returned the value of $F_{\text{calculated}} (MS_{\text{model}} / MS_{\text{residual}}) = 825.68$, higher than $F_{\text{critical}0.05, v_1=9, v_2=98} \text{ ca. } 2.00$, which indicates a significant regression within 95% confidence interval. On the other hand, the lack of fit test returned $F_{\text{calculated}} (MS_{\text{lack of fit}} / MS_{\text{pure error}}) = 7.76$, higher than $F_{\text{critical}0.05, v_1=8, v_2=90} \text{ ca. } 2.06$, which indicates lack of fit within 95% confidence interval. However, taking into account that the lack of fit presented by the model is relatively small and probably represents a minor region in the surface; that the percentage of variation explained by regression and maximum percentage of variation explained are close; and the model residue presented normal behavior through Kolmogorov normality test for 95% confidence ($p\text{-value} > 0.05$) The response surface was built from the experiments (Figure 2 and S1) and indicated as higher response the region (% dissolution) between 40-80 rpm (Table S3), using 1% m/v SLS. This response surface was achieved fixing variable apparatus in "paddle" (level +1). This was made because both apparatuses lead to similar results in the dissolution experiments, thus the decision was made taking into account that the paddle has a more widespread utilization

**Figure 2.** Response surface for the $3^2 \times 2$ mixed factorial design. X_1 = stirring speed (rpm); X_2 = dissolution medium.

than the basket. With respect to the stirring speed, once 40, 60 and 80 rpm led to very similar theoretical dissolution result, it was chosen to work with 80 rpm.

The optimal conditions (stirring speed set at 80 rpm) would theoretically lead to a percentage of dissolution of about 92.15%. Since these conditions provided by the response surface methodology had not been tested previously, the experiment was run again using the provided levels by the methodology, with six replicates. The average percentage dissolved for these conditions was 95.45%, higher than that found in the initial screening experiments and even higher than the predicted value obtained with the response surface methodology (in the whole optimized region for stirring speed, i.e. 40-80 rpm). Though it has presented a lower value compared to the real one, the model was able to predict which one were the best levels for each parameter of the test. So, it can be inferred that this methodology is highly relevant to the objectives proposed here. Therefore, the established conditions to validate the dissolution test were: $X_1 = 80$ rpm, $X_2 = \text{SLS } 1\% \text{ m/v aqueous solution}$ and $X_3 = \text{paddle}$.

Method validation

The linearity of the method used was evaluated on a calibration curve of the absorbance vs. the analyte concentration ($\mu\text{g mL}^{-1}$). The calibration curves, performed in genuine triplicate, returned the model $y (\pm 0.0035) = 0.0096 (\pm 0.000015) x + 0.0029 (\pm 0.0027)$, with $r^2 = 0.99996$. To validate this model, it was performed the analysis of variance described in Table 3. The test of significance of regression returned the value of $F_{\text{calculated}} (MS_{\text{model}} / MS_{\text{residual}}) = 391107.8$, greater than $F_{\text{critical}0.01, v_1=1, v_2=13} = 4.67$, which confirms the existence of a significant linear relationship between the two variables, with 95% confidence. The test of lack of fit value returned $F_{\text{calculated}} (MS_{\text{lack of fit}} / MS_{\text{pure error}}) = 5.06$, smaller than

Table 3. Analysis of variance for fitting the model of prediction for the validation study

Source	Sum of square (SS)	Number of degrees of freedom	Mean of square (MS)
Model	4.80	1	4.80
Residual	1.59×10^{-4}	13	1.22×10^{-5}
Lack of fit	9.63×10^{-5}	3	3.20×10^{-5}
Pure error	6.33×10^{-5}	10	6.33×10^{-6}
Total	4.80	14	

% variation explained by regression = 99.99; % maximum of variation explained = 99.99.

$F_{\text{critical } 0.01, v_1=3, v_2=10} = 6.55$, which indicates no lack of fit, with 99% confidence.

The lowest concentration of an analyte in a sample that it is possible to detect, but not to quantify is considered the LOD. The LOQ has the same definition, but determines the lowest concentration of an analyte in a sample that can be determined/quantified with acceptable precision and accuracy.^{30,31} By calculations using equations 4 and 5, the LOD was determined as $0.84 \mu\text{g mL}^{-1}$ and the LOQ as $2.81 \mu\text{g mL}^{-1}$, which demonstrates the method is sensitive enough to the proposed objective. The calculation of the LOQ and LOD is useful, in the dissolution case, because some capsules do not dissolve adequate amounts of the active ingredient due to problems in the formulation, caused mainly by the choice of the excipients. Thus the method has to be sensitive enough to be applied to a wide range of possibilities found in the pharmaceutical industries.

