

# Worldwide distribution, symptoms and diagnosis of the coinfections between malaria and arboviral diseases: a systematic review

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The coinfection between malaria (ML) and arboviral diseases represents a major global public health problem, particularly in tropical and subtropical countries. Despite its relevance, this topic is still insufficiently discussed in the current literature. Here, we aimed to investigate the worldwide distribution, symptoms, and diagnosis during coinfection between ML and arboviral diseases. We conducted a systematic review following the Preferred reporting items for systematic reviews and meta-analyses (PRISMA) statement and assessed the selection and eligibility criteria, created and diagrammed maps, and analysed major symptoms with 95% confidence intervals (CI) using prevalence ratio and effect size, also performing latent class analysis. A total of 85,485 studies were retrieved, of which 56 were included: 57.14% in Asia, 25% in Africa, 14.30% in South America, and 3.56% in Europe. A total of 746 individuals were reported to be coinfecting with *Plasmodium* and arbovirus. Concurrent ML, Dengue (DEN), Chikungunya (CHIK), and Zika (ZIK) patients are more likely to present headache and skin rash. Regarding diagnosis, 58,253 were made, of which 38,176 were positive (ML and at least one arboviral disease). The magnitude of these pathogens' coexistence points out the pressing need for improvements in public health policies towards diagnosis and prevention of both diseases, especially in endemic areas.

Key words: arbovirus infections - epidemiology - *Plasmodium* - vector borne diseases.

Arthropod-borne diseases (ABD) are among the major global health problems, responsible for more than 17% of all infectious diseases, and more than 700 thousand annual deaths in the world, with children being the most affected.<sup>(1)</sup> Between them, Malaria (ML), Dengue (DEN), Chikungunya (CHIK), Zika (ZIK) and Yellow fever (YF) are among the most significant ABD.<sup>(1,2,3)</sup>

Malaria caused approximately 247 million cases and 619 thousand deaths worldwide in 2021.<sup>(4)</sup> Among the species responsible for causing human ML, *Plasmodium falciparum* and *P. vivax* are responsible for the highest mortality and morbidity, respectively.<sup>(3,4)</sup> Meanwhile, approximately 2,8 million cases of DEN, 274 thousand of CHIK, more than 40 thousand of ZIK, and 203 of YF were reported in 2022.<sup>(1,2,3)</sup>

These diseases share similar symptoms with each other and also with other infectious and non-infectious diseases.<sup>(1-8)</sup> The most recurrent symptoms are febrile syndrome, myalgia, arthralgias, dizziness, vomiting, fatigue, anaemia, and headaches.<sup>(1,2,3,4,5)</sup> This unspecific symptomatology could lead to misdiagnosis, especially during coinfection. Regardless of this limitation, the importance of these coinfections has been reported.<sup>(5,6,7)</sup>

Despite the clinical presentation similarities, clinical management during ML requires the use of antimalarial drugs, while no treatment is available for the viruses. Usually, the most effective method of controlling viruses is through vaccination. However, there are no vaccines or drugs available for CHIKV and ZIKV, and clinicians rely on supportive therapy, while for YFV a vaccine is recommended solely in the case of populations living in endemic areas.<sup>(1,6,7,8,9)</sup> Furthermore, for DENV, two vaccines are available to the population: Dengvaxia (Sanofi Pasteur), since 2022,<sup>(10)</sup> and QDENGGA (Takeda), since 2023.<sup>(11)</sup>

A better understanding of the current knowledge about ML and arboviral diseases coinfection, encompassing their hotspots, diagnostic bottlenecks, and how it affects the patient's follow-up and clinical management, depends on the careful examination over the sum of all information available on the reported cases. Thus, the aim of this systematic review is to assess the worldwide distribution of coinfections between ML and four arboviral diseases, as well as to report whether mono and coinfections present differences in symptoms as well as diagnostic/screening methods.

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## MATERIALS AND METHODS

*Search strategy* - A systematic search was conducted following the Preferred reporting items for systematic reviews and meta-analyses (PRISMA) statement<sup>(12)</sup> to identify relevant studies on the frequency, worldwide distribution, main symptoms, and diagnostic/screening methods on ML and arboviral diseases (DEN, CHIK, ZIK e YF) coinfection. The search for articles was performed in the PubMed, Google Scholar, Science Direct, and Scientific Electronic Library Online (SciELO) databases for studies published up to August 30th, 2023.

The research question was structured in the PICO format, where P = Patients with ML and arboviral diseases; I = *Plasmodium* spp. and arbovirus coinfection (DENV, CHIKV, ZIKV, YFV); C = Patients without coinfection; and O = Frequency, worldwide distribution, symptoms and diagnostic/screening methods of coinfection reported. Thus, the following questions were formulated: Is there a high frequency and worldwide distribution of coinfection between ML and the arboviral diseases addressed in this review? If yes, which symptoms are more prevalent in coinfection compared with mono-infection cases? Also, is the most frequently used method for screening/diagnosis the one that is recommended by the World Health Organisation (WHO)?

The following search terms were used: (“Malaria” AND “Arbovirus infections”); (“Malaria” AND “Arbovirus infections” AND “Symptomatology”); (“Malaria” AND “Arbovirus infections” AND “Diagnostic techniques and Procedures”); (“Malaria” AND “Arbovirus infections” AND “Quick diagnosis units”); (“Malaria” AND “Arbovirus infections” AND “Prevalence”); (“Malaria” AND “Dengue” AND “Prevalence”); (“Malaria” AND “Yellow fever” AND “Prevalence”); (“Malaria” AND “Chikungunya fever” AND “Prevalence”); (“Malaria” AND “Zika virus infection” AND “Prevalence”); (Malaria” AND “Arbovirus infections” AND “frequency”); (“Malaria” AND “Arbovirus infections” AND “Frequency”); (“Malaria” AND “Dengue” AND “ Frequency”); (“Malaria” AND “Yellow fever” AND “ Frequency”); (“Malaria” AND “Chikungunya fever” AND “ Frequency”).

*Selection and eligibility criteria* - The titles and abstracts of all returned studies were assessed for suitability. Studies were selected if they met the following criteria: (1) Peer-reviewed articles published in journals with a description of the sample strategy and study design; (2) Studies that included cases of coinfection between ML and arboviral diseases; (3) Surveys performed containing symptomatology, methods of screening/diagnosis and a description of the pathogen species; (4) Studies that included demographic information (children and/or adults, continent/country of residence/frequency); (5) Studies published up to August, 30th 2023. Full texts of potentially relevant studies were further analysed for coinfection prevalence data. Retrospective analysis and case reports with full text availability and reporting data about all the potential coinfections were included in the study.

The present work excluded studies carried out in non-humans, reviews, letters, opinion pieces, grey literature, as well as studies that did not have elucidated outcomes. A reference manager, EndNote Software (Version x9), was

used to check and exclude duplicate articles. The risk of bias was assessed in each paper by two reviewers using three of the Joanna Briggs Institute’s (JBI) Critical Appraisals Checklists for Case-Report and Analytical Cross Sectional Studies.<sup>(13)</sup> Only papers considered to have a moderate score ( $\geq 50\%$ ) were included in this study.

*Data extraction* - The data extracted from the selected publications included: (i) Citation, (ii) Place/Continent where the study was carried out, (iii) Study design, (iv) Sample number, (v) Positive for coinfection, (vi) Age, (vii) Symptomatology, (viii) Diagnostic test, (ix) Remarks. All data were stored in Microsoft Excel<sup>®</sup> 2020, and checked by three researchers.

*Frequency and global distribution mapping* - The frequencies and distributions of ML and arboviral diseases coinfections were summed, plotted on openly available maps (<https://www.freeworldmaps.net>), and then diagrammed using Ibis Paint X software (Version 10.0.2).

*Data analysis* - The frequency of each symptom for the diseases mentioned in the studies included in this review (ML, DEN, ZIK, CHIK and YF, as well as coinfections) was analysed through cross-reference tables, using Microsoft Excel software (2020) as a tool. This aimed to define the most frequently mentioned symptomatology among individuals.

Subsequently, the symptoms were analysed using the Chi-Square and Fisher’s Exact tests, aiming to determine if there is a distinct symptoms’ profile during coinfection. The prevalence ratio (PR), and the respective confidence interval (CI), were calculated for the occurrence of symptoms, based on the control group. Effect size (ES) measures were calculated using Cohen’s W statistic,<sup>(14,15)</sup> which considers an effect to be insignificant for values less than 0.19, small effect for values between 0.20 and 0.49, medium effect for values between 0.50 and 0.79, large effect for values between 0.80 and 1.29, and very large effect for values equal to or greater than 1.30. All analyses were performed in R version 4.2.2, and the significance level adopted was 5%.<sup>(16)</sup>

Latent class analysis (LCA)<sup>(17,18,19)</sup> was performed to understand the profile of symptoms. This statistical procedure seeks to group individuals according to similar patterns of responses, forming with greater homogeneity of interest and greater interclass heterogeneity. The choice of class number was made by means of the following statistical estimator: Akaike information criterion (AIC) and Bayesian information criterion (BIC). To test the association of latent classes with the different types of infection, a multinomial logistic regression (OR) model and their respective 95% CI were estimated as a measure of effect.

## RESULTS

In the present systematic review, a total of 85,485 studies were identified, 97.45% of which were eliminated in the analysis of titles/abstracts along with duplicate articles, remaining 2,180 articles. These proceeded to the full reading stage, evaluation of selection, eligibility criteria, and risk of bias. With that, 56 articles were included in this study (87.66% were excluded; Fig. 1).

