

# A Critical Review on Chagas Disease Chemotherapy

José Rodrigues Coura/<sup>+</sup>, Solange L de Castro\*

Departamento de Medicina Tropical \*Departamento de Biologia Celular e Ultraestrutura, Instituto Oswaldo Cruz-Fiocruz, Av. Brasil 4365, 21045-900 Rio de Janeiro, RJ, Brasil

*In this "Critical Review" we made a historical introduction of drugs assayed against Chagas disease beginning in 1912 with the works of Mayer and Rocha Lima up to the experimental use of nitrofurazone. In the beginning of the 70s, nifurtimox and benznidazole were introduced for clinical treatment, but results showed a great variability and there is still a controversy about their use for chronic cases. After the introduction of these nitroheterocycles only a few compounds were assayed in chagasic patients. The great advances in vector control in the South Cone countries, and the demonstration of parasite in chronic patients indicated the urgency to discuss the etiologic treatment during this phase, reinforcing the need to find drugs with more efficacy and less toxicity. We also review potential targets in the parasite and present a survey about new classes of synthetic and natural compounds studied after 1992/1993, with which we intend to give to the reader a general view about experimental studies in the area of the chemotherapy of Chagas disease, complementing the previous papers of Brener (1979) and De Castro (1993).*

Key words: Chagas disease - chemotherapy - review - *Trypanosoma cruzi* - drug development - clinical treatment

## BACKGROUND

Chagas disease is endemic in Latin America, affecting 16-18 million people, with more than 100 million exposed to the risk of infection (WHO 1997). Its etiological agent is *Trypanosoma cruzi*, an hemoflagellate protozoan (family Trypanosomatidae, order Kinetoplastida) (Hoare & Wallace 1966), whose life cycle involves obligatory passage through vertebrate (mammals, including man) and invertebrate (hematophagous triatomine bugs) hosts, in a series of stages. The trypomastigote ingested by the insect differentiates into the proliferative epimastigote form that, on reaching the posterior intestine, evolves to metacyclic trypomastigotes. This latter form, following invasion of vertebrate host cells, undergoes differentiation into amastigotes, which after several reproductive cycles transform to trypomastigotes, the form responsible for the dissemination of the infection. The transmission of the disease occurs mainly by the vector (80 a 90%), blood transfusion (5 a 20%) and congenital routes (0.5 a 8%) (Dias 2000).

In humans, after infection and a subsequent incubation period, the acute phase of Chagas disease begins, and in the absence of specific treatment, the symptoms persist for about two months, with a mortality rate of 2 to 8%, especially among children. *T. cruzi* is able to invade and multiply within different host cells, including macrophages, smooth and striated muscles, fibroblasts and even neurons. The first reaction to *T. cruzi* is focal mononuclear inflammation due to rupture of parasitized cells. Within days to two weeks can be detected in the serum the presence of immune complexes, decrease in C3 level, besides

necrosis in the inflammatory foci. Severe inflammation is usually accompanied by necrosis of parasitized and non-parasitized cells, especially in the heart. Platelet aggregation, eosinophils degranulation, microvascular pathology, edema, thrombosis, blood stasis and ischaemia have also been demonstrated (Andrade & Andrade 1999).

After the acute infection, the patient presents strong evidence of immunity, but has a tendency to remain infected. Some parasites evading the immune response and focal inflammatory lesions are seen in several organs. Amastigote forms can be detected by conventional histology, by immunofluorescence and genomic markers by *in situ* hybridization. The combination of rising immunity against the parasite with specific immunological suppression of hypersensitivity and reduction of inflammatory reaction seem to be the main pathways to the indeterminate phase of Chagas disease (Andrade 1999).

In the chronic phase that follows, most patients remain asymptomatic, with about 20 to 50% of the cases, according to the endemic area analyzed, developing the characteristic symptoms of this phase, namely cardiac, digestive or neurological disturbances (revised in Brener et al. 2000). Chronic, active, fibrosing myocarditis have been attributed to hypersensitivity to parasite antigens, neoantigens or autoimmunity. The presence of cross-reactive antigens between heart muscle and *T. cruzi* has been demonstrated, but the autoimmunity does not entirely explain Chagas heart disease. The high positivity of xenodiagnosis and hemoculture, and reactivation of chronic disease by immunosuppression demonstrate the presence of the parasite in chronic cases. High frequency of parasites and/or antigens associated with myocardial inflammation is an important guide to the therapeutic procedures in the chronic phase.

The pathogenesis of Chagas disease is not yet completely defined and understood, with two basic inflammatory lesions, one focal and the other diffuse. The focal lesion is associated with the parasite, and occurs when parasitized cells are disrupted. The diffusion lesion is out of proportion in relation to the presence of parasites. During the acute infection there are diffuse lesions in the heart and focal lesions in several other organs. In the chronic heart disease, severe fibrosis and inflammatory lesions seem not to be related only to the presence of *T. cruzi*, but

This research was supported by grants from the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Programa de Apoio à Pesquisa Estratégica em Saúde (Papes/Fiocruz), Fundação Carlos Chagas Filho de Amparo à Pesquisa do Rio de Janeiro (Faperj), and Fundação Nacional da Saúde (Funasa).

\*Corresponding author. Fax: +55-21-2280.3740. E-mail: coura@ioc.fiocruz.br

Received 5 November 2001

Accepted 10 December 2001

also to a strong delayed hypersensitivity host response and ischaemic lesions (Andrade 1999, Andrade & Andrade 1999, Higuchi 1999). Two mechanisms are proposed for pathogenesis in chronic chagasic infections: the persistence of the parasite results in chronic inflammatory reactivity and it induces immune responses targeted at self-tissues (reviewed in Tarleton 2001). Several clinical reports reinforces the first mechanism (Higuchi et al. 1993, 1997, Jones et al. 1993, Anez et al. 1999), while the main support of the second one is that signs of the disease are evident in tissues in the apparent absence of parasites.

Several reviews about clinical and/or experimental Chagas disease treatment have been published as articles (Coura & Silva 1961, Prata 1963, Brener 1975, 1979, 1984, De Castro 1993, Coura 1996, Urbina 1999, Stoppani 1999) and as book chapters (Brener 1968, 2000, Cañado & Brener 1979, Cañado 1968, 1985, 1997, 1999, 2000, Rassi & Luqueti 1992, Storino et al. 1994). We will detail here just some of the publications and refer readers to the reviews cited above.

The aim of this “Critical Review” is to analyse drugs employed in the clinics since the 70s bringing attention to the recommendations about treatment, and evaluation of cure, and the studies about the development of new drugs, considering potential targets in the parasite and summarizing the experimental studies performed with new compounds assayed against *T. cruzi* after 1992/1993. For a more complete coverage of experimental *in vitro* and *in vivo* studies we suggest the reviews of Brener (1975, 1979) and De Castro (1993).

#### EXPERIMENTAL AND CLINICAL TREATMENT

##### Drugs assayed up to the decade of 70

The first compounds assayed experimentally for the treatment of Chagas disease, soon after its discovery by Carlos Chagas in 1909, were atoxyl (arsenical), fucsin (rosanilin dye), tartar emetic (or antimony potassium tartarate, a pentavalent antimonial) and mercury chloride, employed experimentally by Mayer and Rocha Lima (1912, 1914) and all of them without effective results. Until the publication of the “Manual de Doenças Tropicais e Infectuosas” by Carlos Chagas and Evandro Chagas (1935) “there was no specific treatment for American trypanosomiasis. Drugs with trypanocidal activity have been assayed by a great number of researchers, but without success”, affirm the authors (p. 144).

Among the chemotherapeutic agents employed until 1962 stand out quinolein derivatives, several other antimalarials, arsenobenzoles, and other arsenicals, phenantridines, salts of gold, bismuth, copper and tin, sodium iodide, gentian violet, aminopterin, para-amino salicylic acid, nicotinic acid hydrazide, antihistaminics, sulfonamides, ACTH, cortisone, stylomycin derivatives, amphotericin B, more than 30 antibiotics and some nitrofurans (reviewed in Coura & Silva 1961, Brener 1968, Cañado 1968).

