

Chemical and Spectroscopical Characterization of Humic Acids from two South Brazilian Coals of Different Ranks

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Ácidos húmicos (HA) extraídos de carvões de diferentes ranks, de suas amostras regeneradas e de uma amostra nitrada, foram caracterizados por análise elementar e por espectroscopia de infra-vermelho (FTIR), de ressonância magnética nuclear de ¹³C no estado sólido (NMR) e de ressonância paramagnética eletrônica (EPR). O HA de carvão de baixo rank apresentou maiores valores de teor de C e razão C/N e menores valores de teor de O e razões H/C e O/C do que o de alto rank. Resultados de RMN mostraram que ambas amostras foram mais aromáticas e menos carboxílicas do que HA comuns em solos. Estas características podem limitar o uso de HA de carvão como condicionadores e fertilizantes adequados para solos. A regeneração não provocou alterações relevantes, exceto pelo decréscimo no teor de radicais livres determinados por espectroscopia de EPR. Provavelmente as condições e o tempo de regeneração não foram adequados para oxidar as amostras. Os espectros de FTIR obtidos foram semelhantes, excentuando-se aquele da amostra nitrada, onde a banda de absorção em 1533 cm⁻¹ comprova a presença de grupos nitrogenados. A nitratação aumentou o teor de N e reduziu a razão C/N a valores comparáveis aos observados em HA de solos, porém a aromaticidade permaneceu alta e o teor de carboxílicos diminuiu após o processo.

Humic acids (HA) extracted from two coals of different ranks, from their regenerated samples and from a nitrated sample, were characterized by elemental analysis and by infra-red (FTIR), solid state ¹³C nuclear magnetic resonance (NMR) and electronic paramagnetic resonance (EPR) spectroscopies. The low rank coal HA presented higher C and lower O contents, higher C/N and lower H/C and O/C ratios than high rank coal HA. NMR results showed that both samples were more aromatic and less carboxylic than common soil HA. Those characteristics may limit the coal HA efficiency as an appropriate soil conditioner and fertilizer. The regeneration process did not produce major alterations in the coal HA, except a decrease of the free radical content as determined by EPR spectroscopy. Probably, the regeneration conditions and time were not adequate to oxidize the samples. The obtained FTIR spectra were much alike, except that from the nitrated sample, where the absorption band at 1533 cm⁻¹ confirms the presence of nitrated groups. The nitration process increased the N content and reduced the C/N ratio to values comparable to those reported for soil HA, but the aromaticity still remained high and the carboxylic content was lowered after the procedure.

Keywords: humic substances, CP/MAS-NMR, MAS-NMR, FTIR, EPR, coal regeneration, aromaticity, free radicals, coal nitrification

Introduction

The soil organic matter has a vital role in maintaining soil quality.¹ The application of organic matter in degraded soils is agronomically very important because it restores soil fertility, improves soil aggregation and hinders soil

erosion, among other effects.^{2,3} Amendments to soils of composted organic matter from different sources, like animal manure and sewage sludges, is ecologically very attractive since it also provides an useful fate for those residues.^{4,5,6} Another promising alternative for the use and recycling of non-pedological humic substances is its use in hydroponic solutions to produce plantules under controlled conditions.⁷