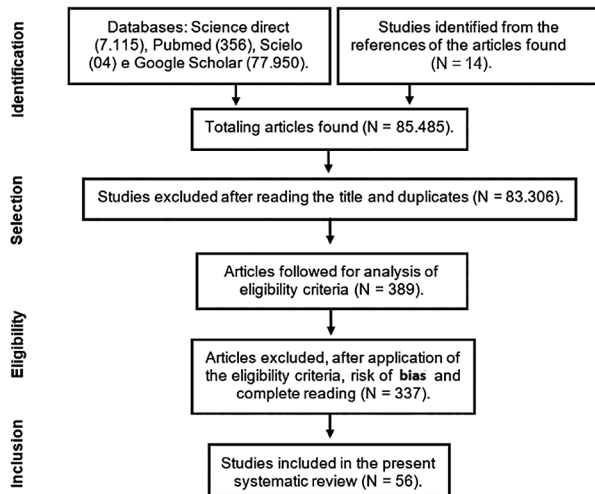


Fig. 1: flowchart of the selection of studies for the systematic review on the worldwide distribution, symptomatology, and diagnosis of coinfections between malaria (ML) and arboviral diseases.

Among the 56 included studies, 67.85% were observational analytical cross-sectional studies while the remaining 32.15% were case reports, all published between the years of 2005-2020. A total of 52,913 individuals were analysed from the included articles, 746 of them parasitised by *Plasmodium* spp. and at least one Arbovirus (Table I). These coinfections were: 656 ML/DEN [Supplementary data (Table I)], 58 ML/CHIK [Supplementary data (Table II)], 25 ML/ZIK, and 07 ML/YF [Supplementary data (Table III)]. Coinfections were detected in all age groups (according to the eligible data from 51 articles). However, there was a higher prevalence in young adults aged 28-30 years.<sup>(20-76)</sup>

As for the *Plasmodium* species, there was a predominance of *P. vivax*<sup>(26,28-29,31,36-38-41,43,45-46,50,53-54,61-62,64,70,73)</sup> with 3,483 monoinfected individuals, followed by *P. falciparum* with 1,557 cases, and the concomitant infection between *P. falciparum* and *P. vivax* with 175 cases. As for DEN, 124 individuals were positive for DENV-1,<sup>(36,39,45,48,56,58)</sup> 221 for DENV-2,<sup>(28,31,36-37,39,41,48,56)</sup> 611 for DENV-3,<sup>(36,38,41,45,56,58)</sup> and 368 for DENV-4.<sup>(36-37,39,41,55-56)</sup> Among the articles, three<sup>(21,35,45)</sup> reported one death, each from coinfection between ML and arboviral diseases. No *P. malariae*, *P. simium*, *P. cynolmolgi*, *P. inui* and arbovirus Oropouche and Mayaro coinfection reports were retrieved.

As for the worldwide frequency of coinfection between ML and arboviral diseases, fewer cases have been reported in Europe, Southeast Asia, and Eastern Africa (between 02 to 25 cases). However, in South America, Western and Central, the incidence of coinfection notifications increases, reaching over 130 cases. In relation to continental distribution, 57.14% were conducted in Asia, followed by 25% in Africa, 14.30% in South America, and 3.56% in Europe. No investigations answering the research questions were found in Oceania, North or Central America [Fig. 2 and Supplementary data (Figs 1,2,3,4)].

Among the 56 articles used to analyse the symptoms, 3.57%<sup>(40,49)</sup> were left out because they did not present symptoms as an inclusion criterion. Of the articles analysed, a total of 49,048 individuals participated in these studies and presented the symptoms for inclusion in our research.

A similar series of symptoms was observed in patients who had ML or an arboviral disease in single infections, as in those who were simultaneously monoinfected with any of the five pathogens (ML, DENV, CHIKV, ZIKV, YFV). The symptoms most commonly reported by coinfecting individuals were fever, headache, vomiting, tinnitus, abdominal pain, bleeding, and diarrhoea.

Table II describes the patients who had simultaneous infections for ML and arboviral diseases with a prevalence ratio and effect size between medium for the following symptom: Rash (PR: -, *p*-value: 0.000, ES: 0.506). The other selected symptoms showed non-significant (e.g.: Febrile syndrome (PR: 1, *p*-value: 0.000, ES: 0.022) or small results (e.g.: Nausea (PR: 10.51, *p*-value: 0.000, ES: 0.317).

In order to describe the clinical profiles, the LCA was performed (Fig. 3), gathering symptoms into three groups. In this analysis, the second group contained 85.71% of all individuals in the study. Thus, in both mono and coinfection cases, a predominance of febrile symptoms was observed. During the analysis of the association of latent classes with the different diseases (Table III), it was observed that the individual who was coinfecting with ML/arboviral diseases had a 12.49x chance of developing the symptoms present in Group 3 (Fig. 3).

Regarding diagnosis, 58,253 of them were performed by distinct methodologies. Of these, a total of 38,176 were positive (ML and/or some arboviral disease), 609 individuals were coinfecting, with seven (*n* = 7) of them presenting simultaneous infection for ML/DEN/CHIK.<sup>(22,26,63)</sup> The thick blood film was the main *Plasmodium* spp. diagnostic method used (*n* = 11,960), although molecular techniques were also observed. Overall, for DENV, the enzyme-linked immunosorbent assay (ELISA) for M immunoglobulin (ELISA IgM) and the ELISA for NS1 antigen (ELISA NS1) were more frequently chosen (5,894 and 4,516 tests, respectively). For CHIKV, the ELISA IgM (*n* = 238) and ELISA IgG (*n* = 208) were also used. The combined ELISA IgM/IgG test (*n* = 48) was the main choice for ZIKV antibody detection. Finally, for YFV the ELISA IgM test was more frequently applied (*n* = 11) (Table IV). The reverse transcription-polymerase chain reaction (RT-PCR) diagnosis was used in few studies.

## DISCUSSION

Diseases transmitted by vectors such as ML, DEN, CHIK, ZIK, and YF are of global epidemiological importance due to their mode of transmission, which involves a vector mosquito, humans, and the environment, characterising a one health problem.<sup>(1,2,3,4,5)</sup> In turn, vector species involved in the transmission of ML parasites and arboviruses have different habits and environmental conditions in the development of their biological cycle.<sup>(1,5,7)</sup> Therefore, it should be added that transmission may be related to the individual's place of residence, as well as their work activities, such as mining, deforestation, hunting, and fishing.<sup>(29,30,31)</sup>

TABLE I  
 Studies for the systematic review on the worldwide distribution, symptomatology, and diagnosis of coinfections between malaria (ML) and arboviral diseases (ABV)

Num Citation	Place/Continent	Study design	N	Positive for coinfection (%)	Demography	Symptomatology	Diagnostic test ML/ABV	Remarks
1 <sup>(38)</sup>	Pakistan, Asia	Analytical cross-sectional studies	114	22.80%	Ages between 13 and 70 years	Fever ≤ 10 days duration, severe body aches, rash, and bleeding.	Blood smear, ELISA (IgM/IgG)	<i>Plasmodium vivax</i> and <i>P. falciparum</i> , DENV (no serotype)
2 <sup>(29)</sup>	India, Asia	Analytical cross-sectional studies	223	4.03%	Ages between 22 and 56 years	Acute febrile illness (<2 weeks), associated with nausea, vomiting, and headache.	Blood smear, ELISA (IgM)	<i>Plasmodium vivax</i> and <i>P. falciparum</i> , DENV (no serotype)
3 <sup>(40)</sup>	India, Asia	Case report	1	100%	42 years	Fever, chills, rigor, altered sensorium (Cerebral malaria and dengue).	Blood smear, RDT, ELISA (NSI/IgM/IgG)	<i>Plasmodium falciparum</i> and DENV (no serotype)
4 <sup>(35)</sup>	Yemen, Asia	Analytical cross-sectional studies	270	30.37%	Ages between 15 and 60 years	Episodes of fever, headache, arthralgia, myalgia, and retro-orbital pain.	Blood Smear, RDT, ELISA (IgM/IgG)	<i>Plasmodium falciparum</i> and <i>P. vivax</i> , DENV (no serotype)
5 <sup>(31)</sup>	India, Asia	Case report	2	100%	35 and 63 years	Fever, chills, with vomiting and abdominal pain.	Blood smear, ELISA(NSI/IgM/IgG)	<i>Plasmodium vivax</i> and DENV-2, DENV-3
6 <sup>(28)</sup>	Pakistan, Asia	Analytical cross-sectional studies	856	1.99%	Ages between 12 and 32 years	Fever of 2-10 days, myalgia, arthralgia, retro-orbital pain.	Blood smear, RT-PCR, ELISA(NSI/IgM)	<i>Plasmodium vivax</i> and <i>P. falciparum</i> and DENV-2
7 <sup>(25)</sup>	Nigeria, Africa	Analytical cross-sectional studies	118	12.71%	All between 1 and 78 years	Fever > 37.5°C.	RDT, ELISA (IgM/IgG) Immunochromatography	<i>Plasmodium</i> sp. and ZIKV
8 <sup>(26)</sup>	Nigeria, Africa	Analytical cross-sectional studies	60	6%	Ages between 3 and 70 years	Fever, chills, headache, joint pain, muscle pain.	Blood smear, RDT, PCR, ELISA (NSI/IgM/IgG)	<i>Plasmodium vivax</i> and <i>P. falciparum</i> , DENV/CHIKV (no serotype)
9 <sup>(53)</sup>	India, Asia	Case report	3	100%	Between 8 months and 12 years	Fever for 5-8 days, cough, and body ache.	Blood smear, RDT ELISA(NSI, IgM/IgG)	<i>Plasmodium vivax</i> and DENV (no serotype)
10 <sup>(67)</sup>	French Guiana, South America	Analytical cross-sectional studies	1723	0.99%	NM	Episodes of fever lasting up to 4 days.	Blood smear, ELISA(IgM), RT-PCR, virus isolation	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and <i>P. malariae</i> , DENV-1, DENV-3
11 <sup>(74)</sup>	India, Asia	Case report	1	100%	28 years	Fever, chills in the last 7 days, abdominal pain, vomiting.	Blood smear, ELISA(IgM)	<i>Plasmodium falciparum</i> and DENV (no serotype)
12 <sup>(58)</sup>	France, Europe	Case report	1	100%	37 years	Fever, conjunctival jaundice, vomiting, diarrhoea.	Blood smear, IgM/IgG ELISA	<i>Plasmodium falciparum</i> and DENV-3
13 <sup>(42)</sup>	Malaysia, Asia	Case report	1	100%	59 years	Dyspnoea, chest discomfort, dry cough.	Blood smear, PCR, ELISA (NSI)	<i>Plasmodium knowlesi</i> and DENV (no serotype)
14 <sup>(24)</sup>	Malaysia, Asia	Case report	1	100%	59 years	Fever, headache, myalgia, arthralgia, and poor oral intake.	Blood smear, ELISA(NSI/IgM)	<i>Plasmodium knowlesi</i> and DENV (no serotype)