Brener (1968) made a meticulous evaluation of the experimental drugs assayed *in vitro* and *in vivo* against *T. cruzi*, registering 27 compounds and more than 30 antibiotics, assayed between 1912 and 1962, that were inactive. He also considered that the following compounds had a suppressive effect on the parasitemia but were not curative: the bisquinaldine Bayer 7602 (Cruzon, Imperial Chemical Industry, UK), the phenantridine carbidium sulfate (74C48), aminoquinolines (pentaquine, isopentaquine and

primaquine), trivalent arsenicals (Bayer 9736 and Bayer 10557 also named spirotrypan), aminoglycoside of stylomycin, nitrofurans and antibiotics.

Packchianian (1952, 1957) opened a new and promising line of potential drugs with the nitrofurans that led to nitrofurazone (5-nitro-2-furaldehyde-semicarbazone). This derivative administered by oral route for 53 days consecutively in the dose of 100 mg/kg/day mice experimentally infected with *T. cruzi* cured 95.4% of the animals (62/65) (Brener 1961)

Ferreira (1961, 1962) and Ferreira et al. (1963) treated the first ten cases of acute Chagas disease with this nitro-furan, obtaining “good clinical results” with few collateral effects but the xenodiagnosis became positive in five cases after treatment Coura et al. (1961, 1962) treated 14 chronic cases with this drug in long-term schemes, observing in the first four patients, that received progressive doses of 10 to 30 mg/kg/day, important side effects that led to suspension of the treatment due to severe sensitive polyneuropathy, that began at the third week of nitrofurazone administration. With reduction of the dose to 10 mg/kg/day and association with complex B, administered by parenteral route, five patients tolerated the treatment for 60 days, in spite of the side effects (anorexia, weight loss, paresthesia, and sensitive polyneuropathy). Another patient was treated with 20 mg/kg/day and presented paresthesics manifestations only at the 53th day of treatment, evolving to a severe sensitive polyneuropathy, with termination of the treatment by his own decision (informed consent). From the six patients submitted to long-term treatment, two of them were considered cured, based on xenodiagnosis and serology (complement fixation) persistently negative. Cañado et al. (1964) also treated five chronic patients with 10 mg nitrofurazone/kg/day during 10 to 34 days when the treatment had to be suspended due to polyneuritis, being all the five patients considered as therapeutic failures.

In a critical analysis of the literature about the clinical experiences, Cañado (1968) emphasized the lack of methods in the execution, preferential selection of acute cases, based on the remission of the symptoms and signs that could also be spontaneous. He cited, as example, the reports of Mazza et al. (1937, 1942) and Pifano (1941) about the results with Bayer 7602 and Bayer 9736 that were considered “excellent results” only based on the reduction of the symptoms and signals. Both compounds were in fact ineffective, since the xenodiagnosis after treatment remained positive and untreated cases had also reduction of the symptomatology. In subsequent works (Cañado et al. 1973, Cañado 1981) reviewed the results of therapeutic trials in the period of 1936 to 1965 and proposed basic criteria for the evaluation of the specific treatment, that were later updated by 15 experts from Latin America (OPS/OMS 1998).

Following the requirements of the World Health Organization (WHO) the ideal drug for the treatment of Chagas disease should fulfill the following requirements: (i) parasitological cure of acute and chronic cases; (ii) effective in single or few doses; (iii) accessible to the patients, in other words, of low cost; (iv) no collateral or teratogenic effects; (v) no need of hospitalization for the treatment; (vi) no induction of resistance.

As we will see below this ideal drug does not exist and possibly it will take a long period of time to be obtained. Since the end of 1960 beginning of the 70s two drugs have

been used for the treatment of Chagas disease: nifurtimox and benznidazole.

### Nifurtimox and benznidazole

The drugs and Chagas disease treatment

Nifurtimox (Nif) is a 5-nitrofuran (3-methyl-4-(5'-nitrofururylideneamine) tetrahydro-4H-1,4-tiazine-1,1-dioxide (Bayer 2502) and benznidazole (Bz) is a 2-nitroimidazole (N-benzyl-2-nitroimidazole acetamide (RO 7-1051), commercialized, respectively, with the names Lampit and Rochagan in Brazil (Radanil in Argentina) (Fig. 1a,b). Nif that was the most active 5-nitrofururylidene derivative experimentally assayed (Bock et al. 1969) and Bz showed a high in vitro and in vivo activity against *T. cruzi* (Richle 1973). Since the 80s Nif had its commercialization discontinued, first in Brazil, and then in Argentina, Chile and Uruguay. The mode of action of Nif involves generation of nitroanion radical by nitroreductases that, in the presence of oxygen, led to reactive intermediates and being *T. cruzi* is partially deficient in free radical detoxification mechanisms, it is susceptible to such intermediates (reviewed in DoCampo & Moreno 1986). On the other hand, this oxidative damage was not the key action of Bz, the detection of corresponding nitroanion radical occurred only at concentrations much higher than those that killed the parasite. The action of Bz could involve covalent bond or other interactions of nitroreduction intermediates with parasite components (Polak & Richle 1978), or binding to DNA, lipids and proteins (Diaz de Toranzo et al. 1988).

Nif and Bz have been employed by different authors, especially in Brazil, Chile and Argentina (Cañado et al. 1969, 1973, 1975, Cañado & Brener 1979, Bocca-Tourres 1969, Rubio & Donoso 1969, Schenone et al. 1969, 1972, 1975, 1981, Rassi & Ferreira, 1971, Rassi & Luquetti 1992, Cerisola et al. 1972, 1977, Prata et al. 1975, Ferreira 1976, 1990, Coura et al. 1978, 1997, Macêdo & Silveira 1987, Viotti et al. 1994, Andrade et al. 1996, Sosa Estani et al. 1998).

The results obtained with both drugs varied according to the phase of Chagas disease, the period of treatment and the dose, the age and geographical origin of the patients. Good results have been achieved in the acute phase, in recent chronic infection (children under 12 years old), congenital infection and laboratory accidents. For the acute phase treatment and congenital cases it is recommended 8 to 10 mg/kg/day of Nif or 5 to 7.5 mg/kg/day of Bz during 30 to 60 days consecutively, and divided in two or three daily doses. Patients with less than 40 kg can take up to 12 mg/kg/day of Nif and up to 7.5 mg/kg/day for Bz during 30 to 60 days (OPAS/OMS 1998). For recent chronic infection (children under 12 years old) or individuals infected in the last 10 years, the treatment should be made with 8 mg/kg/day of Nif or 5 mg/kg/day of Bz during 30 to 60 days. In the case of accidental infection the treatment must begin immediately and last for only 10 to 15 consecutive days. Cases of late chronic infections without clinical manifestation or with mild cardiac or digestive manifestations should be treated during 60 to 90 days, in accordance with the tolerance to the drugs, aiming to prevent or reduce the evolution of Chagas disease to more severe forms, a fact that is not yet definitely proved.

### Side effects and contraindications

The more frequent collateral effects with Nif treatment are anorexia, loss of weight, psychic alterations, excitability or sleepiness and digestive manifestations, such as nausea, vomit and occasionally intestinal colic and diar-

rhea. The adverse reactions with Bz could be classified in three groups: (i) symptoms of hypersensitivity, dermatitis with cutaneous eruptions (usually appearing between the 7th and 10th day of treatment), generalized edema fever, lymphadenopathy, articular and muscular pain; (ii) depression of bone marrow, thrombocytopenic purpura and agranulocytosis, the most severe manifestation; (iii) polyneuropathy, paresthesia and polyneuritis of peripheral nerves.