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Humic acids (HA) occur naturally in immature coals and can be easily extracted with alkaline solution.^{8,9} On the other hand, the amount of extractable HA in mature coals, like bituminous coals, is lower.^{9,10} As the coal rank increases, *i.e.* coal matures, the carbon content gradually increases (from about 700 g.kg⁻¹ in low rank coals to about 950 g.kg⁻¹ in high rank coals) with a corresponding decrease in the oxygen content.⁹ This behavior has also been observed in the respective HA. Additionally, an increase in the aromaticity degree of HA with the coal rank is also reported.⁹ In comparison to soil HA, coal HA contain in general a higher content of C and lower contents of O and N.^{11,12} The reactions of humic substances are determined by their chemical composition and molecular structure¹ and the quantity and quality of HA in the sample may influence the significance of coal as a soil fertilizer.¹³⁻¹⁵ However, the amount of HA in coals can be increased by oxidizing the sample,^{3,16} resulting what is called "reversed/regenerated coal". Regeneration can be achieved by employing oxidizing agents like oxygen, nitric acid, alkaline permanganate and hydrogen hydroxide, or by exposing the coal sample to the atmospheric oxygen at the earth surface for a larger period, what also leads to the formation of weathered/regenerated coals.⁹ The addition of humic substances extracted from an oxidized coal to a loamy soil and to a sandy-loamy soil was able to retard the onset of the runoff process under rainfall and a significant reduction of soil loss was also observed.¹⁵ Furthermore, an increase of soil cation exchange capacity, pH and organic matter was verified after the incorporation of coal derived humic acids in two Nigerian low-fertile tropical soils.¹⁷ These improvements reflected in a higher crop productivity and showed that the coal humic substances are potentially good soil conditioners.

The main objectives of this study are: (1) to characterize HA from different coal samples through appropriate chemical and spectroscopic techniques (*e.g.*, elemental analysis, FTIR, solid state ¹³C-NMR, EPR); (2) to evaluate the influence of the coal rank, of the regeneration and nitration processes on their chemical structure and composition.

Experimental

Coal samples

The studied coal samples, collected by bulk sampling, were as following: a high rank coal from Cambuí Mine, Figueiras, Paraná State, Brazil (HI); a regenerated high rank coal from a 10 years old pile from Cambuí Mine (HR); a

low rank coal from Seival Mine, Candiota, Rio Grande do Sul State, Brazil (LO); a regenerated low rank coal from a 15 years old pile from Seival Mine (LR). The fifth studied sample (LN) was obtained by nitrating the sample LO with concentrated HNO₃ at room temperature (soil:solution ratio 8:2). The coal samples were dried at 40 °C, grounded in a quartz mortar and kept in plastic flasks at 4 °C to prevent oxidation.

Humic acids isolation

The humic acid extraction was based on Swift.¹⁸ The coal sample (0.1 g) was shaken during 2 h with 30 mL of 0.1 mol. L⁻¹ HCl; the acid supernatant was separated by centrifugation (10 minutes, 2000 rpm) and discarded. This procedure was repeated three times. After the acidic pretreatment, the samples were extracted with 30 mL of 0.5 mol. L⁻¹ NaOH under mechanical shaking during 3 h. The alkaline extract was separated by centrifugation and stored in a polyethylene flask. This procedure was performed three times. The HA was separated by precipitation with 0.1 mol. L⁻¹ HCl added to the extract until pH 2 was reached. After 24 h, the precipitated HA was separated from the fulvic acid (FA) by centrifugation and purified by treating the sample with 5% HCl / HF (1:1) solution for 2 h under mechanical shaking (three times). The purified HA was washed three times with distilled water, frozen overnight, and dried at 60 °C in vacuum oven. The dried samples were maintained in a dessicator with silica gel before further analysis. For all coal samples, the fulvic acid fraction was of such low yield as to be ignorable (FA content < 0.01% of total humic substances).

Elemental analysis

The HA elemental composition was determined by a Perkin Elmer 2400 elemental analyzer and the content of O was calculated by difference of 1000 mg.g⁻¹ after correcting the C, H and N values for an ash-free basis. The ash content was obtained gravimetrically after calcinating the HA sample at 750 °C for 4 h.