Num Citation	Place/Continent	Study design	N	Positive for coinfection (%)	Demography	Symptomatology	Diagnostic test ML/ABV	Remarks
15 (22)	Tanzania, Africa	Analytical cross-sectional studies	364	33 (9.06%)	Ages between 2 and 13 years	Fever, measured axillary or rectal temperature (37.5 or 38°C / 99.5 or 100.4°F).	Blood smear, ELISA(IgM/IgG), PCR	<i>Plasmodium</i> sp., and DENV/CHIKV (no serotype)
16 (43)	India, Asia.	Case report	1	1 (100%)	27 years	Myalgia (1 day before returning home to California from India after a 3 month period).	Blood smear, ELISA(IgM/IgG)	<i>Plasmodium vivax</i> and DENV (no serotype)
17 (39)	French Guiana, South America	Analytical cross-sectional studies	208	104 (50%)	Ages between 15 and 75 years	Episodes of fever > 40°C, tachycardia, initial hypotension, nausea.	Blood smear, RT-PCR, ELISA(NSI/IgM/IgA)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> , and DENV-1, DENV-2/ DENV-3
18 (40)	Bangladesh, Asia	Analytical cross-sectional studies	720	1 (0.14%)	4 years	Febrile patients > 38°C, headache, bodyaches, muscle pain.	RDT compatible with Blood smear, ELISA(IgM)	<i>Plasmodium vivax</i> and DENV (no serotype)
19 (46)	Peru, South American	Analytical cross-sectional studies	95	17 (17.89%)	Ages between 5 and 17 years	Fever, measured axillary > 37.5°C, abdominal pain, nausea, vomiting.	Blood Smear, PCR, ELISA (IgM/IgG), Immunofluorescence	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV-1, DENV-3
20 (32)	Nigeria, Africa	Analytical cross-sectional studies	340	2 (0.59%)	All ages	Febrile complaints (temperature > 37.5°C / 99.5°F).	Blood Smear, ELISA(IgM)	<i>Plasmodium</i> sp., and DENV (no serotype)
21 (23)	Thailand, Asia	Case report	1	1 (100%)	11 years	Fever, chills.	Blood smear, ELISA (NSI/ IgM/IgG)	<i>Plasmodium falciparum</i> and DENV (no serotype)
22 (58)	Cambodia, Asia	Analytical cross-sectional studies	9997	15 (0.15%)	Ages between 8 and 17 years	Fever in the last 24 hours and for < 10 days, muscle pain.	Blood Smear, ELISA (IgM/IgG), Nested PCR	<i>Plasmodium falciparum</i> , <i>P. vivax</i> and DENV-1, DENV-2, DENV-3, DENV-4
23 (46)	India, Asia	Case report	1	1 (100%)	26 years	Fever, headache, severe body pain and nausea for 10 days, chills every other day.	Blood smear, ELISA (IgM/IgG)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)
24 (27)	Tanzania, Africa	Analytical cross-sectional studies	400	8 (2.0%)	Ages between 10 and 50 years	Fever > 38°C, headache, skin rashes, joint pain.	Blood smear, RDT, ELISA (IgM/ IgG)	<i>Plasmodium</i> sp. and CHIKV (no serotype)
25 (60)	Nigeria, Africa	Analytical cross-sectional studies	176	5 (2.84%)	Ages between 10 and 70 years	Febrile illness.	RDT, ELISA (IgM ELISA) RT-PCR	<i>Plasmodium</i> sp., and DENV (no serotype)
26 (49)	Brazil, South America	Case report	1	1 (100%)	52 years	Chills, fever, headache, arthralgia, myalgia, choluria.	Blood smear, RDT, PCR, ELISA (IgM/ IgG/ NSI)	<i>Plasmodium ovale</i> and DENV (no serotype)
27 (41)	Brazil, South America	Analytical cross-sectional studies	132	11 (8.33%)	Ages between 16 and 92 years	Febrile, chills, myalgias, arthralgias, headache.	Blood smear, RT-PCR, ELISA(NSI)	<i>Plasmodium vivax</i> and DENV-3, DENV-4
28 (37)	Brazil, South America	Analytical cross-sectional studies	1578	44 (2.79%)	Ages between 14 and 60 years	Episodes of fever in the past 10 days.	Blood smear, RT-PCR, ELISA (IgM/NSI)	<i>Plasmodium vivax</i> and DENV-2, DENV-4



Num Citation	Place/Continent	Study design	N	Positive for coinfection (%)	Demography	Symptomatology	Diagnostic test ML/ABV	Remarks	
29 <sup>(50)</sup>	Thailand (Burmese border), Asia	Analytical cross-sectional studies	203	1	0.49%	Ages between 15 and 41 years	Febrile episodes up to 3 days (aural temperature 37.5°C), headache, anorexia, muscle pain.	Blood smear, ELISA (IgM/NSI)	<i>Plasmodium falciparum</i> , <i>P. vivax</i> and DENV (no serotype)
30 <sup>(36)</sup>	Brazil, South America	Analytical cross-sectional studies	72	30	41.67%	Ages between 20 and 44 years	Acute febrile syndrome.	Blood smear, PCR/RT-PCR, ELISA (IgM/NSI)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV-1, DENV-2, DENV-3, DENV-4
31 <sup>(44)</sup>	India, Asia	Analytical cross-sectional studies	2547	11	0.43%	≥ 18 years	Febrile illness with duration of 5-14 days, rash, hepatomegaly and abdominal pain.	Blood smear, RDT, ELISA (NSI/IgM)	<i>Plasmodium</i> sp., and DENV (no serotype)
32 <sup>(61)</sup>	India, Asia	Analytical cross-sectional studies	469	27	5.76%	NM	Fever for < 7 days, running nose, myalgia, headache, and bleeding manifestations.	Blood smear, ELISA (IgM/NSI)	<i>Plasmodium falciparum</i> , <i>P. vivax</i> and DENV (no serotype)
33 <sup>(62)</sup>	India, Asia	Analytical cross-sectional studies	1564	78	4.98%	≥ 5 years	Fever temperature ≥ 38°C (100.4°F) and febrile illness with duration of 2-14 days.	Blood smear, ELISA (IgM/IgG/NSI), and Blood cultures.	<i>Plasmodium falciparum</i> and DENV/CHIK (no serotype)
34 <sup>(63)</sup>	Cambodia, Asian	Analytical cross-sectional studies	1193	27	2.26%	Ages between 7 and 49 years	Febrile illness (> 38°C), sore throat, cough, and running nose.	RDT, Nested-PCR, and RT-PCR.	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)
35 <sup>(64)</sup>	Mozambique Africa	Analytical cross-sectional studies	163	2	1.23%	Ages between 5 and > 40 years	Acute febrile illness > 37.5°C, headache, arthralgia, myalgia.	RDT, ELISA (IgM/IgG/NSI), qRT-PCR	<i>Plasmodium falciparum</i> and CHIKV (no serotype)
36 <sup>(54)</sup>	India, Asia	Case report	1	1	100%	25 years	Fever, chills, myalgias, headache, severe headache, and high fever of 102°F (38.9°C).	Malaria Ag (pLDH/HRP2), Blood smear, ELISA (IgM)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)
37 <sup>(76)</sup>	Tanzania, Africa	Analytical cross-sectional studies	448	13	2.90%	All ages between 2 and 70 years	Participants without complaints, randomly selected.	Blood Smear, ELISA (IgM)	<i>Plasmodium</i> sp. and CHIKV (no serotype)
38 <sup>(66)</sup>	Cameroon, African	Analytical cross-sectional studies	349	68	19.48%	Ages between 6 months to 15 years	Children presenting episodes of fever (37.8-41°C, or 100.04-105.8°F), vomiting, diarrhoea, and fatigue.	Blood Smear, ELISA (NSI/IgM/IgG)	<i>Plasmodium falciparum</i> , <i>P. vivax</i> and DENV (no serotype)
39 <sup>(67)</sup>	Nigeria, Africa	Analytical cross-sectional studies	529	35	6.62%	Ages between < 18 and 58 years	Episodes of fever (axillary temperature > 37.8°C/100.04°F).	Blood smear, ELISA (IgM/IgG/NSI)	<i>Plasmodium falciparum</i> and DENV (no serotype)
40 <sup>(56)</sup>	Nigerian, Africa	Analytical cross-sectional studies	188	19	10.11%	Ages between 4 and 82 years	Acute fever (> 38°C)	NM ELISA (IgG/IgM, NSI)	<i>Plasmodium</i> sp., and DENV (no serotype)
41 <sup>(20)</sup>	India, Asia	Case report	1	1	100%	25 years	Fever (temperature = 101°F, or 38.3°C), dyspnoea, erythematous rash.	Blood smear, ELISA (NSI/IgG/IgM)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)