The two most serious complications induced by Bz are agranulocytosis, initiated by neutropenia, sore throat, fever and septicemia, and thrombocytopenic purpura, characterized by reduction of platelets, petechiae, hemorrhagic blister and even mucosal bleeding. At the first signs of such manifestations, medication must be immediately suspended and should be started a treatment with antibiotics in the case of septicemia plus corticosteroids for the control of the agranulocytosis and of the thrombocytopenic purpura. Other manifestations of intolerance and hypersensitivity could be circumvented with the reduction of the dose or suspension of the drug, depending on their intensity, introduction of symptomatic medication and eventually of anti-histaminics and corticosteroids.

Teixeira et al. (1990, 1994) have been alerting for the appearance of lymphomas and mutagenic and carcinogenic activities in experimental animals (rabbits and mice) treated with Bz. However, a broad review of thousands of patients treated with these drugs by several authors has not demonstrated such effects.

Bz and Nif should not be indicated for pregnant patients, in cases of severe diseases associated with Chagas disease, such as systemic infections, cardiac, respiratory, renal or hepatic insufficiency, hemopathies and neoplasias without the possibility of treatment, old-aged and very debilitated persons.

### Effect of the treatment on the evolution of Chagas disease

Since 1969 several therapeutic experiences have been performed in acute and chronic cases of Chagas disease using Nif (Bocca-Tourres 1969, Rubio & Donoso 1969, Schenone et al. 1969, 1972, Rassi & Ferreira 1971, Cerisola et al. 1972, 1977, Silva et al. 1974, Prata et al. 1975, Cañado et al. 1975, 1976, Cañado & Brener 1979), Bz (Schenone et al. 1975, Ferreira 1976, Coura et al. 1978, Viotti et al. 1994, Andrade et al. 1996, Sosa Estani et al. 1998) and also comparing the efficacy and tolerance of both drugs in different groups of patients, therapeutic schemes and periods of follow-up and cure evaluation criteria (Schenone et al. 1981, Ferreira 1990, Coura et al. 1997, Lazzari & Freilig 1998).

The results of such experiences showed a great variability, according to the authors, and the type of casuistic and of cure control employed. In general, results obtained were good for acute phase and recent infection cases, especially among children, who, besides tolerating a long-term treatment much better, presented high indexes of cure, as demonstrated in a randomized field study with Bz in Brazil (Andrade et al. 1996) and in Argentina (Sosa Estani et al. 1998). For acute cases and recent infections an average index of parasitological cure around 60% is estimated. In relation to chronic infection cases, results have been poor, for the Brazilian experience, while in the South Cone, they were much better, possibly due to the type of parasite strain (Silva et al. 1974, Cerisola et al. 1977).

Studies on the clinical evolution of Chagas disease after specific treatment are controversial and results are not convincing, due to differences in casuistics, methods

of evaluation, time of follow-up and interpretation of the data. Macêdo and Silveira (1987) studying 171 adults with chronic disease treated with Nif or Bz with a follow-up of 7 years, observed electrocardiographic (ECG) evolution of cardiopathy in 6.7% of the cases, against 8.8% for untreated patients, indicating no significant differences between the two groups. Ianni et al. (1993) monitored 33 adults in the indeterminate phase for 8 years, reported the evolution in 13.3% of the cases treated with Bz (n = 15) and 0% of the cases that received placebo (n = 18), not allowing a definitive conclusion, due to the small casuistic. Miranda et al. (1994 *apud* OPAS/OMS 1998), in 120 patients (adults and children), observed ECG evolution in 10.5% of those treated with Bz and 63.6% of the placebo group. Although these authors monitored the patients for 10 to 16 years, the combination of the data obtained with adults and children and the ECG interpretation made the analysis of the results obtained difficult.

Viotti et al. (1994) in a well-designed study with 201 adult patients monitored for 8 years, observed ECG evolution in 7/131 cases (5.3%) treated with Bz (5 mg/kg/day for 30 days) and 16/70 (22.8%) in the control group. In patients with more than 50 years old, the ECG alterations occurred in 3/36 (8.3%) of the treated cases and 7/40 (17.5%) of the untreated ones; differences were not statistically significant. For those under 50 years old, such alteration occurred in 4/95 (4.2%) and 9/30 (30%), respectively, for treated and untreated patients, and were significantly different. Two patients died during the follow-up, one treated and one untreated. Although the study has been well conducted, after 8 years of follow-up, 68% of the untreated patients presented positive serology against 48.2% of the Bz-treated group. The low number of ECG alterations and their frequent mutability in chronic cases make the interpretation of the data difficult.

On the other hand, Fragata Filho et al. (1995) reported, in a study with 120 chronic patients with follow-up for 7-8 years, ECG evolution in 7% for Bz-treated cases (n = 71) and 14.3% for the untreated group (n = 49).

Andrade et al. (1996) performed a randomized field study in the State of Goiás (Brazil) treating children between 7 and 12 years old with positive serology for Chagas disease. Sixty-four patients received 7.5 mg/kg/day of Bz for 60 days and 65 received placebo. From these 129 children, 88.7% (58 treated and 54 control) were monitored for three years. The authors considered the treatment effective in 55.8% of the treated cases (most of them with a significant decrease in serological titers). However, no significant results were observed when the ECG abnormalities were compared. In a similar way, in Salta (Argentina), Sosa Estani et al. (1998) treated 55 children from 6 to 12 years old with 5 mg Bz/mg/day for 60 days and 51 children with placebo, and monitored the study for four years. They observed negatization or significant decrease of the specific serology in 62% of the treated group and none for the control group. In relation to xenodiagnosis the positivity was 4.7% and 51.2%, respectively for treated and untreated children, indicating an important suppressive activity of the treatment, however, the ECG alterations were similar for both groups: 2.5% (1/40) and 2.4% (1/41).

#### Rules and recommendations for the clinical treatment

A meeting of 13 specialists promoted in Brasília by the Ministry of Health of Brazil summarized by Luquetti (1997) and another one by the Pan American Health Organization and World Health Organization (OPAS/OMS 1998) that

took place in Instituto Oswaldo Cruz (Fiocruz) in April 23-25 of 1998, established some rules and recommendations for the etiologic treatment of Chagas disease, with the present available drugs, that are summarized below:

#### Acute phase

In this phase the parasite is detected by direct examination of peripheral blood by wet smear or in stained cover slides or by concentration methods (centrifugation of the blood and microscopic examination of leukocyte cream or stained quantitative buffy coat techniques). With or without clinical manifestations, the detection of the parasite by direct methods or determination of IgM levels allows the diagnostic of the acute phase. Independently of the mechanism of transmission (vectorial, transfusional, by oral route, or laboratory accident) the patients must be treated, as indicated previously, since about 60% of them could be cured in the acute phase.

#### Congenital infection

The diagnostic of congenital infection is based in cases of children from infected mothers, serologically positive, who presented *T. cruzi* in the blood of umbilical cord, specific IgM in the serum soon after birth, or IgG after 6 months, when the possibility of vectorial, transfusional and oral transmission are absent. The treatment is similar to that of the acute phase patients.

#### Accidental infection

The person, technician or researcher, that working with *T. cruzi*, was accidentally punctured by infected needle, ingested or had any contact with infected materials in lesions, wounds, mucosal, or any other form indicative of the possibility of the parasite penetration is considered infected. In these cases, blood is collected for serological test and treatment immediately begins, during 10 to 15 days, repeating the serology after 15, 30, and 60 days after the accident. It is recommended that all the laboratories that work with *T. cruzi* have the drug available.

#### Chronic phase

*Recent chronic phase* - Recent chronic phase is considered when the infection was acquired in the last 10 years, including children up to 12 years old or adults that have been infected occasionally in endemic areas of by blood transfusion in a known interval of time. The published work indicates that the results obtained by the etiologic treatment in this phase are much better than that performed in late chronic patients.

*Late chronic phase* - Patients with more than 10 years of infection are considered late chronic cases. There is no agreement about the clinical evolution of such cases, and parasitological cure is obtained in 10 to 20% of the patients, according to different experiences. The treatment must be elective, with priority to cases in the indeterminate form or with minor pathology that may be monitored by long periods of time, after treatment.