Infrared Spectroscopy

Fourier transform infrared (FTIR) spectra of pellets samples (100 mg KBr and 1 mg HA) were recorded on a Win-Bomem & Michaelson spectrometer on a spectral range of 4000 to 400 cm⁻¹, employing a resolution of 4 cm⁻¹, 32 scans and 21 scans.min⁻¹. The band assignments were made according to Mangrich *et al.*,⁶ Baranciková *et al.*¹⁹ and Rivero *et al.*¹⁹

¹³C Nuclear Magnetic Resonance

The solid state ¹³C-NMR spectra were obtained on a Varian INOVA-300 spectrometer (VT CP/MAS probe). Two experimental techniques were tested: (1) single pulse-magic angle spinning technique (MAS), (2) cross polarization with magic angle spinning technique (CP/MAS). The following experimental conditions were employed: (1) ¹³C-MAS: a resonance frequency of 75.4 MHz, a 90° pulse width, a pulse delay of 100 s, 500 to 900 accumulation transients (corresponding to an overnight acquisition), a 7 mm silicium nitrate rotor with Kel-F caps and a MAS spinning rate of 5.60 KHz; (2) ¹³C-CP/MAS: a resonance frequency of 75.4 MHz, a 90° pulse width, a pulse delay of 2 s, contact time of 500 μs, 3200 accumulation transients (corresponding to an overnight acquisition), a 7 mm silicium nitrate rotor with Kel-F caps and a MAS spinning rate of 5.60 KHz. ¹³C chemical shifts are reported relative to hexa-methylbenzene. Chemical shifts were assigned according to Kögel-Knabner:²¹ alkyl-C from 0 to 65 ppm ; O-alkyl-C from 65 to 100 ppm; aromatic-C from 100 to 165 ppm and carboxylic-C from 165 ppm to 190 ppm. The abundance of each carbon type was estimated by integration of the respective spectral region. The region 190 ppm to 230 ppm was not integrated because the MAS rotation was limited to 5600 Hz.

Electron Paramagnetic Resonance

The EPR spectra were obtained in the solid state at room temperature (300 K) in quartz tubes. A Bruker ESP 300E spectrometer was used operating at a frequency of 9.5 GHz (X-band), with a 100 KHz modulation frequency, 2.024 G modulation amplitude and 20 mW microwave power. HA free radicals were detected and quantified using the approximation:²² intensity × ΔH². The areas of the EPR peaks were calibrated with that corresponding to the EPR signal of a “weak pitch” reference of known free radical content obtained from Bruker.

Results and Discussion

Elemental composition

The elemental composition and elemental ratios of the humic acids are listed in Table 1. The obtained values for C, N, H and O contents are within the range observed for a diversified group of coal HA's.⁹ The coal HA from the Seival Mine (HA-LO) differed from that from the Cambuí Mine (HA-HI) basically due to the higher C and lower O and N contents, higher C/N and lower O/C ratios. The increase in the molecular condensation and in the carbon reduction along the coalification development usually reported for bulk coal samples,^{9,10,11} was not verified for the coal humic acids in the present study. It seems that the coal rank did not affect their HA chemical composition. HA with such composition as that verified in the present study are expected to be quite stable with respect to mineralization process.¹⁹ The O/C and C/N ratios of samples HA-HI and HA-LO were higher and lower, respectively, than those reported for Australian brown coal HA samples.^{8,12} However, when compared to soil samples, HA-HI and HA-LO presented higher C, lower N and O contents than those reported for HA from Brazilian soils²³ and tropical Venezuelan soils.²⁰ On the other hand, similar values for C, H and O contents and for H/C and O/C ratios observed in the present study have been obtained for HA from Slovakian soils¹⁹ and from sheep manure.⁶

The chemical composition alterations caused by the regeneration process were different in the two coal samples (Table 1). In the high rank coal HA (HA-HR) a C content increase and an O content decrease were observed, leading to a lowering of the O/C ratio. In the low rank coal HA, a decrease in the C/N ratio due to the N content increase, was observed in the regenerated sample (HA-LR).

Values for N content and C/N ratio comparable to those reported in soil HA,^{1,19,20,23} were successfully obtained by the nitration procedure (Table 1). Nevertheless, the O/C and H/C ratios in the nitrated sample (HA-LN) still remained relatively low.