Num Citation	Place/Continent	Study design	N	Positive for coinfection (%)	Demography	Symptomatology	Diagnostic test ML/ABV	Remarks	
42 <sup>(75)</sup>	India, Asia	Analytical cross-sectional studies	100	3	3.0%	Ages between 5 and ≥ 60 years	Fever, abdominal pain and bleeding.	Blood smear, RDT/ELISA	<i>Plasmodium</i> sp., and DENV (no serotype)
43 <sup>(66)</sup>	India, Asia	Analytical cross-sectional studies	1980	22	1.11%	Ages between 5 and > 15 years	Febrile illness (38.3–39.4°C / 100.94–102.92°F), headache, retro-orbital pain, fever for 2–15 days.	Blood smear, RDT, ELISA (IgM/ IgG, NSI) RT-PCR	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)
44 <sup>(47)</sup>	India, Asia	Case report	1	1	100%	17 years	Fever, chills in the last 5–6 days, abdominal pain, and vomiting.	RMAT, PCR, ELISA (IgM)	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV (no serotype)
45 <sup>(48)</sup>	Brazil, South America	Analytical cross-sectional studies	111	2	1.80%	Ages > 18 years	Episodes of fever, headache, and shivering.	Blood smear, RT-PCR, Nested-PCR	<i>Plasmodium vivax</i> , <i>P. falciparum</i> and DENV-1
46 <sup>(33)</sup>	Madagascar, Africa	Analytical cross-sectional studies	1216	2	0.16%	Pregnant women, all ages	NM	ELISA (IgG/IgM), IIFA, PCR	<i>Plasmodium falciparum</i> and ZIKV
47 <sup>(59)</sup>	Spain, Europe	Case report	1	1	100%	27 years	Fever, constipation, and joint pain.	Blood smear, RT-PCR, ELISA (IgM/ IgG/NSI)	<i>Plasmodium falciparum</i> and DENV-4
48 <sup>(70)</sup>	India, Asia	Analytical cross-sectional studies	8364	27	0.32%	NM	Fever compatible with malaria and/or dengue.	Blood smear, ELISA (NSI/ IgM)	<i>Plasmodium falciparum</i> and DENV (no serotype)
49 <sup>(71)</sup>	India, Asia	Analytical cross-sectional studies	1141	9	0.79%	Ages between 12 and 80 years	Acute febrile illness.	Blood smear, ELISA (IgM, NSI)	<i>Plasmodium</i> sp., and DENV (no serotype)
50 <sup>(6)</sup>	Senegal, Africa	Analytical cross-sectional studies	13845	19	0.13	Ages between 1 and 90 years	Acute febrile illnesses (> 38 °C), headache, myalgia, eye pain, arthralgia.	Blood smear, RDT, ELISA (IgM), RT-PCR	<i>Plasmodium</i> sp., and DENV/CHIKV/ ZIKV/YFV (no serotype)
51 <sup>(52)</sup>	Ghana, Africa	Analytical cross-sectional studies	218	7	3.21%	Ages between 2 and 14 years	Febrile illness, headache, nausea, chills.	RDT, ELISA (IgM/ IgG), RT-PCR	<i>Plasmodium</i> sp., and DENV (no serotype)
52 <sup>(72)</sup>	Bangladesh, Asia	Analytical cross-sectional studies	659	5	0.76%	Ages between 0 and 90 years	Fever > 37.5°C, fatigue, fever, dizziness and headache.	RDT, PCR, Blood smear, ELISA (IgM)	<i>Plasmodium falciparum</i> , <i>P. vivax</i> , <i>P. malariae</i> and DENV (no serotype)
53 <sup>(73)</sup>	India, Asia	Case report	1	1	100%	22 years	Fever > 39° C, chills, rigors, cough up to 3 days.	Blood smear, ELISA (IgM)	<i>Plasmodium vivax</i> and DENV (no serotype)
54 <sup>(21)</sup>	East Timor, Asia	Case report	1	1	100%	7 years	Fever, headache, fatigue, anorexia.	Blood Smear, RDT, ELISA (IgM)	<i>Plasmodium falciparum</i> and DENV (no serotype)
55 <sup>(34)</sup>	Pakistan, Asia	Analytical cross-sectional studies	159	5	3.74%	Ages > 12 years	Acute febrile illness and found to have thrombocytopenia.	Blood smear, IgM	<i>Plasmodium falciparum</i> and DENV (no serotype)
56 <sup>(51)</sup>	Indonesia, Asia	Case report	1	1	100%	49 years	Fever, chills, rigors, myalgia.	Blood smear, ELISA (IgM/ NSI)	<i>Plasmodium falciparum</i> and DENV (no serotype)

ELISA: enzyme-linked immunosorbent assay; PCR: polymerase chain reaction; qRT-PCR: reverse transcription real-time PCR; RDT: rapid diagnostic test; CHIK: Chikungunya; DENV: Dengue virus; YFV: Yellow fever virus; ZIKV: Zika virus.

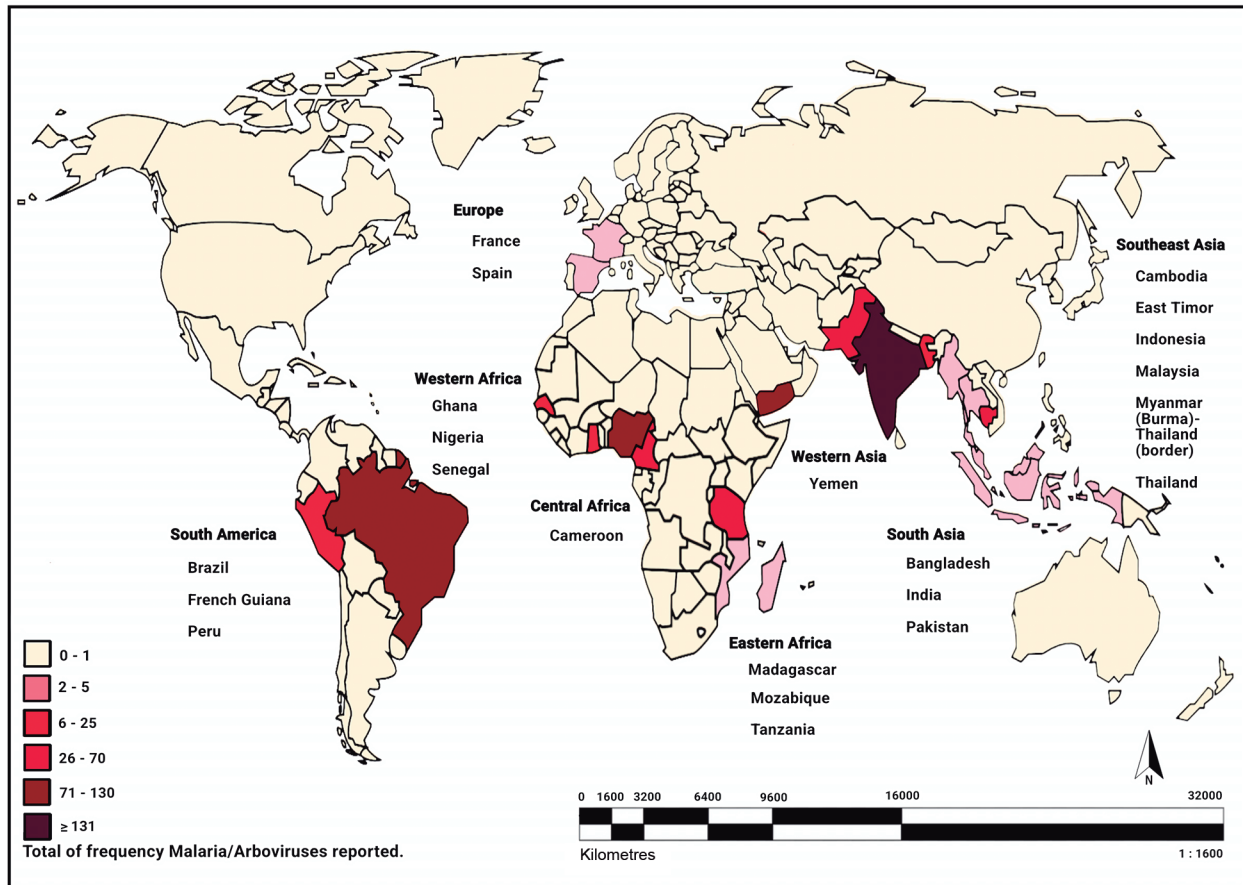


Fig. 2: worldwide frequency and distribution map of malaria (ML) and arboviruses coinfections, according to the studies in this systematic review.