*Transplant of organs* - In cases of transplants it is always necessary to perform specific serological reaction both in the donor and the receptor. The transplant of organs from patients with infection by *T. cruzi* could transmit the parasite to the receptor, especially during the immunosuppression phase. On the other hand, in the infected receptor a reactivation of the disease could occur through the immunosuppression, compromising even the transplanted organ, mainly in cardiac cases. Jatene et al. (1997) refer to reactivation of Chagas disease in 40% of

cases derived from immunosuppression in heart transplants. Both the donor and the receptor must be treated with Bz in the dose of 5 mg/kg/day for at least 60 days. Tomitori-Yamashita et al. (1997) considered that "allopurinol seems a safe and effective treatment for reactivated Chagas disease after heart transplantation, although it is not recommended as a post-transplantation prophylaxis because reactivation of the disease is unpredictable".

#### Reactivation of Chagas infection

Reactivation of the chronic disease can occur due to immunosuppression by several diseases, such as leukemia, lymphoma and other neoplasias, infection by HIV/AIDS and in the cases of transplants with immunosuppression. Meningoencephalitis and acute myocarditis are the most frequent manifestations (reviewed in Ferreira et al. 1997a). In chronic patients parasites were detected in cutaneous lesions after transplant with immunosuppressive treatment and in the smooth muscle after cancer chemotherapy. Ferreira et al. (1997a,b) made an extensive review about this topic, recommending as first choice the treatment with Nif or Bz, indicating as alternatives triazole derivatives and allopurinol. Long-term secondary prophylaxis should be recommended for patients who respond to therapy, although it is uncertain which drug to use for this purpose (Ferreira et al. 1997b). Anyway all HIV-positive cases, patients with neoplasias or candidates to transplants must be thoroughly investigated for the possibility of concomitance with chagasic infection. There is no consensus about the prophylactic indication of the etiologic treatment of the infection, in cases without clinical reactivation, but with xenodiagnosis or hemoculture positive for *T. cruzi*.

Patients in the chronic phase and receiving corticoid because of concomitant diseases were treated with Bz since the beginning of the use of corticoid (n=12) or 15 days afterwards (n = 6) (Rassi et al. 1999). The authors observed that Bz prevented an increase of parasitemia, and suggested that in immunocompromized patients with chronic Chagas disease this drug could be useful.

#### Where and who should treat the patient

The patients in severe acute phase and the congenital symptomatic cases with diagnostic at birth, must be hospitalized for treatment. The acute oligosymptomatic or chronic cases can be treated in basic health care units under the supervision of an experienced physician. Bz and Nif must be considered drugs of high complexity, and recommended only by professionals with solid information about the side effects and the disease in itself. The acute phase and the accidental infection are emergency situations, and treatment must start immediately even with a professional without experience that must search a colleague or a qualified institution for orientation.

#### Evaluation of cure

The evaluation of cure of Chagas disease is certainly the more complex aspect of its treatment, leading several times to diverse and controversial results, in relation to both parasitological cure and clinic cure. The term parasitological cure itself is of difficult interpretation and the evaluation is almost impossible, since it would mean the total elimination of the parasite not only from the blood but also from all tissues. So, in humans it is not viable to be confirmed. On the other hand, clinic cure is the long-term evaluation and several times uncertain due to the pathogenesis of the disease, which involves the action of parasite and the immune and autoimmune response of the pa-

tient and in antigenic complexes deposition, generation of antibodies, inflammatory reactions, tissular lesions with cellular degeneration, ischaemia, fibrosis and their consequent clinical manifestations, sometimes for long periods of time.

For the evaluation of experimental animals, mostly in drug assays, the situation is less complex. The in vitro tests (tissue culture) are not necessarily reproduced in vivo. On the other hand, as occurs in humans, a suppressive effect on the parasitaemia does not correspond exactly to the effect of a drug in tissues. In his pioneer work Brener (1961) demonstrated parasitological cure in 94.5% of the mice treated with nitrofurazone. Later, Brener et al. (1969) demonstrated by electron microscopy that 13.5% of the amastigotes of the Y strain were intact in heart cells of the treated animals. A question remained: were these findings due to populations of the parasite with primary resistance or the drug did not reach all the infected cells? The group of Andrade reported development of resistance to both nitroheterocycles and the influence of *T. cruzi* strain in the cure rate, for example, Bz cured 87% of the mice infected with the Peru strain, and only 16.7% in the case of Colombian strain (Andrade et al. 1975, 1977, Andrade & Figueira 1977). Resistance to both drugs, including cross resistance, was also observed in animals infected with the Y strain (Costa Silva et al. 1990). Could this be due to a mechanism similar to that questioned above?

#### Parasitological evaluation

The suppressive activity on the parasitaemia is almost immediate after the beginning of the treatment when the strain (population) of *T. cruzi* is susceptible to the drug employed. In acute cases this fact could be verified by the direct examination of blood in fresh or stained preparations or by concentration methods. In chronic cases, the usual methods are xenodiagnosis standardized with 40 nymphs of 3rd/4th stage of *Triatoma infestans* or by 20 nymphs of this species and another 20 from *Panstrongylus megistus* or of other species that could give similar yield. The nymphs are distributed in four boxes (10 per box); two are placed in the internal face of each arm for about 30 min (Coura et al. 1991). In this study we observed a positivity of 50.7% in 570 xenodiagnosis performed in 246 patients. Nowadays, due to ethical questions and comfort for the patients, the preferential method is the artificial xenodiagnosis that consists in collection of 10 ml of the blood that are placed in a condom type membrane, with external heating, and then the nymphs are added and the reading done after 30, 45 and 60 days. The yield obtained is similar to that of the natural test (Pineda et al. 1998).

In a multicenter study that involved researchers from 10 Brazilian institutions and 312 Bz-treated patients monitored by a media of 12 xenodiagnosis performed after treatment, suppression of parasitaemia was demonstrated in 78% of the cases (Coura et al. 1978). In another comparative controlled study with 77 chronic patients treated with Bz (n = 26), Nif (n = 27) or placebo (n = 24), suppression of the parasitaemia monitored xenodiagnosis by one year after treatment was achieved in 98.1% (2/110) of the Bz group, and in 90.4% (75/83) of the Nif group (Coura et al. 1997). However, this result does not imply that cases with xenodiagnosis negative are cured since only 34.3% of the control group was positive.

The hemoculture is the second parasitological method of choice for the control of cure in chronic Chagas disease, being equivalent in sensibility to xenodiagnosis

(Chiari & Brener 1966). Both methods have a tendency to increase the positivity with the number of tests performed, amount of blood employed, cultivation medium, interval of time between blood collection and cultivation and other factors emphasized by Chiari et al. (1989) aiming standardization of the assay. Using 30 ml of blood seeded in six test tubes with LIT medium and readings up to 60 days, Chiari et al. (1989) obtained a positivity up to 50%, while with small modifications of the technique such as direct seeding soon after blood collection, refrigerated centrifugation for a short time, gentle homogenization and up to 120 days led Luz et al. (1994) to a positivity of 94%, not achieved by any other author, for assays with chronic cases of Chagas disease.

Polymerase chain reaction (PCR) was a major advance for the parasitological control of the cure of Chagas disease, with positivity 2 to 3 times higher for chronic cases when compared to routine xenodiagnosis and hemoculture. By this technique it is possible to detect one parasite or a fragment of *T. cruzi* DNA in 20 ml of blood (Ávila et al. 1991).

Sturm et al. (1989) amplifying minicircles of DNA of *T. cruzi* obtained fragments of 83 and 22 pairs of base (bp) from variable regions that were employed for detection of the parasite (Moser et al. 1989). Ávila et al. (1991) using a solution of 6 M guanidine plus EDTA and equal amount of blood of chronic patients, promoted lysis of proteins and maintained the integrity of the DNA sample at room temperature. The treatment of this lysate with phenantroline-copper (OP-Cu<sup>2+</sup>) led to the cleavage of DNA and liberation of minicircles, allowing the identification of a single parasite in 20 ml of blood when three initiators for the fragments of 83, 122 and 330 bp from the variable and constant regions of the minicircles. By this technique, the authors identified *T. cruzi* in samples of 10 ml of blood of five chronic patients, four of them with negative xenodiagnosis.