The precision of the method (repeatability and intermediate precision) is a measure of the degree of repeatability of an analytical method under normal operation. Its evaluation is very important in order to assess the reliability of the dissolution test data.²⁵ In the case of dissolution test (as previously informed, a “category III - performance test”), it is needed to verify the repeatability through a minimum of 6 determinations/experiments with the work concentration of the substance (100%, *i.e.* $166 \mu\text{g mL}^{-1}$). A coefficient of variation (CV) of less than 5% is considered appropriate.^{30,31} It was achieved repeatability values of 3.0% ($n = 7$).

For the intermediate precision, it is recommended to perform the test in two different days with two different analysts, aiming to assess the inherent variability of the method. For this test, it is also considered as satisfactory a CV value lower than 5%. The intermediate precision coefficient of variation achieved was 4.2% ($n = 14$).

Other parameters such as accuracy and specificity were not performed because the legislation²⁵ does not consider them as mandatory for dissolution tests. Since the emphasis of this study was not the validation, but the application of the mixed-level factorial design in dissolution tests development, the authors chose not to include these two figures of merit.

To sum up, this work opens a new perspective in the dissolution test development methodology, since it is the first time that the mixed-level factorial design was used for this purpose. As it was successfully applied for the OCBZ capsules case, it can be inferred that it can be also applied for virtually any drug that needs a dissolution study. Using the fundamentals here presented, one can develop it for a variety of pharmaceutical dosage forms, composed by several different active ingredients.

Conclusions

On balance, this work provided a simple and validated procedure for dissolution test of oxcarbazepine capsules by UV-Vis spectrophotometric analysis. The $3^2 \times 2$ mixed factorial design carried out an optimized dissolution test for: apparatus 2 (paddle) at 80 rpm and sodium lauryl sulfate 1% m/v aqueous solution, as the dissolution medium. Data analysis demonstrates the method was validated with respect to linearity, precision and limits of detection and quantification. The method is useful and adequate for oxcarbazepine capsules dissolution test, once there is no official monograph regarding it. Moreover, the mixed-level factorial design presented can be applied for the dissolution method development of virtually any drug substance candidate, or even the already marketed ones.

Supplementary Information

Supplementary information is available free of charge at <http://jbc.org.br> as a PDF file.

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References

1. Buggy, Y.; Layton, D.; Fogg, C.; Shakir, S. A. W.; *Epilepsia* **2010**, *51*, 818.
2. Novartis Pharmaceuticals; *Trileptal (R), Summary of Product Characteristics*, 2009.
3. Shorvon, S.; *Seizure* **2000**, *9*, 75.
4. Tecoma, E. S.; *Epilepsia* **1999**, *40*, S37.
5. Bill, P. A.; Vigonius, U.; Pohlmann, A.; Guerreiro, C. A.; Kochen, S.; Saffer, D.; Moore, A.; *Epilepsy Res.* **1997**, *27*, 195.
6. Dam, M.; Ekberg, R.; Løyning, Y.; Waltimo, O.; Jakobsen, K.; *Epilepsy Res.* **1989**, *3*, 70.
7. Schmidt, D.; Elger, C. E.; *Epilepsy Behav.* **2004**, *5*, 627.
8. Agência Nacional de Vigilância Sanitária (ANVISA); *Cr terios adicionais de Boas Pr ticas de Manipula o de Medicamentos em Farm cias*, Resolu o RDC n. 354, Brasil, 2003.
9. Cohen, J. L.; Hubert, B. B.; Leeson, L. J.; Rhodes, C. T.; Robinson, J. R.; Roseman, T. J.; Shefter, E.; *Pharm. Res.* **1990**, *7*, 983.