Table 1. Elemental composition, elemental ratios and ash content of the humic acids

Sample	HA-HI	HA-HR	HA-LO	HA-LR	HA-LN
C*, mg.g ⁻¹	580 ±1.2	640 ±1.2	633 ±1.9	630 ±1.3	588 ±0.0
H*, mg.g ⁻¹	42 ±0.6	46 ±0.4	44 ±0.4	44 ±0.4	42 ±3.5
N*, mg.g ⁻¹	16 ±0.0	18 ±0.0	12 ±0.1	14 ±0.0	48 ±0.0
O**, mg.g ⁻¹	362 ±3.2	296 ±1.6	312 ±2.5	312 ±2.5	322 ±3.8
C/N	37	36	53	46	12
H/C***	0.88	0.86	0.83	0.84	0.86
O/C***	0.47	0.35	0.37	0.37	0.41
Ash, mg.g ⁻¹	36.0	24.0	5.0	59.0	92.0

* Calculated on an ash-free basis; ** calculated by difference; *** atomic ratios. HA-HI: high rank coal HA, HR: regenerated HA-HI; HA-LO: low rank coal HA; HA-LR: regenerated HA-LO; HA-LN: nitrated HA-LO

Infrared spectroscopy

The FTIR spectra of the isolated coal HA are shown in Figure 1. The spectra of the HA-HI and HA-HR samples were very similar and practically no changes due to the regeneration were detected. The most important features of those spectra are: a) a broad band around 3400 cm^{-1} due to O-H stretching of various groups like phenols and alcohol; b) a couple of peaks at 2922 and 2860 cm^{-1} and at 1430 and 1334 cm^{-1} due to C-H stretching; c) a well defined band at 1706 cm^{-1} due to C=O stretching of ketonic and carboxylic groups; d) a strong peak at 1615 cm^{-1} assigned to aromatic C=C stretching; e) a broad band around 1250 cm^{-1} assigned to C-O stretching of aryl esters and of OH deformation of COOH; f) a discrete peak at 1031 cm^{-1} due to Si-O stretching.

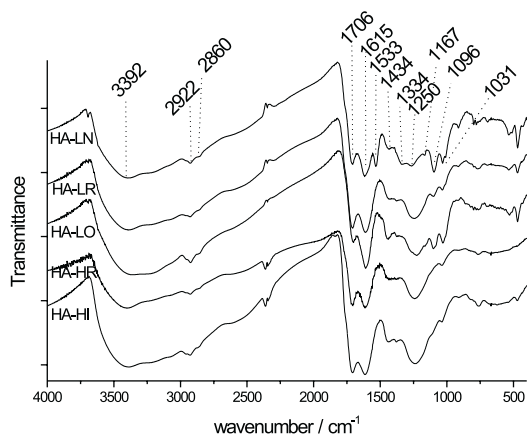


Figure 1. Infrared (FTIR) spectra of the humic acids

Opposite to the HA-HI, HA-HR and HA-LR spectra, the HA-LO sample presented more prominent peaks at 1615 and 1434 cm^{-1} (Figure 1) attributed to the presence of metal-coordinated carboxylate, which absorbs in this region (antisymmetrical and symmetrical COO^- stretching, respectively). This feature is probably more related to the lower effectiveness of the purification process rather than

to the regeneration or to the coal rank. Both low rank coal HA samples (HA-LO and HA-LR) showed a well-defined peak at 1096 cm^{-1} due to C-O stretching of ethers and phenols. This absorption band was also observed in the FTIR spectra of an Australian coal HA.¹⁰ A sharp absorption band at 1530 cm^{-1} due to NO_2 stretching observed in the sample HA-LN (Figure 1) is an indicative of the nitration reaction, corroborating the findings from the elemental analysis (Table 1). The intensity decrease of the peak at 1250 cm^{-1} suggests that nitration occurred at the expenses of the carboxylic groups, while the intensity increase of the band at 1096 cm^{-1} is probably due to the contribution of the bending of the C-N group, that also absorbs in this region.²⁴

^{13}C -NMR and EPR spectroscopy

The chemical shifts assignments of the ^{13}C -NMR-spectra and their relative abundance obtained with the MAS and with the CP/MAS technique are given in Table 2. The ^{13}C -NMR-MAS spectra are shown in Figure 2.