As expected, our results demonstrate that the majority of studies on these diseases come from countries located in tropical and subtropical regions of Asia, Africa, and the Americas, respectively. In these continents, hot and rainy weather form water collections, unplanned urban development, poor sanitation, rampant deforestation, and intense population flow between endemic and non-endemic regions of these pathologies.<sup>(6,20)</sup> In such a scenario, the adaptation of the microorganism to the vector and environmental conditions favours the proliferation of *Anopheles* spp. and *Aedes* spp., as well as the development of these diseases, also as coinfections.<sup>(22,31,71-85)</sup>

ML, DEN, CHIK, ZIK, and YF are diseases that were first reported in Africa, representing the second continent with the highest number of reported coinfection cases in this review. This is likely due to the fact that Asia presents an overpopulation, leading to more cases. However, there may be underreporting in the African continent, as a significant portion of its population lives in war-torn regions, which also contributes to difficulties in healthcare access.<sup>(25,33,73,74)</sup>

America, the third continent with the highest number of coinfections, presents similar numbers of mono and coinfections compared to Africa. Presumably, the aforementioned information is somehow interconnected, since their vulnerable population faces environmental and socioeconomic challenges, leading to an increase in

infectious disease burden.<sup>(25-33)</sup> In this review, only two European studies described coinfection cases, which happened during trips to Central America and to Africa. The cold climate hinders the spread of vector mosquitoes along with the advantageous socio-economic conditions and effective prophylactic measures, compared to those observed in the African, Asian, and American continents. These are responsible for the lower frequency of coinfections.<sup>(58-61)</sup>

Coinfection between ML and DEN was the most common, followed by ML+CHIK, ML+ZIK, and ML+YF. These data reflect the pathogens' ability to complete their biological cycle in the vector and the vectorial competence to transmit these arboviral diseases, with a better shape in the host-parasite relationship for concurrent infection between ML+DEN.<sup>(28,29,30,31)</sup> This scenario is likely to change in view of the promising DENV vaccines, which are already in the testing/implementation phase.<sup>(10,11,86-88)</sup> In contrast, the detection of seven cases of coinfection between ML and YF, the only arboviral disease that currently has an effective vaccine, reflects the global vaccination policy.<sup>(28-34)</sup>

In Brazil, South America, the main vectors of ML and DEN, *Anopheles darlingi* and *Aedes aegypti*, respectively, have different behaviours, the former more associated with rural areas and the latter with urban areas. However, nowadays, it has been observed that *A.*



TABLE II  
Prevalence ratio (PR) and effect size (ES) of symptoms that *Plasmodium*/arboviral diseases coinfecting individuals may develop

Symptoms	N (%)	PR	p-value	ES
Headache				
Malaria	2147 (17.6)	3.88 (3.63, 4.14)	0.000	0.440
Arbovirus	3615 (46.9)	10.32 (9.74, 10.94)		
Coinfection	413 (56.2)	12.37 (11.39,13.44)		
Control	1323 (4.5)	1		
Vomiting				
Malaria	1934 (15.9)	4.76 (4.42, 5.13)	0.000	0.385
Arbovirus	2690 (34.9)	10.48 (9.78, 11.22)		
Coinfection	413 (56.2)	16.87 (15.44, 18.44)		
Control	970 (3.3)	1		
Nausea				
Malaria	1620 (13.3)	4.02 (3.72, 4.34)	0.000	0.317
Arbovirus	2215 (28.7)	8.7 (8.1, 9.34)		
Coinfection	255 (34.7)	10.51 (9.35, 11.81)		
Control	962 (3.3)	1		
Abdomen pain				
Malaria	1399 (11.5)	417.58 (208.49,836.39)	0.000	0.364
Arbovirus	1702 (22.1)	803.71 (401.46, 1608.99)		
Coinfection	290 (39.5)	1436.68 (714.42, 2889.14)		
Control	8 (0)	1		
Joint pain				
Malaria	881 (7.2)	NA	0.000	0.336
Arbovirus	1610 (20.9)	NA		
Coinfection	79 (10.7)	NA		
Control	0 (0)	1		
Arthralgia				
Malaria	533 (4.4)	3.53 (3.09, 4.02)	0.000	0.269
Arbovirus	1293 (16.8)	13.53 (12.07, 15.16)		
Coinfection	128 (17.4)	14.05 (11.65, 16.96)		
Control	361 (1.2)	1		
Rash				
Malaria	791 (6.5)	NA	0.000	0.506
Arbovirus	2974 (38.6)	NA		
Coinfection	183 (24.9)	NA		
Control	0 (0)	1		
Chills				
Malaria	381 (3.1)	NA	0.000	0.229
Arbovirus	322 (4.2)	NA		
Coinfection	158 (21.5)	NA		
Control	0 (0)	1		
Myalgia				
Malaria	687 (5.6)	1.24 (1.13, 1.36)	0.000	0.225
Arbovirus	1507 (19.5)	4.3 (4.01, 4.61)		
Coinfection	212 (28.8)	6.35 (5.6, 7.2)		
Control	1323 (4.5)	1		
Diarrhea				
Malaria	1642 (13.5)	NA	0.000	0.397
Arbovirus	2117 (27.5)	NA		
Coinfection	290 (39.5)	NA		
Control	0 (0)	1		
Febrile syndrome				
Malaria	12199 (100)	1 (1, 1)	0.000	0.022
Arbovirus	7711 (100)	1 (1, 1)		
Coinfection	735 (100)	1 (1, 1)		
Control	29096 (99.9)	1		

NA: not applicable.

*darlingi* due to anthropological actions, such as the construction of hydroelectric plants, illegal mining, unlawful deforestation, and urbanisation, has changed its behaviour, causing urban ML, especially in the Brazilian Amazon. In this way, these two vectors end up coexisting in the same space, which can increase the risk of infection, especially by *Plasmodium* spp. + DENV, which accounts for the majority of coinfection cases. This fact raises new challenges for public health in the control of both diseases, since, in this country, the surveillance of ML and DEN is usually developed separately.<sup>(37,38,42,48,49)</sup>

It is important to note that, in this systematic review, coinfection was observed between ML and various arboviral diseases (DEN, CHIK, ZIK, and YF). However, no reports were presented regarding the detection of cases by *Plasmodium* spp. and the Mayaro and Oropouche viruses. This fact is likely due to the absence of routine laboratory diagnosis of these two arboviruses in the studied areas.<sup>(22-26)</sup> We emphasise the need for the investigation of arboviruses, specifically the Mayaro and Oropouche viruses, to be implemented in routine diagnosis, given reports of their co-circulation in ML-endemic regions.<sup>(85,86)</sup>

*Symptomatology: aspects that lead to similarity in the clinical manifestation of mono and coinfections between ML and arboviral diseases - Plasmodium* spp. and the arbovirus investigated here are challenged to thrive in spite of the immunological factors involved in the host-parasite interaction, which are intrinsically permeated by the human host genetic variability.<sup>(32-80)</sup> In spite of all this variety and its consequent possible outcomes, intriguingly, we could not observe major symptoms and/or clinical manifestations to be named as pathognomonic in patients with concurrent ML and arboviral diseases. As a matter of fact, in such infectious diseases, the pro-inflammatory cytokine cascades, during coinfections, play a crucial role in the severity of symptoms.<sup>(24,25,26,27)</sup> Likewise, undifferentiated febrile syndrome, neurological symptoms, joint pain, and anaemia were constantly recorded for the majority of all cases.

Nevertheless, this systematic review brings to light symptoms which can be considered of attention to healthcare providers working in endemic areas for ABD. The first of these was that concurrent ML, DEN, CHIK, and ZIK patients are more susceptible to presenting headache and skin rash. Therefore, an important public health measure could be to implement *Plasmodium* spp. investigation whenever skin rash and headache appear as symptoms in an arbovirus-infected patient, at least in the endemic area of both diseases, when these diseases coincide spatially and affect the same population groups. Secondly, when the three clinical aspects are assessed (febrile syndrome, bleeding, and thrombocytopenia), the probability of concurrent ML and DEN is observed.<sup>(50-60)</sup> Undoubtedly, this triad can worsen the clinical condition, especially in immunocompromised individuals, pregnant women, and children.<sup>(55-60)</sup>

Anaemia reported in individuals infected with *Plasmodium* spp. has multifactorial origins. One of the studied factors may be related to coinfection with arbovirus-

TABLE III

Multinomial logistic regression, with the odds ratio (OR) as the measure of effect, and their respective 95% confidence intervals (CI), regarding patient-reported symptoms in this systematic review

Variables	Group 1		Group 3	
	OR	CI 95%	OR	CI 95%
Malaria	14.95	55.64 - 85.53	2.14	1.97 - 2.33
Arbovirus	16.32	54.27 - 86.91	6.01	5.54 - 6.53
Coinfection	16.84	53.75 - 87.42	14.92	12.49 - 17.82
Control	1		1	

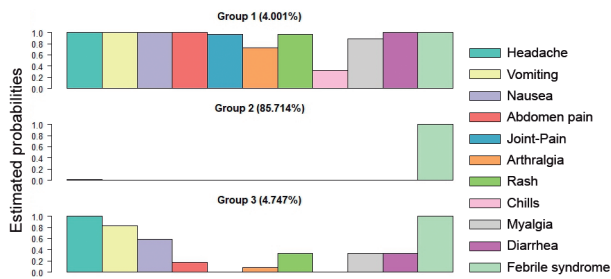


Fig. 3: latent class analysis (LCA). The choice of class number was made using the statistics Akaike information criterion (AIC) and Bayesian information criterion (BIC), according to the studies in this systematic review.

es, especially DEN.<sup>(20-24,37-40)</sup> Haemorrhages associated with febrile syndrome can worsen the clinical picture, making therapeutic approaches by healthcare professionals challenging. Added to this concern, severe DEN associated with simultaneous ML infection becomes a problem, as some of the antimalarial drugs cannot be prescribed to pregnant women, neonates, and glucose-6-phosphate dehydrogenase deficiency carriers.<sup>(32,33,34)</sup>

*Diagnosing coinfection between ML and arboviral diseases: a lasting issue* - Delaying the identification of the cause of the illness in endemic areas of *Plasmodium* and arbovirus can lead to clinical management uncertainty or even critical misconduct. One of its complex consequences is returning the patient to its community in the condition of a constant source of transmission.<sup>(38-50)</sup> Therefore, there is a need for the implementation of diagnostic strategies for these microorganisms, their vectors, and also their environmental conditions to develop effective prevention and control measures, as recommended by the WHO and Pan American Health Organisation (PAHO).<sup>(89,90)</sup>