10. The United States Pharmacopeial Convention, *The United States Pharmacopeia*, USP 33, Rockville, 2010.
11. The United States Food and Drug Administration (USFDA), *Guidance for Industry: Dissolution Testing of Immediate Release Solid Oral Dosage Forms*, Food and Drug Administration, Rockville, 1997.
12. Frost, S. M. A.; *Dissolution Technol.* **2004**, *11*, 19.
13. Davydova, N.; Stippler, E.; Jin, P.; Giancaspro, G.; *J. Pharm. Biomed.* **2010**, *53*, 295.
14. *European Pharmacopoeia*, 6th ed., Council of Europe: Strasbourg, 2008.
15. *British Pharmacopoeia*, Her Majesty's Stationery Office: London, 2010.
16. The Pharmaceutical Society of Japan, *Japanese Pharmacopoeia*, 15th ed., Tokyo, 2006.
17. *Farmacopéia Brasileira*, 4a. ed., Atheneu Editora: São Paulo, Brasil, 2005.
18. Corrigan, O. I.; Stanley, T.; *J. Pharm. Pharmacol.* **1982**, *34*, 621.
19. ICH, International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use, Q2B(R1), *Guideline on Validation of Analytical Procedure: Text and Methodology*, Geneve, 2005.
20. Agência Nacional de Vigilância Sanitária (ANVISA); *Estudos de Equivalência Farmacêutica e de Perfil de Dissolução Comparativo*, Resolução RDC n. 31, Brasil, 2010.
21. Agência Nacional de Vigilância Sanitária (ANVISA); *Guia para Ensaio de Dissolução para Formas Farmacêuticas Sólidas Oraís de Liberação Imediata (FFSOLI)*, Resolução RDC n. 483, Brasil, 2010.
22. Montgomery, D. C.; *Design and Analysis of Experiments*, 6th ed., Wiley: New York, 2004.
23. Barros Neto, B.; Scarminio, I. S.; Bruns, R. E.; *Como fazer Experimentos: Pesquisa e Desenvolvimento na Ciência e na Indústria*, 4a. ed., Bookman: Porto Alegre, Brasil, 2010.
24. Frick, A.; Moller, H.; Wirbitzki, E.; *Eur. J. Pharm. Biopharm.* **1998**, *46*, 305.
25. Agência Nacional de Vigilância Sanitária (ANVISA); *Validação de Métodos Bioanalíticos*, Resolução RE n. 899, Brasil, 2003.
26. Ansari, M.; Kazemipour, M.; Talebnia, J.; *Dissolution Technol.* **2004**, *11*, 16.
27. Mohd, A. A.; Dean, J. R.; Tomlinson, W. R.; *Analyst* **1992**, *117*, 1743.
28. Furlanetto, S.; Maestrelli, F.; Orlandini, S.; Pinzauti, S.; Mura, P.; *J. Pharm. Biomed. Anal.* **2003**, *24*, 159.
29. Shah, V. P.; Konecny, J. J.; Everett, R. L.; McCullough, B.; Noorzadeh, A. C.; Skelly, J. P.; *Pharm. Res.* **1989**, *6*, 612.
30. Shabir, G. A.; *J. Chromatogr. A* **2003**, *987*, 57.
31. Instituto Nacional de Metrologia (INMETRO); *Normalização e Qualidade Industrial, Orientações sobre Validação de Métodos de Ensaio Químicos*, Documento DOQ-CGCRE-008, Brasil, 2003.

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Supplementary Information

Optimization of a New Dissolution Test for Oxcarbazepine Capsules using Mixed-Level Factorial Design

Hudson C. Polonini,^a Marcone A. L. de Oliveira,^b Anderson O. Ferreira,^c
 Nádia R. B. Raposo,^a Livia N. Grossi^c and Marcos A. F. Brandão^{*a}

^aFaculty of Pharmacy and ^bDepartment of Chemistry, Federal University of Juiz de Fora, 36036-900 Juiz de Fora-MG, Brazil

^cOrtofarma – Controle de Qualidade, 36120-000 Matias Barbosa-MG, Brazil

Table S1. “b” Coefficient calculation

	Coefficients	Significant intervals	
Mean	60.03591	59.42733	60.64448999
X ₁	5.415861	5.082528	5.749194615
X ₂	13.55133	13.218	13.8846618
X ₃	0.886864	0.614698	1.1590292
X ₁₂	-4.3891	-4.96645	-3.81174705
X ₂₂	17.08021	16.50286	17.65755911
X ₁ X ₂	-2.25243	-2.66068	-1.84418205
X ₁ X ₃	0.490914	0.157581	0.824247368
X ₂ X ₃	-0.15214	-0.48547	0.181192138
X ₁ X ₂ X ₃	0.798203	0.389955	1.206451261

The coefficients were calculated using: $(X^T X)^{-1} X^T y$; significant intervals were calculated using: $b_i \pm \text{error} \times t_{0.05, v=90}$