In comparison to the MAS, the CP/MAS technique intensified the alkyl-C signals and depleted the carboxylic-C and aromatic-C signals. This cross polarization effect is well reported in the literature and the quantitative interpretation of the ^{13}C -NMR-CP/MAS spectra of soil organic matter and humic substances should be taken with caution.^{16,21,25} Nevertheless, in the present study, the CP/MAS data showed the same tendency verified with the MAS data, and for comparison purposes between similar samples, the cross polarization technique (which is much faster) is usually considered adequate.^{16,21,25} In the following discussion of functional groups distribution only the NMR-MAS data obtained in this study will be considered.

All five HA presented a much higher aromatic and lower carboxylic contents than those obtained for an Australian brown coal HA⁸ and those usually reported for soil HA,^{1,21} but are in the range of the values verified for Spanish coal

Table 2. Main resonance regions of the ^{13}C -nuclear magnetic resonance spectra and its relative abundance (%), employing the MAS technique (^{13}C -NMR-MAS) and the CP/MAS technique (^{13}C -NMR-CP/MAS) of the humic acids

HA Sample	^{13}C -NMR-MAS				^{13}C -NMR-CP/MAS			
	Alkyl-C	O-alkyl-C	Aromatic-C	Carboxylic-C	Alkyl-C	O-alkyl-C	Aromatic-C	Carboxylic-C
HA- HI	13.9	2.7	75.8	7.6	32.5	1.0	63.7	2.8
HA-HR	10.3	3.5	78.3	7.9	24.2	1,1	72.4	2.3
HA-LO	9.5	6.1	77.3	7.1	19.2	9.0	69.7	2.1
HA-LR	10.3	6.1	76.3	7.3	24.4	3.7	69.4	2.5
HA-LN	14.7	3.3	80.2	1.8	34.8	1.7	62.1	1.4