Several authors (Wincker et al. 1994, Britto et al. 1995, 2001, Junqueira et al. 1996) have demonstrated the efficiency of PCR for the diagnosis of chronic disease and for the control of cure after treatment. However, Junqueira et al. (1996), in a comparative study among PCR, xenodiagnosis and hemoculture in 101 chronic cases, observed positivity of, respectively, 59.4%, 35.6% and 25.7%, but in five cases with positive xenodiagnosis and/or hemoculture, the result of PCR was negative. Recently, Britto et al. (2001) comparing PCR and xenodiagnosis in the control of cure of 85 chagasic patients submitted to specific treatment and 15 chronic asymptomatic cases that received placebo, reported that in all the cases of positive xenodiagnosis, positivity was obtained also by PCR. On the other hand when xenodiagnosis was negative, PCR was positive in 18.5% of the acute phase group (n = 37), 29% of the chronic phase group (n = 45) and 57.1% of the control group. These results demonstrate the advantage of PCR over conventional techniques to demonstrate persistent infections in patients that underwent chemotherapy.

#### Serological evaluation

This evaluation is certainly the most simple, more broad and reliable for the control of cure of chagasic infection after treatment, especially in the chronic phase when the serology is positive in almost 100% of the untreated cases. Whereas the positivity of parasitological methods depends on the random presence of the parasite in the blood sample,

the presence of antibodies is almost warranted in all the samples. On the other hand, the serology since the Guerreiro and Machado reaction (1913) until the qualitative and quantitative reaction of complement fixation (Kelser 1936, Freitas & Almeida 1949), the indirect immunofluorescence assay (Fife & Muschel 1959, Camargo 1966), the hemagglutination assay (Neal & Miles 1970), the conventional ELISA (Voller et al. 1975) and with recombinant antigens (reviewed in Silveira 1992, Silveira et al. 2001), the lysis mediated by complement (Kretzli & Brener 1982, Kretzli et al. 1984) and finally techniques using immunoblots have been improved as confirmatory tests due to their sensibility and specificity (Umezawa et al. 1996, 2001).

The three basic serological reactions for the diagnosis of Chagas disease are indirect immunofluorescence, hemagglutination and ELISA. During the years, the great polemic is their negatization in cases of parasitological cure. Some authors as Cançado (1963, 1997) consider cure as “definitive post-therapeutic reversion to negativity of parasitological and serological tests”, whereas others, like Rassi and Luquetti (1992), Andrade et al. (1996) and Sosa Estani et al. (1998) admit a long period of negatization of the reactions and even low serological titers as criteria of cure. Andrade et al. (1991) demonstrated that, in mice infected with *T. cruzi* and parasitologically cured by chemotherapy, parasite antigens persist in interstitial dendritic cells in the spleen and the animals present positive serology. Recently Andrade et al. (2000) reported the importance of the presence of parasite antigens in the same type of cells in the heart of infected dogs and the pathogenesis of the chagasic myocarditis probably by presentation of *T. cruzi* antigens to immune-competent cells, and, as consequence, maintenance of the response to the infection.

#### Clinical evaluation

This type of the cure evaluation after chemotherapy is perhaps the most difficult and long topic to be addressed. In this review, we have already discussed some aspects of the clinical evaluation when analyzing the evolution of Chagas disease after treatment with Nif or Bz, so, in this topic we will discuss only some essential clinical tests for monitoring the disease, before, during and after treatment.

In the clinical evaluation of the treatment we must consider, besides the anamnesis and clinical examination, the electrocardiographic and radiologic aspects together with other non-invasive tests with high sensibility, such as dynamic electrocardiography (Holter) for the study of arrhythmias and echocardiography for the anatomophysiological evaluation of the cardiac function, the endoscopy and manometry for the anatomofunctional study of the digestive system and some other tests for evaluation of the autonomous nervous system and neuronal lesion, besides biopsies for histological and histochemical studies, that will not be evaluated here in depth.

A careful clinical examination after a detailed anamnesis, especially analyzing the cardiovascular, digestive and neurologic systems, before, during and after treatment, monthly in the first year and at least once a year subsequently is fundamental importance for the evolutive study of the patients. The ideal condition, when ethics allows, would be the monitoring of a control group, of the same age and sex, untreated or receiving a placebo, with aspect similar to the drug, for evaluation of collateral effects and a comparative study between the treated and the control groups.

The standard electrocardiogram, with the limb leads (D1, D2, D3, aVR, aVL and aVF) and chest leads from V1 to V6 is the most simple and most important examination in the clinical evaluation. This test must be associated with the anamnesis and the physical examination in each consultation during all the follow-up period.

The radiological examination is a less sensible method and more expensive than ECG, so it should be performed once before treatment, 6 and 12 months later and then once per year of monitoring. This examination should consist of a chest RX with postero-anterior and lateral views with esophagus contrasted immediately after ingestion of barium and also after 1 min for the evaluation of the time necessary to drain the contrast (Rezende et al. 1960). The barium enema with previous preparation and radiography of colon is the only test capable of evaluating an established megacolon (Rezende 1959).

The dynamic electrocardiography and the echocardiography are suitable techniques for evaluation of arrhythmias and anatomophysiology of the heart and must be performed by a cardiologist. In the same way, endoscopic and manometric tests need an experienced gastroenterologist. The examination of the autonomous and peripheric nervous systems, with or without stimulation with cholinergic drugs, such as pilocarpine, can be done by a physician (Macêdo 1997).

The histopathology study of biopsies fragments of the digestive system or endomyocardic with conventional microscopy or analysis by immunoperoxidase or immunofluorescence are indicated only in some research cases and could only be performed following strict protocols from the ethics point of view.

### New drugs in clinical tests

For several years in Brazil, and more recently in Argentina, Chile and Uruguay, only Bz is commercially available as the development of drugs for tropical diseases is of little interest for the pharmaceutical industry (Fairlamb 1999). After the introduction of Nif and Bz, few compounds were assayed in chagasic patients.

#### Allopurinol

The results obtained with allopurinol (4-hydroxyxypyrazolo (3,4-*d*) pyrimidine HPP, Fig. 1c) in experimental animals and the knowledge about its mode of action led to its clinical assays for the treatment of Chagas disease. This compound is a hypoxanthine analog that acts as an alternative substrate of hypoxanthine-guanine phosphoribosyltransferase (HGPRT) and is incorporated into the RNA. This incorporation leads to formation of non-physiological nucleotides and to blockade of the synthesis *de novo* of purine nucleotides (reviewed in Marr 1991).