Thick blood film is the gold standard of the ML diagnosis, as verified. However, this technique has limitations, as it can result in false negatives, especially in cases of low parasitaemia and mixed infections. Additionally, it requires the expertise of a microscopist to identify the species, its stage, as well as atypical forms of *Plasmodium* spp.<sup>(83)</sup> The rapid diagnostic test (RDT) is the second laboratory protocol option for ML detection found in this

TABLE IV

Types of diagnoses and frequency used to identify malaria (ML)/arboviral disease (ABV) of the studies in this systematic review

Groups	Types of diagnostic tests / frequency (%)											TOTAL			
	BS	RDT	PCR	RT-PCR	Nested PCR	ELISA (NSI)	ELISA (IGM)	ELISA (IGG)	ELISA (IGM/IGG)	Blood culture	Virus isolation	NT	P (n)	TPCa	%
ML	RP 11.960	RP 89.43	RP 9.499	RP 71.03	RP 7.57	RP 905	RP 87	RP 57.77	RP 71.03	RP 7.57	RP 905	RP 87	RP 57.77	RP 71.03	RP 7.57
ML/ABV	RP 698	RP 93.57	RP 431	RP 1.033	RP 13.87	RP 14	RP 227	RP 3.26	RP 1.206	RP 17.34	RP 845	RP 12.15	RP 4.516	RP 64.91	RP 5.894
ABV	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA
DENV	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA
CHIKV	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA
ZIKV	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA
YFV	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA	RP NA
TOTAL	RP 13,374	RP 22,989	RP 171.89	RP 44	RP 0.33	RP 13,374	RP 22,989	RP 171.89	RP 44	RP 0.33	RP 13,374	RP 22,989	RP 171.89	RP 44	RP 0.33

BS: blood smear; RDT: rapid diagnostic tests; PCR: polymerase chain reaction; RT-PCR: reverse-transcription polymerase chain reaction; nested PCR: nested polymerase chain reaction; ELISA (NSI): enzyme-linked immunosorbent assay for NSI antigen; ELISA (IgM): enzyme-linked immunosorbent assay for M immunoglobulin; ELISA (IgG): enzyme-linked immunosorbent assay for G immunoglobulin; ELISA (IgM/IgG): enzyme-linked immunosorbent assay for M and G immunoglobulins; NT: analysis type not mentioned; RP: reported positives; P (n): number of individuals with positive results; TPCa: total positive case analysis; DENV: Dengue virus; CHIKV: Chikungunya virus; ZIKV: Zika virus; YFV: Yellow fever virus; NA: not applicable.

review, being the most widely used test in remote areas.<sup>(62-66,81)</sup> However, it has limitations such as cost and inability to quantify parasitaemia and demonstrate the parasitic stage.<sup>(36,40)</sup> It is worth noting that polymorphisms affecting the expression of the rapid test recognition parasite protein have been observed, leading to false negative results. Diagnostic serology is not efficient for ML, and molecular biology techniques still have high operational costs, limiting their use in routine diagnosis.<sup>(60,61,62,63,72)</sup>

This systematic review showed that the IgM/IgG ELISA was the most commonly used diagnostic protocol for detecting arboviruses. However, these serological tests do not define current infection but only indicate their circulation in endemic regions.<sup>(50,51,52,53)</sup> Most articles describe that arbovirus investigation typically follows *Plasmodium* spp. Investigation.<sup>(60-65)</sup> The DENV NS1 antigen detection test by ELISA was also used, as well as the RT-PCR for all other arboviruses.<sup>(62-67)</sup> However, these protocols were not observed in the majority of studies. The limitations of serological tests in detecting antibodies against arboviruses should be considered, as seroconversion takes an average of 6-10 days.<sup>(76-80,84,85)</sup>

*In conclusion* - Coinfection and co-circulation between *Plasmodium* spp. and arbovirus are predominantly found in tropical and subtropical countries, where socio-environmental-sanitary conditions favour transmission. The review of vaccination programmes against YF is crucial in controlling this arbovirus. Protocols related to symptoms and diagnosis need to be redefined to distinguish coinfection from co-circulation, requiring molecular tests. Therefore, the current scenario of coinfection between ML and arboviral diseases still needs more extensive study, calling for efficient public health policies and investment in health education. The ultimate goal is to mitigate these diseases and improve the quality of life for the population.

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#### AUTHORS' CONTRIBUTION

MCF, ARSB, LMST and RLDM - Conception and design of the study, acquisition of data, analysis and interpretation of data, preparation of the manuscript, revision, and final approval of the submitted version; MLD and ESM - acquisition of data, analysis and interpretation of data, preparation of the manuscript, revision, and final approval of the submitted version; MCSJ, MPS, NFR and JRSS - analysis and interpretation of data, revision, and final approval of the submitted version. The authors declare that they have no competing interests, and confirm that all data underlying the findings are fully available without restriction. The funders had no participation in the study's design, data collection and analysis, publishing decision, or manuscript preparation.

#### REFERENCES

1. WHO - World Health Organization. Vector-borne diseases. 2020. Available from: <https://www.who.int/news-room/fact-sheets/detail/vector-borne-diseases>. [accessed 01 December 2022].
2. PAHO - Pan American Health Organization. Arbovirus Bulletin 2022. Epidemiological update for Dengue, Chikungunya and Zika in 2022. 2022. Available from: <https://www3.paho.org/data/index.php/en/mnu-topics/indicadores-dengue-en/annual-arbovirus-bulletin-2022.html>. [accessed 01 December 2023].
3. Mordecai EA, Caldwell JM, Grossman MK, Lippi CA, Johnson LR, Neira M, et al. Thermal biology of mosquito-borne disease. *Ecol Lett*. 2019; 22(10): 1690-1708. doi: 10.1111/ele.13335.
4. WHO - World Health Organization. World Malaria Report 2022. Geneva: World Health Organization; 2022. Available from <https://www.who.int/teams/global-malaria-programme>. [accessed 8 December 2023].
5. Githeko AK, Lindsay SW, Confalonieri UE, Patz JA. Climate change and vector-borne diseases: a regional analysis. *Bull World Health Organ*. 2000; 78(9): 1136-47.
6. Sow A, Loucoubar C, Diallo D, Faye O, Ndiaye Y, Senghor CS, et al. Concurrent malaria and arbovirus infections in Kedougou, southeastern Senegal. *Malar J*. 2016; 28(15): 47. doi: 10.1186/s12936-016-1100-5.
7. Chiuya T, Villinger J, Falzon LC, Alumasa L, Amany F, Bastos ADS, et al. Molecular screening reveals non-uniform malaria transmission in western Kenya and absence of *Rickettsia africae* and selected arboviruses in hospital patients. *Malar J*. 2022; 21(1): 268. doi: 10.1186/s12936-022-04287-3.
8. WHO - World Health Organization. Yellow fever - East, West, and Central Africa. 2022. Available from: <https://www.who.int/emergencies/disease-outbreak-news/item/2022-DON405>. [accessed 9 December 2023].
9. PAHO - Pan American Health Organization. Epidemiological alert: Yellow fever. 2022. Available from: <https://www.paho.org/en/documents/epidemiological-alert-yellow-fever-31-august-2022>. [accessed 09 December 2022].
10. Ma E, Cheng G. Host immunity and vaccine development against Dengue virus. *Infect Med (Beijing)*. 2022; 1(1): 50-8. doi: 10.1016/j.imj.2021.12.003.
11. Patel SS, Winkle P, Faccin A, Nordio F, LeFevre I, Tsoukas CG. An open-label, Phase 3 trial of TAK-003, a live attenuated dengue tetravalent vaccine, in healthy US adults: immunogenicity and safety when administered during the second half of a 24-month shelf-life. *Hum Vaccin Immunother*. 2023; 19(2): 2254964. doi: 10.1080/21645515.2023.2254964.
12. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. 2021; 29(71): 372. doi: 10.1136/bmj.n71.
13. Moola S, Munn Z, Tufanaru C, Aromataris E, Sears K, Sfetcu R, et al. Systematic reviews of etiology and risk. In: Aromataris E, Lockwood C, Porritt K, Pilla B, Jordan Z, editors. *JBIM Manual for Evidence Synthesis*. JBI. 2024. Available from: <https://synthesis-manual.jbi.global>.
14. Cohen J. *Statistical power analysis for the behavioral sciences*. 2nd ed. New York: Lawrence Erlbaum Associates, Publishers; 1988.
15. Siegel S, Castellan NJ. *Nonparametric statistics for behavioral sciences*. 2nd ed. McGraw-Hill Book Company; 1988.
16. The R Core Team. R: a language and environment for statistical computing. R Foundation for Statistical Computing. 2022. <https://www.r-project.org/>. [Accessed 10 October 2023].
17. Bbandeen-Roche K, Miglioretti DL, Zeger SL, Rathouz PJ. Latent variable regression for multiple discrete outcomes. *J Am Stat Assoc*. 1997; 92(440): 1375-86.