Legend: see Table 1

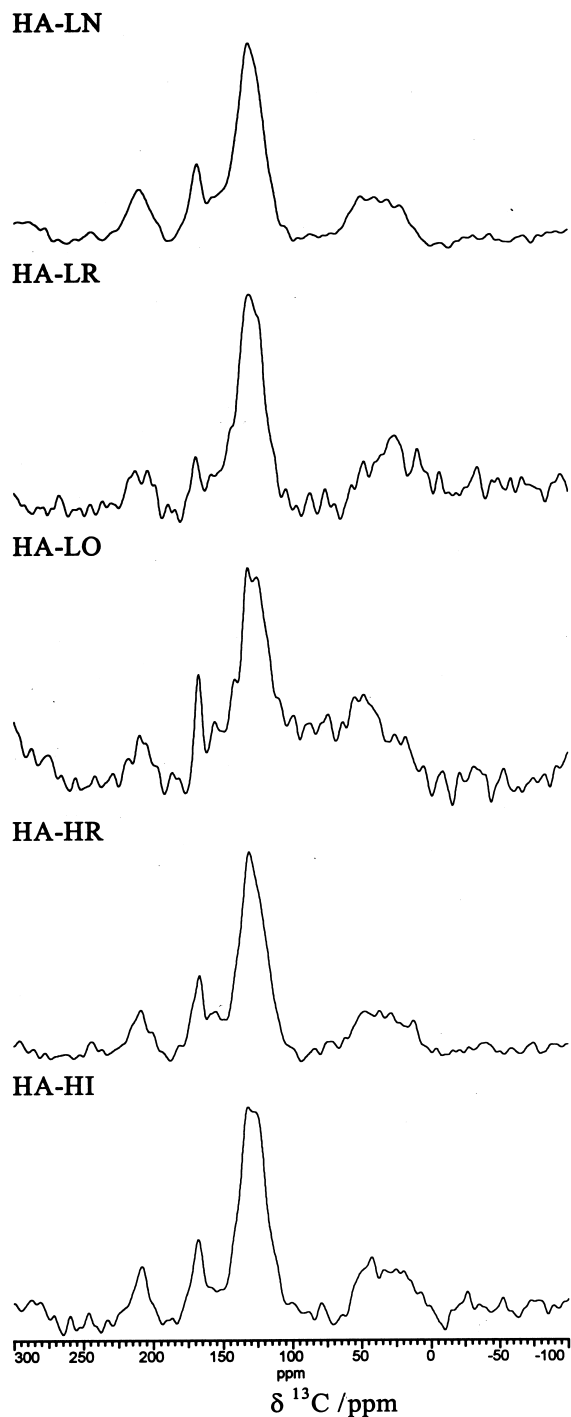


Figure 2. ^{13}C - nuclear magnetic resonance spectra (^{13}C -NMR-MAS) of the humic acids

HA.¹⁰ In the low rank samples (HA-LO and HA-LR) the abundance of O-alkyl-C groups was greater than that in the high rank coal HA, and this is consistent with the results from the FTIR analysis. This result may be associated to the preservation of carbohydrate-like structures in the low rank coal HA. Analogous to the infrared spectroscopy, no

changes in the chemical composition and structure of the humic acids due to coal regeneration in pile were evidenced by the NMR data. On the contrary, the nitration process of the low rank coal HA (HA-LR), produced an increase of the alkyl-C and of the aromatic-C, and a significant decrease of the carboxylic C=O and $\text{CH}_n\text{-O}$ groups.

The concentrations of free radicals as determined by EPR analysis were similar in the low and in the high rank coal HA (HA-LO and HA-HI) (Table 3) and higher than in the other HA samples. The values obtained in the HA from fresh coal samples are in the range of those already reported for soil HA^{19,20} and are indicative of a higher humification degree.²⁶ The decrease of the free radical concentration in the sample submitted to regeneration (HA-LR) may be due to semiquinone structures oxidation. The same reaction probably occurred by nitration of the sample, although in a lower extent than in the aged sample. The g-values, a typical EPR parameter for molecular structural study, oscillated between 2.0055 and 2.0058 and the line widths between 4.1 and 5.6 G. These g-values are more appropriated for 1,4- than for 1,2-semiquinone structures.²⁷

Table 3. Electron paramagnetic resonance (EPR) spectral data of humic acids

HA Sample	EPR data		
	Free radical, (spins $\times 10^{-17}\text{g}^{-1}$) [*]	Line widths (Gauss)	g-values
HA-HI	8.0	5.0	2.0056
HA-HR	n.m.	n.m.	n.m
HA-LO	7.8	5.6	2.0058
HA-LR	4.1	5.2	2.0056
HA-LN	6.7	4.1	2.0055

n.m. not measured

* Calculated on an ash-free basis. Legend: see Table 1

Conclusions

The analyzed coal humic acids presented a higher aromatic character, a lower carboxylic content and a lower C/N ratio than those usually observed for soil HA. These characteristics may limit the application of those two south Brazilian coals as chelating agents, fertilizer or soil conditioner.

The elemental composition obtained in the low rank HA pointed to more hydrophobic character in this sample when compared to the high rank HA. Those results, though, were not confirmed by the IR, ^{13}C -NMR and EPR data. It seems that the coal rank did not influence in a great extent the chemical and molecular characteristics of their humic acids. The regeneration process did not cause major

alterations in the coal HA, except in the free radical content as determined by EPR. The regeneration period and/or condition (in pile) was not sufficient to produce HA with chemical and molecular characteristics similar to those of soil HA. The nitration process achieved to reduce the C/N ratio of the HA to a value comparable to that obtained in soil HA, but the aromaticity still remained high and the carboxylic content was lowered after the procedure.