In the treatment of six acute phase patients, allopurinol was ineffective, with maintenance of positive xenodiagnosis and serology (Lauria-Pires et al. 1988). In a study, with chronic patients, Galleano et al. (1990) treated two groups of patients with 600 and 900 mg/kg/day of allopurinol for 60 days, compared with other two treated with Nif and Bz. In the four groups the percentage of negativation of xenodiagnosis was in the range of 75-92%, and those treated with allopurinol presented less collateral effects. Allopurinol (600 mg/day for 2 months) was administered to two cases of reactivation of Chagas disease due to cardiac transplant. Erythematous lesions on the superior and/or inferior members characterized this reactivation. In one patient the lesions disappeared in 3 weeks, and in the sec-

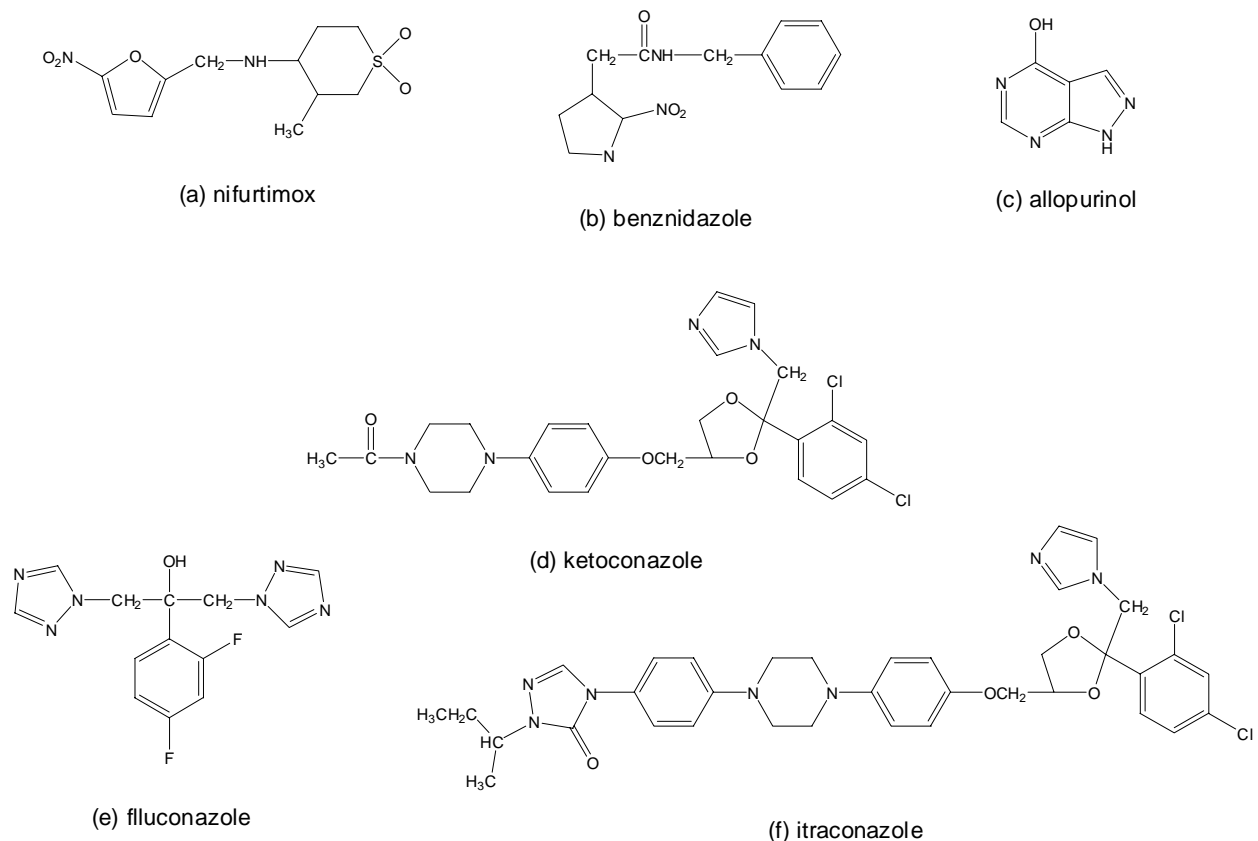


Fig. 1: structure of compounds used in the treatment of Chagas disease after the 70s.

ond one, after 2 weeks there was a clinical improvement of the lesions. In both cases after treatment, xenodiagnosis and hemoculture tests were negative and in the follow-up of 38 and 17 months, respectively, no reactivation of Chagas disease occurred, even with continued immunosuppression (Tomimori-Yamashita et al. 1997). Apt et al. (1998) treated 104 chronic patients with allopurinol (8.5 mg/kg/day for 60 days) that were monitored by clinical examination, serology, xenodiagnosis, hemoculture and electrocardiogram. Parasitological cure was achieved in 44% of the allopurinol-treated patients. The criteria for parasitological cure were maintenance of negative xenodiagnosis and/or complement-mediated lysis for at least four years. A double blind randomized longitudinal study must be performed to reevaluate the efficacy of this drug for the treatment of Chagas disease.

#### Ketoconazole

In eight chronic patients, ketoconazole (*cis-(dl)*-1-acetyl-4-[4-[(2,2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-ioxolan-4-yl]methoxy]-phenyl]-piperazine, Fig. 1d) was administered in doses between 3.1 and 8.7 mg/kg by oral route during 51 to 96 days and cure evaluation was performed by hemoculture, conventional serology and complement-mediated lysis. The patients were monitored up to 60 months and it was observed that the drug was unable to eradicate the parasites, from 6 out of 8 patients with positive hemoculture and two others with positive serology (Brener et al. 1993).

In a case of reactivation of Chagas disease in a patient in the indeterminate phase due to infection by HIV, ketoconazole (400 mg/day) was administered for 70 days leading to a negative xenodiagnosis. The treatment was suspended by the patient's own decision and after one month occurred signs of reactivation of the disease, including development of myocarditis. Bz was introduced (200 mg/day) and after four days, although negativation of parasitemia, the patient presented signs of neurological deterioration. This drug was maintained for 45 days and then replaced by ketoconazole (400 mg/day) as a suppressive treatment. However, after a general clinical improvement, there was a new neurologic deterioration and the patient died (Galhardo et al. 1999).

Ketoconazole was one of the first imidazoles that showed in vitro activity against *T. cruzi*, with accumulation of metabolites of sterol metabolism in epimastigotes. In vivo ketoconazole led to parasitological cure in experimental animal in the acute phase, but was ineffective in the chronic phase (reviewed in De Castro 1993). A synergic effect of ketoconazole and Bz was observed in mice infected with the CL or Y strain, what did not occur in the case of the Bz-resistant Colombian strain (Araújo et al. 2000).

#### Fluconazole and itraconazole

An haemophylic patient was infected by blood transfusion with HIV and *T. cruzi* and brain biopsy revealed the presence of amastigotes inside glial, macrophages and endothelial cells. Initially he was treated with Bz (400 mg/day) but due to general worsening of his clinical condition, the medication was changed to itraconazole (200 mg/day), and latter, aiming a better CNS penetration, to fluconazole (400 mg/day). The azoles were administered for 11 weeks, and during this period the fever resolved and neurological symptoms stabilized. No significant collateral reaction was observed and three months after treat-

ment the xenodiagnosis was negative and the titer of indirect hemagglutination test was 1:16 (Solari et al. 1993). Following the same methodology described for the treatment with allopurinol, Apt et al. (1998) treated 135 chronic patients with itraconazole (6 mg/kg/day for 120 days) observing parasitological cure and normalization of ECG in 36.5% of the treated patients but new abnormalities of the ECG appeared in 48.2% after treatment. As in the case of allopurinol these azoles must be further investigated in a well-designed protocol for treatment of chagasic patients.

The azoles fluconazole ( $\alpha$ -(2,4-difluorophenyl)- $\alpha$ -(1H-1,2,4-triazol-1-ylmethyl)-1H-1,2,4-triazol-1-ethanol, Fig. 1e) and itraconazole (*cis*-4-[4-4-[[2-(2,4-dichlorophenyl)-2-(1H-1,2,4-triazol-1-methyl)-1,3-dioxolan-4-yl]-1-piperazinyl]phenyl]-2,4-dihydro-2-(1-methyl-propyl)-3H-1,2,4-triazol-3-one, Fig. 1f) have been previously assayed in experimental animals, and their mechanism of action against *T. cruzi* involve interference on ergosterol synthesis (reviewed in De Castro 1993). A more recent study showed potent effect of the D(+) isomer of fluconazole – compound D0870 – in both acute and chronic mice models, with 30-50 times higher activity than ketoconazole and Nif and leading to 60-70% of parasitological cure (Urbina et al. 1996). A formulation of D0870 as loaded nanospheres administered by intravenous route to mice, showed also a significant cure rate (Molina et al. 2001). It is important to stress that this compound, based on several cure parameters was also active in a chronic phase model. We believe that the pharmacokinetics of D0870 is now being investigated.