18. Hagens JA, McCutcheon AL. Introduction to applied latent class analysis. In: Hagens JA, McCutcheon AL, editors. Applied Latent Class Analysis. Cambridge University Press; 2002.
19. Vanebles WN, Ripley BD. Modern applied statistics with S. 4th ed. Springer; 2002.
20. Pande A, Guharoy D. A case report of *Plasmodium vivax*, *Plasmodium falciparum* and dengue co-infection in a 6 months pregnancy. Ann Med Health Sci Res. 2013; 3(Suppl. 1): S16-17.
21. Ward DI. A case of fatal *Plasmodium falciparum* malaria complicated by acute dengue fever in East Timor. Am J Trop Med Hyg. 2006; 75(1): 182-5.
22. Chipwaza B, Mugasa JP, Selemani M, Amuri M, Moshaf F, Ngantunga SD, et al. Dengue and Chikungunya fever among viral diseases in outpatient febrile children in Kilosa district hospital, Tanzania. PLoS Negl Trop Dis. 2014; 8(11): e3335. doi: 10.1371/journal.pntd.0003335.
23. Ayuthaya SI, Wangjirapan A, Oberdorfer P. An 11-year-old boy with *Plasmodium falciparum* malaria and dengue co-infection. BMJ Case Rep. 2014; 2014: bcr2013202998. doi: 10.1136/bcr-2013-202998.
24. Chong SE, Zaini RHM, Suraiya S, Lee KT, Lim JA. The dangers of accepting a single diagnosis: case report of concurrent *Plasmodium knowlesi* malaria and dengue infection. Malar J. 2017; 16(1): 2. doi: 10.1186/s12936-016-1666-y.
25. Otu AA, Udoh UA, Ita OI, Hicks JP, Ukpeh I, Walley J. Prevalence of Zika and malaria in patients with fever in secondary healthcare facilities in south-eastern Nigeria. Trop Doct. 2020; 50(1): 22-30. doi: 10.1177/0049475519872580.
26. Ayorinde AF, Oyeyiga AM, Nosegbe NO, Folarin OA. A survey of malaria and some arboviral infections among suspected febrile patients visiting a health centre in Simawa, Ogun State, Nigeria. J Infect Public Health. 2016; 9(1): 52-9. doi: 10.1016/j.jiph.2015.06.009.
27. Kinimi E, Patrick BN, Misinzo G. Serological evidence of chikungunya and malaria co-infection among febrile patients seeking health care in Karagwe district, Tanzania. Tanzania J Hlth Res. 2018; 30: 1-8. doi.org/10.4314/thrb.v20i4.1.
28. Assir MZ, Masood MA, Ahmad HI. Concurrent dengue and malaria infection in Lahore, Pakistan during the 2012 dengue outbreak. Int J Infect Dis. 2014; 18: 41-6. doi: 10.1016/j.ijid.2013.09.007.
29. Ahmad S, Dhar M, Mittal G, Bhat NK, Shirazi N, Kalra V, et al. A comparative hospital-based observational study of mono- and co-infections of malaria, dengue virus and scrub typhus causing acute undifferentiated fever. Eur J Clin Microbiol Infect Dis. 2016; 35(4): 705-11. doi: 10.1007/s10096-016-2590-3.
30. Alam A. A case of cerebral malaria and dengue concurrent infection. Asian Pac J Trop Biomed (India). 2013; 3(5): 416-7.
31. Arya SC, Mehta LK, Agarwal N, Agarwal BK, Mathai G, Moondhara A. Episodes of concurrent dengue and malaria. Dengue Bull. 2005; 29: 208-9.
32. Idoko MO, Ado SA, Umoh VJ. Prevalence of Dengue virus and Malaria in patients with febrile complaints in Kaduna metropolis, Nigeria. Microbiol Res J Int. 2015; 8(1): 343-7. https://doi.org/10.9734/BMRJ/2015/15588.
33. Schwarz NG, Mertens E, Winter D, Maiga-Ascofaré O, Dekker D, Jansen S, et al. No serological evidence for Zika virus infection and low specificity for anti-Zika virus ELISA in malaria positive individuals among pregnant women from Madagascar in 2010. PLoS One. 2017; 16: 12(5): e0176708. doi: 10.1371/journal.pone.0176708.
34. Shazia Y, Muhammad Owais R, Faisal M, Komal O. Co-existence of dengue fever & malaria in thrombocytopenic patients presented with acute febrile illness. PJMD. 2014; 3(3): 19-23.
35. Al-Areeqi A, Alghalibi S, Qais Y, Al-Masrafi I, Al-Kamarany MA. Epidemiological characteristic of malaria coinfecting with Dengue fever in Hodeidah, Yemen. Int J Trop Dis Health. 2020; 40(3): 1-10.
36. Mendonça VR, Andrade BB, Souza LC, Magalhães BM, Mourão MP, Lacerda MV, et al. Unravelling the patterns of host immune responses in *Plasmodium vivax* malaria and dengue co-infection. Malar J. 2015; 14: 315. doi: 10.1186/s12936-015-0835-8.
37. Magalhães BM, Siqueira AM, Alexandre MA, Souza MS, Gimaque JB, Bastos MS, et al. *P. vivax* malaria and dengue fever co-infection: a cross-sectional study in the Brazilian Amazon. PLoS Negl Trop Dis. 2014; 8(10): e3239. doi: 10.1371/journal.pntd.0003239.
38. Abbasi A, Butt N, Sheikh QH, Bhutto AR, Munir SM, Ahmed SM. Clinical features, diagnostic techniques and management of dual dengue and malaria infection. J Coll Physicians Surg Pak. 2009; 19(1): 25-9.
39. Epelboin L, Hanf M, Dussart P, Ouar-Epelboin S, Djossou F, Nacher M, et al. Is dengue and malaria co-infection more severe than single infections? A retrospective matched-pair study in French Guiana. Malar J. 2012; 11(11): 142. doi: 10.1186/1475-2875-11-142.
40. Faruque LI, Zaman RU, Alamgir AS, Gurley ES, Haque R, Rahman M, et al. Hospital-based prevalence of malaria and dengue in febrile patients in Bangladesh. Am J Trop Med Hyg. 2012; 86(1): 58-64. doi: 10.4269/ajtmh.2012.11-0190.
41. Magalhães BM, Alexandre MA, Siqueira AM, Melo GC, Gimaque JB, Bastos MS, et al. Clinical profile of concurrent dengue fever and *Plasmodium vivax* malaria in the Brazilian Amazon: case series of 11 hospitalized patients. Am J Trop Med Hyg. 2012; 87(6): 1119-24. doi: 10.4269/ajtmh.2012.12-0210.
42. Che Rahim MJ, Mohammad N, Besari AM, Wan Ghazali WS. Severe *Plasmodium knowlesi* with dengue coinfection. BMJ Case Rep. 2017; 20: bcr2016218480. doi: 10.1136/bcr-2016-218480.
43. Deresinski S. Concurrent *Plasmodium vivax* malaria and dengue. Emerg Infect Dis. 2006; 8(11): 1082.
44. Mittal G, Ahmad S, Agarwal RK, Dhar M, Mittal M, Sharma S. Aetiologies of acute undifferentiated febrile illness in adult patients - an experience from a tertiary care hospital in northern India. J Clin Diagn Res. 2015; 9(12): DC22-4. doi: 10.7860/JCDR/2015/11168.6990.
45. Halsey ES, Baldeviano GC, Edgel KA, Vilcarrromero S, Sihuinchu M, Lescano AG. Symptoms and immune markers in *Plasmodium*/Dengue virus co-infection compared with mono-infection with either in Peru. PLoS Negl Trop Dis. 2016; 29; 10(4): e0004646. doi: 10.1371/journal.pntd.0004646.
46. Kaushik RM, Varma A, Kaushik R, Gaur KJ. Concurrent dengue and malaria due to *Plasmodium falciparum* and *P. vivax*. Trans R Soc Trop Med Hyg. 2007; 101(10): 1048-50. doi: 10.1016/j.trstmh.2007.04.017.
47. Saksena R, Matlani M, Singh V, Kumar A, Anveshi A, Kumar D, et al. Early treatment failure in concurrent dengue and mixed malaria species infection with suspected resistance to artemisinin combination therapy from a tertiary care center in Delhi: a case report. Int Med Case Rep J. 2017; 16(10): 289-94. doi: 10.2147/IMCRJ.S139729.
48. Santana VS, Lavezzo LC, Mondini A, Terzian AC, Bronzoni RV, Rossit AR, et al. Concurrent dengue and malaria in the Amazon Region. Rev Soc Bras Med Trop. 2010; 43(5): 508-11. doi: 10.1590/s0037-86822010000500007.
49. Lupi O, Ridolfi F, da Silva S, Zanini GM, Lavigne A, Nogueira RM, et al. Dengue infection as a potential trigger of an imported *Plasmodium ovale* malaria relapse or a long incubation period in a non-endemic malaria region. Int J Infect Dis. 2016; 44: 20-4. doi: 10.1016/j.ijid.2016.01.008.