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References

1. Stevenson, F. J.; *Humus Chemistry. Genesis, Composition, Reactions*, John Wiley: New York, 1994.
2. Piccolo, A. In *Humic Substances in Terrestrial Ecosystems*; Piccolo, A. ed.; Elsevier: Amsterdam, 1996, p 225.
3. Piccolo, A.; Pietramallara, G.; Mbagwu, J.S.C.; *Geoderma* **1997**, *75*, 267.
4. Inbar, Y.; Chen, Y.; Hadar, Y.; *Soil Sci. Soc. Am. J.* **1990**, *54*, 1316.
5. Rumpel, C.; Knicker, H.; Kögel-Knabner, I.; Skjemstad, J.O.; Hüttl, R.F.; *Geoderma* **1998**, *86*, 123.
6. Mangrich, A.S.; Lobo, M.A.; Tanck, C.B.; Wypych, F.; Toledo, E.B.S.; Guimarães, E.; *J. Braz. Chem. Soc.* **2000**, *11*, 164.
7. Chen, Y. and Aviad, T. In *Humic Substances in Soil and Crop Sciences: Selected Readings*; McCarthy, P., Clapp, C. E., Malcolm, R.L., Bloom, P.R. eds.; Am. Soc. Agron., Soil Sci. Soc. Am.: Madison, 1990, p 161.
8. Pang, L.S.K.; Vassalo, A.M.; Wilson, M.A.; *Org. Geochem.* **1989**, *16*, 853.
9. Lawson, G.J.; Stewart, D. In *Humic Substances II: In Search of Structure*; Hayes, M.H.B., McCarthy, P., Malcolm, R.L., Swift, R.S. eds.; John Wiley and Sons: Chichester, USA, 1989, p 641.
10. Ibarra, J.V.; Juan, R.; *Fuel* **1985**, *64*, 650.
11. Hatcher, P. G.; *Energy and Fuels*, **1988**, *2*, 48.
12. Spark, K. M.; Well, J.D.; Johnson, B.B.; *Aust. J. Soil Res.* **1997**, *35*, 89.
13. Ibarra, J.V.; Orduna, P.; *Fuel* **1986**, *65*, 1012.
14. Stevenson, F.J.; Chen, Y.; *Soil Sci. Soc. Am. J.* **1991**, *55*, 1586.
15. Piccolo, A.; Pietramallara, G.; Bazzofi, P.; Pellegrini, S. In *Humic Substances in the Global Environment and Implications on Human Health*; Senesi, N. and Miano, T. M. eds.; Elsevier: Amsterdam, 1994, p 683.
16. Wilson, M.A.; *NMR Techniques and Applications in Geochemistry and Soil Chemistry*, Pergamon Press: Oxford, England, 1987.
17. Mbagwu, J.S.C.; Piccolo, A. In *The Role of Humic Substances in the Ecosystems and in the Environmental Protection*; Drodz, J., Gonet, S.S., Senesi, N., Weber, J. eds.; Polish Society of Humic Substances: Wroclaw, Poland, 1997, p 921.
18. Swift, R.S. In *Methods of Soil Analysis. Part 3. Chemical Methods*; Sparks, D. L. ed.; Soil Science Society of America: Madison, USA, 1996, p 1001.
19. Baranciková, G.; Senesi, N.; Brunetti, G.; *Geoderma* **1997**, *78*, 251.
20. Rivero, C.; Senesi, N.; Paolini, J.; D'Orazio V.D.; *Geoderma* **1998**, *81*, 227.
21. Kögel-Knabner, I.; *Geoderma* **1997**, *80*, 243.
22. Gonçalves, A.B.; Mangrich, A.S.; Zarbin, A.J.G.; *Synthetic Metals* **2000**, *114*, 119.
23. Dick, D.P.; Gomes, J.; Rosinha, P.B.; *Rev. Bras. Ci. Solo* **1998**, *22*, 603.
24. Silverstein, R.M.; Bassler, G. C.; Morrill, T.C.; *Identificação Espectrofotométrica de Compostos Orgânicos*, Editora Guanabara Koogan AS.: Rio de Janeiro, 1994, p 380.
25. Preston, C.M.; *Soil Sci.* **1996**, *161*, 144.
26. Senesi, N.; *Adv. Soil Sci.* **1990**, *14*, 77.
27. Pedersen, J.A.; *CRC Handbook of EPR Spectra from Quinones and Quinols*; CRC Press: Boca Raton, Florida, 1985, p 57.

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