Table S2. Standard error calculation of the “b” coefficients

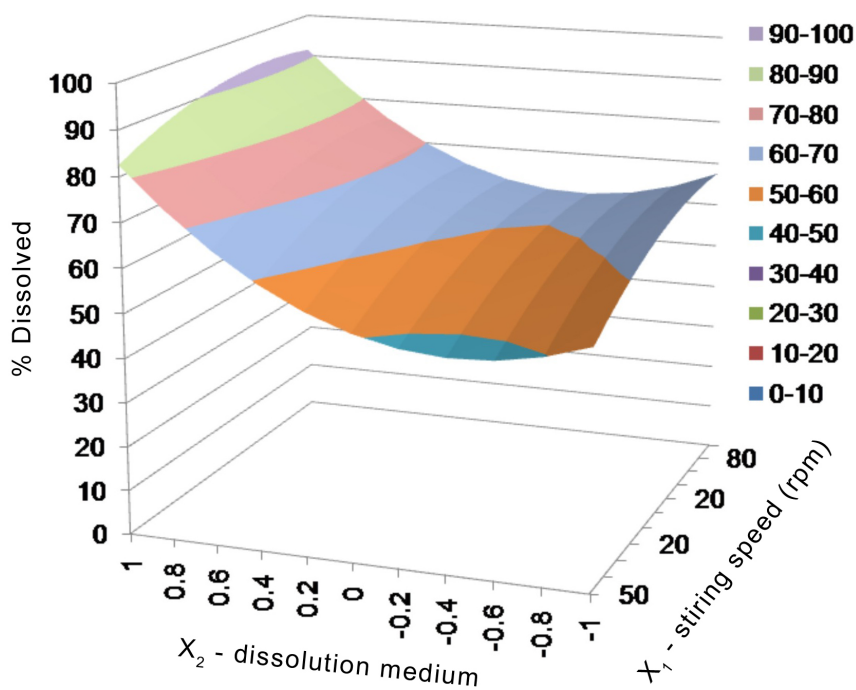
0.30429	0	0	0	0	0	0	0	0	0
0	0.166667	0	0	0	0	0	0	0	0
0	0	0.166667	0	0	0	0	0	0	0
0	0	0	0.136083	0	0	0	0	0	0
0	0	0	0	0.288675	0	0	0	0	0
0	0	0	0	0	0.288675	0	0	0	0
0	0	0	0	0	0	0.204124	0	0	0
0	0	0	0	0	0	0	0.166667	0	0
0	0	0	0	0	0	0	0	0.166667	0
0	0	0	0	0	0	0	0	0	0.204124

The standard errors were calculated as the square root of $(X^T X)^{-1} \times s_{\text{clustered}}^2$.

*e-mail: marcosbrand@uol.com.br

Table S3. Plotting of the response surface. The levels were substituted in the statistical model in order to find the region of best response

		X_2									
	1	0.8	0.6	0.4	0.2	0	-0.2	-0.4	-0.6	-0.8	
	50	82.56052388	73.44097	65.68782	59.3011	54.28079	50.6269	48.33943	47.41837	47.86373	49.6755
	40	85.03110851	75.96972	68.27475	61.94619	56.98405	53.38833	51.15903	50.29614	50.79966	52.66961
	30	87.15056535	78.14735	70.51054	64.24016	59.33619	55.79863	53.6275	52.82278	53.38447	55.31259
	20	88.9188944	79.97384	72.39521	66.18299	61.33719	57.85781	55.74484	54.99829	55.61816	57.60444
	10	90.33609567	81.44921	73.92875	67.7747	62.98707	59.56585	57.51106	56.82267	57.50071	59.54516
X_1	0	91.40216915	82.57346	75.11116	69.01528	64.28582	60.92277	58.92614	58.29593	59.03214	61.13476
	20	92.11711485	83.34657	75.94244	69.90473	65.23344	61.92856	59.9901	59.41806	60.21243	62.37322
	40	92.48093276	83.76856	76.4226	70.44306	65.82994	62.58323	60.70294	60.18906	61.0416	63.26056
	60	92.49362289	83.83942	76.55163	70.63026	66.0753	62.88676	61.06464	60.60894	61.51965	63.79678
	80	92.15518523	83.55915	76.32953	70.46633	65.96954	62.83917	61.07522	60.67768	61.64656	63.98186
	100	91.46561979	82.92775	75.7563	69.95127	65.51265	62.44045	60.73467	60.3953	61.42235	63.81582

**Figure S1.** Response surface for the 3² × 2 mixed factorial design. X₁ = stirring speed (rpm); X₂ = dissolution medium.