50. McGready R, Ashley EA, Wuthiekanun V, Tan SO, Pimanpanarak M, Viladpai-Nguen SJ, et al. Arthropod borne disease: the leading cause of fever in pregnancy on the Thai-Burmese border. *PLoS Negl Trop Dis*. 2010; 16(4): e888. doi: 10.1371/journal.pntd.0000888.
51. Yong KP, Tan BH, Low CY. Severe falciparum malaria with dengue coinfection complicated by rhabdomyolysis and acute kidney injury: an unusual case with myoglobinemia, myoglobinuria but normal serum creatine kinase. *BMC Infect Dis*. 2012; 20(12): 364. doi: 10.1186/1471-2334-12-364.
52. Stoler J, Delimini RK, Bonney JH, Oduro AR, Owusu-Agyei S, Fobil JN, et al. Evidence of recent dengue exposure among malaria parasite-positive children in three urban centers in Ghana. *Am J Trop Med Hyg*. 2015; 92(3): 497-500. doi: 10.4269/ajtmh.14-0678.
53. Bhagat M, Kanhere S, Phadke V, George R. Concurrent malaria and dengue fever: a need for rapid diagnostic methods. *J Family Med Prim Care*. 2014; (4): 446-8. doi: 10.4103/2249-4863.148146.
54. Mushtaq MB, Qadri MI, Rashid A. Concurrent infection with dengue and malaria: an unusual presentation. *Case Rep Med*. 2013; 520181. doi: 10.1155/2013/520181.
55. Kasper MR, Blair PJ, Touch S, Sokhal B, Yasuda CY, Williams M, et al. Infectious etiologies of acute febrile illness among patients seeking health care in south-central Cambodia. *Am J Trop Med Hyg*. 2012; 86(2): 246-53. doi: 10.4269/ajtmh.2012.11-0409.
56. Oyero OG, Ayukekbong JA. High dengue NS1 antigenemia in febrile patients in Ibadan, Nigeria. *Virus Res*. 2014; 191: 59-61.
57. Carme B, Matheus S, Donutil G, Raulin O, Nacher M, Morvan J. Concurrent dengue and malaria in Cayenne Hospital, French Guiana. *Emerg Infect Dis*. 2009; 15(4): 668-71. doi: 10.3201/eid1504.080891.
58. Charrel RN, Brouqui P, Foucault C, de Lamballerie X. Concurrent dengue and malaria. *Emerg Infect Dis*. 2005; 11(7): 1153-4. doi: 10.3201/eid1107.041352.
59. Serre N, Franco L, Sulleiro E, Rubio JM, Zarzuela F, Molero F, et al. Concurrent infection with Dengue type 4 and *Plasmodium falciparum* acquired in Haiti. *J Travel Med*. 2015; 22(5): 345-7. doi: 10.1111/jtm.12222.
60. Kolawole OM, Seriki AA, Irekeola AA, Bello KE, Adeyemi OO. Dengue virus and malaria concurrent infection among febrile subjects within Ilorin metropolis, Nigeria. *J Med Virol*. 2017; 89(8): 1347-53. doi: 10.1002/jmv.24788.
61. Mohapatra MK, Patra P, Agrawala R. Manifestation and outcome of concurrent malaria and dengue infection. *J Vector Borne Dis*. 2012; 49(4): 262-5.
62. Mørch K, Manoharan A, Chandy S, Chacko N, Alvarez-Uria G, Patil S, et al. Acute undifferentiated fever in India: a multicentre study of aetiology and diagnostic accuracy. *BMC Infect Dis*. 2017; 17(1): 665. doi: 10.1186/s12879-017-2764-3.
63. Mueller TC, Siv S, Khim N, Kim S, Fleischmann E, Arie F, et al. Acute undifferentiated febrile illness in rural Cambodia: a 3-year prospective observational study. *PLoS One*. 2014; 9(4): e95868. doi: 10.1371/journal.pone.0095868.
64. Mugabe VA, Ali S, Chelene I, Monteiro VO, Guiliche O, Muianga AF, et al. Evidence for chikungunya and dengue transmission in Quelimane, Mozambique: Results from an investigation of a potential outbreak of chikungunya virus. *PLoS One*. 2018; 13(2): e0192110. doi: 10.1371/journal.pone.0192110.
65. Ndosi R, Kwigizile E, Ibrahim U, Dossajee U, Rwiza J, Kabanayana C, et al. Risk factors for concurrent Malaria and arbovirus infections in Handeni, northeastern Tanzania. *Int J Trop Dis Health*. 2016; 20(4): 1-7. <https://doi.org/10.9734/IJTDH/2016/30632>.
66. Nkenfou CN, Fainguem N, Dongmo-Nguefack F, Yatchou LG, Kameni JJK, Elong EL, et al. Enhanced passive surveillance dengue infection among febrile children: prevalence, co-infections and associated factors in Cameroon. *PLoS Negl Trop Dis*. 2021; 15(4): e0009316. doi: 10.1371/journal.pntd.0009316.
67. Onyedibe K, Dawurung J, Iroezindu M, Shehu N, Okolo M, Shobowale E, et al. A cross sectional study of dengue virus infection in febrile patients presumptively diagnosed of malaria in Maiduguri and Jos plateau, Nigeria. *Malawi Med J*. 2018; 30(4): 276-82. doi: 10.4314/mmj.v30i4.11.
68. Joel MR, Annapoorna M, Usha S. A study on dual infections in pyrexia cases. *Int J Med Res Health Sci*. 2016; 5(8): 150-5.
69. Rao MR, Padhy RN, Das MK. Prevalence of dengue viral and malaria parasitic co-infections in an epidemic district, Angul of Odisha, India: an eco-epidemiological and cross-sectional study for the prospective aspects of public health. *J Infect Public Health*. 2016; 9(4): 421-8. doi: 10.1016/j.jiph.2015.10.019.
70. Shah PD, Mehta TK. Evaluation of concurrent malaria and dengue infections among febrile patients. *Indian J Med Microbiol*. 2017; 35(3): 402-5. doi: 10.4103/ijmm.IJMM\_15\_455.
71. Singh R, Singh SP, Ahmad N. A study of etiological pattern in an epidemic of acute febrile illness during monsoon in a tertiary health care Institute of Uttarakhand, India. *J Clin Diagn Res*. 2014; 8(6): MC01-3. doi: 10.7860/JCDR/2014/8965.4435.
72. Swoboda P, Fuehrer HP, Ley B, Starzengruber P, Ley-Thriemer K, Jung M, et al. Evidence of a major reservoir of non-malarial febrile diseases in malaria-endemic regions of Bangladesh. *Am J Trop Med Hyg*. 2014; 90(2): 377-82. doi: 10.4269/ajtmh.13-0487.
73. Thangaratham PS, Jeevan MK, Rajendran R, Samuel PP, Tyagi BK. Dual infection by dengue virus and *Plasmodium vivax* in Alappuzha District, Kerala, India. *Jpn J Infect Dis*. 2006; 59(3): 211-2.
74. Chander N, Singla J, Singh R. Concurrent presence of dengue and *Plasmodium falciparum*. *Trop Med Health*. 2009; 37(2): 69-70.
75. Raja JM, Mary A, Stagopan U. A study on dual infections in pyrexia cases. *Int J Med Res Health Sci*. 2016; 5(8): 150-5.
76. Kajeguka DC, Kaaya RD, Mwakalinga S, Ndossi R, Ndaro A, Chilongola JO, et al. Prevalence of dengue and chikungunya virus infections in north-eastern Tanzania: a cross sectional study among participants presenting with malaria-like symptoms. *BMC Infect Dis*. 2016; 16: 183. <https://doi.org/10.1186/s12879-016-1511-5>.
77. Mioto LD, Galhardi LCF, Amarante MK. Aspectos parasitológicos e imunológicos da malaria. *Biosaude*. 2012; 14(1): 42-55.
78. Donalísio MR, Freitas ARR, Von Zuben APB. Emerging arboviruses in Brazil: clinical challenges and implications for public health. *Rev Saude Publica*. 2017; 51: 30.
79. Pearson RD. Malaria. Family health version msd manual. 2020. Available from: <https://www.msmanuals.com/pt/casa/infeccões/infeccões-parasitárias-protozoários-extraintestinais/malária>. [accessed 19 January 2023].
80. Miccas FL, Batista SHSS. Educação permanente em saúde: metassíntese [Permanent education in health: a review]. *Rev Saude Publica*. 2014; 48(1): 170-85. doi: 10.1590/s0034-8910.2014048004498.
81. Alves ACJ, Dos Santos ACF, Peres JMV, Nascimento JMS, Barbosa DRL, Figueiredo JV, et al. Morphological atypia and molecular profile of *Plasmodium vivax*: findings from an outbreak in the Brazilian Amazon. *Parasite*. 2023; 30: 38. doi: 10.1051/parasite/2023039.
82. Silva TM. Estudo de prevalência e soroconversão (IgG) para arboviroses após a emergência de Zika e Chikungunya em uma amostra de adolescentes [Trabalho de conclusão de curso]. Rio de Janeiro: Universidade Federal do Rio de Janeiro; 2020.

83. SS - Secretária da Saúde do Ceará. Nota Técnica n. 01. Vigilância laboratorial e genômica das arboviroses. 2023. Available from: [https://www.saude.ce.gov.br/wp-content/uploads/sites/9/2018/06/NOTA\\_TECNICA\\_Deteccao-de-Arbovirus-1.pdf](https://www.saude.ce.gov.br/wp-content/uploads/sites/9/2018/06/NOTA_TECNICA_Deteccao-de-Arbovirus-1.pdf).
84. Mourão MPG, Bastos MS, Figueiredo RP, Gimaque JBL, Galuso EL, Kramer VM. Mayaro fever in the city of Manaus, Brazil, 2007-2008. *Vector Borne Zoonotic Dis.* 2012; 12(1): 42-6. doi: 10.1089/vbz.2011.0669.
85. Amélia PAT, Sueli GR, Márcio RTN, Mioni TFM, Jorge FST, Rosa PFC. Epidemia de febre do oropouche em Serra Pelada, município de Curionópolis, Pará, 1994. *Rev Soc Bras Med Trop.* 1996; 29(6): 537-41.
86. Alves AVG. Proteína não-estrutural (NS1) como ferramenta diagnóstica precoce e alvo terapêutico na dengue. *Revista Multidisciplinar em Saúde.* 2021; 2(4): 86. <https://doi.org/10.51161/rem/2230>.
87. Kariyawasam R, Lachman M, Mansuri S, Chakrabarti S, Boggild AK. A dengue vaccine whirlwind update. *Ther Adv Infect Dis.* 2023; 20: 10-20499361231167274. doi: 10.1177/20499361231167274.
88. Ooi EE, Kalimuddin S. Erratum for the Review “Insights into dengue immunity from vaccine trials”. *Sci Transl Med.* 2023; 15(709): eadk1254. doi:10.1126/scitranslmed.adk1254.
89. WHO - World Health Organization. Microscopy examination of thick and thin blood films for identification of malaria parasites. Malaria microscopy standard operating procedures. 2016. Available from: <https://www.who.int/publications/i/item/HTM-GMP-MM-SOP-08>.
90. PAHO - Pan American Health Organization. Tool for the diagnosis and care of patients with suspected arboviral diseases. Washington, DC: PAHO; 2017.