#### DEVELOPMENT OF NEW DRUGS

Development of anti-parasite chemotherapy could emerge from screening of synthetic or natural libraries, of compounds with structural similarities, with a drug with recognized activity, of assays with agents already approved for other diseases or through the determination of a specific target, identified in a key metabolic pathway. Although several putative targets have been presented, there is a need for their validation. The criteria for such validation was discussed by Wang (1997), who suggested that preliminary verifications can be indicated by in vitro activity of an inhibitor of the putative target, but before a major effort is directed to the design of specific inhibitors, three approaches should be used: (i) correlation of the target inhibition and anti-parasite activity among a series of drug derivatives; (ii) the comparison of the target between drug-sensitive and drug-resistant parasites; (iii) the knock out of the gene encoding such target in the parasite. In this paper Wang (1997) also pointed out the inherent difficulties of such approaches of target validation.

#### Promising targets

Recent developments in the study of the basic biochemistry of *T. cruzi* have allowed the identification of novel targets for chemotherapy that include sterol metabolism, enzymes such as trypanothione reductase, cystein proteinase, hypoxanthine-guanine phosphoribosyltransferase, glyceraldehyde-3-phosphate dehydrogenase, DNA topoisomerases, dihydrofolate reductase and farnesylpyrophosphate synthase (reviewed in DoCampo 2001 and in Rodriguez 2001).

#### Sterol synthesis

The knowledge about sterol synthesis on fungi opened the possibility of interference in this pathway, leading sev-



eral pharmaceutical companies to develop drugs for the treatment of different types of superficial mycosis and systemic fungal infections. Since the main sterol of *T. cruzi* is ergosterol, an intensive and fruitful investigation about the potential effect of inhibitors of this sterol, especially by the group of Urbina in Venezuela. Parallel to clinical studies with azole derivatives, the trypanocidal activity and mechanism of action of new compounds is under intensive investigation.

The triazole posaconazole (SCH56592, Schering-Plough), inhibited epimastigote proliferation and ergosterol synthesis at levels 30 to 100 times higher than ketoconazole and D0870. In experimental infections, this compound led to a cure rate of 50% in animals infected with strains resistant to Nif, Bz and ketoconazole (Molina et al. 2000). Another triazole derivative UR-9825 was very active against epimastigotes and intracellular amastigotes. At the minimum inhibitory concentration for epimastigotes occurred also depletion of 4,14-desmethyl endogenous sterols, such as ergosterol, and their replacement by methylated sterols, indicating inhibition of C14- $\alpha$  demethylase, as previously reported for other azoles. This drug induced also alteration in the phospholipid profile of the parasite (Urbina et al. 2000).

The induction of resistance of *T. cruzi* to azoles, such as fluconazole, and also the cross resistance between ketoconazole, miconazole and itraconazole, observed in in vitro experiments point to difficulties in the use of such compounds as chemotherapeutic agents (Buckner et al. 1998). In a subsequent work, Buckner et al. (2001) reported the development of inhibitors of a key enzyme in sterol biosynthesis, oxidosqualene cyclase, which converts 2,3-oxidosqualene to lanosterol. The lead compound, N-(4E,8E)-5,9, 13-trimethyl-4,8, 12-tetradecatrien-1-ylpyridinium, was shown to cause an accumulation of oxidosqualene and decreased production of lanosterol and ergosterol in *T. cruzi*. This compound and 27 related derivatives were tested against *T. cruzi*, and 12 of them were highly active against trypomastigotes.

#### Trypanothione reductase

Trypanosomatids present trypanothione (N<sup>1</sup>,N<sup>8</sup>-bis(glutathionyl)spermidine) and of specific enzymes for this cofactor, trypanothione reductase (TR) and trypanothione oxidase (reviewed in Fairlamb & Cerami 1992). TR is an NADPH-dependent flavoprotein that maintains trypanothione in its reduced form and able to be oxidized by trypanothione oxidase, leading to reduction of free radicals levels and contributing to the maintenance of an intracellular reducing environment. TR has been used as a target for rational drug design against trypanosomiasis and leishmaniasis in a number of laboratories, since this enzyme and the mammalian counterpart (glutathione peroxidase/glutathione reductase system) differ on the substrate specificity (reviewed in Augustyns et al. 2001). The determination of the structure of the active center of TR (Krauth-Siegel et al. 1987) allowed the search of inhibitors of this enzyme, being assayed different classes of compounds. In most cases the studies analyzed the effect of a putative inhibitor on the purified enzyme, and depending on the results obtained, new compounds based on molecular modeling were developed. A first group of inhibitors reported were the so-called "subversive substrates", due to the futile-cycling of TR induced by redox-damaging drugs, such as nitrofurans, and naphthoquinones

(Henderson et al. 1988, Salmon-Chemin et al. 2001). Subsequently the structure of tricyclic neuroleptic showed to be a promising backbone class of TR inhibitors, and based on computational design techniques several tricyclic compounds were investigated (Chan et al. 1998, Gutierrez-Correa et al. 2001). Some compounds of the series of 2-amino diphenylsulfides, that have lower neuroleptic activity than phenothiazines, were potent inhibitors of TR (Girault et al. 1998). Polyamine derivatives (Bonnet et al. 1997, Li et al. 2001), bisbenzylisoquinoline alkaloids (Fournet et al. 1998) and platinum II complexes (Bonse et al. 2000) were also studied in their capacity of inhibiting TR of *T. cruzi*.

#### Cystein protease

Cruzipain, also known as cruzain or GP57/51, is a member of the papain C1 family of cystein proteinases (CPs). The *T. cruzi* enzyme consists of a catalytic moiety with high homology to cathepsins S and L, and is absent in all other C1 families described so far (reviewed in Cazzulo et al. 2001). Irreversible inhibitors of cruzipain, such as several peptidyl diazomethylketones, peptidyl fluoromethylketones and peptidyl vinyl sulphones interfered with the in vitro intracellular cycle of *T. cruzi*, killing the parasite (reviewed in McKerrow 1999).

The treatment of acutely infected mice with the vinyl sulphone N-piperazine-Phe-hPhe-vinyl sulphone phenyl led to the absence of myocardial lesions, lymphocyte infiltration and intracellular amastigote clusters. This drug kills *T. cruzi* by inducing an accumulation of unprocessed cruzipain in the Golgi cisternae, interfering with the secretory pathway (Engel et al. 1998a,b). Cruzipain exposed to biotin-labelled peptidyl diazomethane inhibitors with a spacer arm showed a stronger reaction than the counterparts without such spacer, probably due to differences in the topologies of the binding sites of proteinases, differences that could be exploited to improve specificity against trypanosomal CP (Lalmanach et al. 1996). Roush et al. (2000) substituting the L-leucine residue of the natural peptidylepoxysuccinate E-64, a selective irreversible inhibitor of CP, by a D-threonine obtained a derivative with much higher activity against cruzipain than against bovine cathepsin B. Yong et al. (2000) commented that a possible limitation of CP as a target would be the emergence of parasite populations developing resistance to inhibitors. These authors reported a phenotypically stable *T. cruzi* cell line (R-Dm28) that displays increased resistance to Z-(SBz)Cys-Phe-CHN<sub>2</sub>, an irreversible cysteine proteinase inhibitor, which preferentially inactivates cathepsin L-like enzymes.

#### Hypoxanthine-guanine phosphoribosyltransferase

Trypanosomatids must rely upon the salvage of exogenous purines for nucleotide synthesis, while in mammals these nucleotides are synthesized both *de novo* and salvaged from recycled purine bases. These protozoa convert purine bases to ribonucleotides, by the single enzyme HGPRT. This enzyme can also initiate in these parasites the metabolism of certain cytotoxic purine base analogs, such as allopurinol. This implies that either inhibitors or substrates of HGPRT have the potential of being effective and selective chemotherapeutic agents. The hgpRT genes from *T. cruzi* and other pathogenic trypanosomatids have been cloned, sequenced and overexpressed in *Escherichia coli*, and the recombinant proteins have all been purified and characterized (reviewed in Ullman & Carter 1997).

The purine analogs 3'-deoxyinosine, 3'-deoxyadenosine and allopurinol inhibited the proliferation of amastigotes in HeLa cells, being the latter the most active. Among the pyrimidine analogs, 3'-azido-3'-deoxythymidine showed high activity against *T. cruzi* (Nakajima-Shimada et al. 1996). Purine analogs were assayed for their interaction with the HGPRTs from *T. cruzi* and man and some of them showed affinity for the trypanosomal enzyme (Eakin et al. 1997). A structure-based docking method identified 22 potential inhibitors of the enzyme. Three compounds (2,4,7-trinitro-9-fluorenyl-idenemalononitrite, 3-(2-fluorophenyl)-5-(phenoxy)-1,2,4-triazolo (4,3-C)-quinazoline and 3,5-diphenyl-4'-methyl-2-nitrobiphenyl) were effective against intracellular amastigotes and one [6-(2,2-dichloro-acetamido)chrysene] was a potent inhibitor of the trypanosomal HPRT (Freyman et al. 2000).

#### DNA topoisomerases

DNA topoisomerases II are enzymes that alter the topology of DNA and in kinetoplastids have been the focus of considerable study in the areas of molecular and cellular biology and also experimental chemotherapy. The gene encoding *T. cruzi* type II topoisomerase was isolated and the comparison with the amino acid sequence of the corresponding enzymes of *T. brucei* and *Crithidia fasciculata* showed a high degree of conservation (Fragoso & Goldenberg 1992). The enzyme is expressed in epimastigotes but not in trypomastigotes, although both forms of the parasite present the mRNA encoding the enzyme and is localized exclusively in the nucleus of the parasite (Fragoso et al. 1998).

Several inhibitors of bacterial DNA topoisomerase II showed activity against *T. cruzi*, inhibiting both proliferation and differentiation processes, and causing damage to kinetoplast and/or the nucleus of epimastigotes (Kerschmann et al. 1989, Gonzales-Perdomo et al. 1990), suggesting that both organelles could be the targets of the drugs. Camptothecin, inhibitor of eukaryotic DNA topoisomerase I, induced cleavage of nuclear and mitochondrial DNA in *T. cruzi* (Bodley & Shapiro 1995).

#### Dihydrofolate reductase

Dihydrofolate reductase (DHFR) and thymidylate synthetase exist as a bifunctional protein in different species of protozoa. This enzyme has successfully been used as a drug target in chemotherapy of cancer, malaria and bacterial infections. The gene coding for the DHFR domain from *T. cruzi* was cloned and expressed (Reche et al. 1996). Zucotto et al. (1998) described the modelling of *T. cruzi*'s DHFR based on the crystal structure of *Leishmania major* enzyme. From methotrexate, inhibitor of the human enzyme, among several derivatives synthesized, some of them showed a greater selectivity for the parasite enzyme than for the human counterpart (Zucotto et al. 1999). In the same line, Chowdhury et al. (2001) designed and synthesized novel inhibitors of DHFR of trypanosomatids, however the compounds showed weak activity against both the enzyme and intracellular amastigotes of *T. cruzi*.

#### Glyceraldehyde-3-phosphate dehydrogenase

Since intracellular amastigotes possibly derive its energy from glycolysis, inhibition of glycolytic enzymes such as glyceraldehyde-3-phosphate dehydrogenase (GAPDH) may be a novel approach for the development of anti-*T. cruzi* drugs. The structure of GAPDH from glycosomes was reported and comparison with that of the mammalian counterpart led to the group of Oliva to consider the pos-

sibility of development of specific inhibitors of the parasite enzyme (Souza et al. 1998). In a subsequent work the isolation of flavonoids from the fruits of *Neoraptua magnifica* led to the compound 3',4',5',7-pentamethoxyflavone that showed the highest activity over flavones and pyrano chalcones against the GAPDH of the parasite (Tomazela et al. 2000). Crystal structure of trypanosomatids and human GAPDHs provided details about the interaction of adenosyl moiety of NAD<sup>+</sup> with proteins. Although adenosine is a very poor inhibitor, addition of substituents to the 2' position of ribose and the N6-position of adenosine led to a series of disubstituted nucleosides, and [N6-(1-naphthalenemethyl)-2'-(3-chlorobenzamido) adenosine] inhibited the proliferation of amastigotes without effect on the corresponding human enzyme (Bressi et al. 2001).

#### Farnesylpyrophosphate synthase

In pathogenic protozoa the pathway responsible for the synthesis of a variety of sterols and polyisoprenoids involves the enzyme farnesylpyrophosphate synthase, leading to the formation of farnesylpyrophosphate that marks the branching point of these synthetic routes. A gene encoding the farnesylpyrophosphate synthase of *T. cruzi* (TcFPPS) was cloned and sequenced and the enzyme was inhibited by the nitrogen-containing bisphosphonates, such as pamidronate and risedronate, but was less sensitive to the non-nitrogen-containing bisphosphonate etidronate, which does not affect parasite growth (Montalvetti et al. 2001). Pamidronate caused a decrease in the parasitemia of *T. cruzi*-infected mice and inhibited the in vitro intracellular replication of amastigotes (Urbina et al. 1999). Risedronate inhibited the proliferation of epimastigotes and sterol biosynthesis at a pre-squalene level and based on sterols analysis in treated parasites Martin et al. (2001) associated these results with the inhibition of farnesylpyrophosphate synthase. The effect of a series of bisphosphonates derived from fatty acids were assayed against *T. cruzi* and some of these drugs were potent inhibitors of the proliferation of intracellular amastigotes, but all of them were devoid of activity against epimastigotes (Szajnman et al. 2000). The selective action of nitrogen-containing bisphosphonates against *T. cruzi* in comparison to mammalian cells could result from the preferential drug accumulation in parasite acidocalcisomes, acidic organelles rich in calcium, pyrophosphate, magnesium, sodium, zinc and polyphosphates (reviewed in DoCampo & Moreno 2001).

#### Experimentally assayed drugs after 1992/1993

The present review offers a survey of the available literature about new classes of compounds and also new derivatives from compounds previously assayed in the search for new drugs against *T. cruzi*. Recently other reviews have also been published, most of them exploring selected groups of compounds or inhibitors for *T. cruzi* targets (Rodriguez & Gros 1995, Urbina 1999, DoCampo 2001, Rodriguez 2001). As we have already mentioned with the present review, together with those of Brener (1979) and De Castro (1993), we intend to give the reader a general view of the experimental studies in the area of the chemotherapy of Chagas disease.

#### Synthetic drugs

*Thiadiazine derivatives* - In assays with epimastigotes, most of the 1,3,5-thiadiazine-2-thione derivatives were more active than Nif, while among 1,2,6-thiadiazin-3,5-dione 1,1-

dioxides; although active against the parasite, most of them were also toxic to mammalian cells (Ochoa et al. 1999, Di Maio et al. 1999). Among the less cytotoxic derivatives of the second series, one compound showed activity against intracellular amastigotes similar to the standard drug (Muelas et al. 2001).

*1,2,5-oxadiazole N-oxide derivatives* - A series of 1,2,5-oxadiazole N-oxide, benzo[1,2-c]1,2,5-oxadiazole N-oxide, and of quinoxaline di-N-oxide derivatives were synthesized and the activity against epimastigotes was associated with N-oxide radical formation (Cerecetto et al. 1999).

*1,4-dihydropyridines* - Among nitro-aryl-1,4-dihydropyridines, nicardipine, isradipine and lacidipine inhibited epimastigote proliferation and oxygen uptake in intact parasite, and the first compound showed also a similar effect in mitochondria *in situ* (Nunes-Vergara et al. 1997). For a series of 3-chloro-phenyl-1,4-dihydropyridine derivatives a positive correlation between trypanocidal effect and easiness of oxidation of the dihydropyridine ring was found (Maya et al. 2000).

*Acridine derivatives* - Since the 80s several acridine and acridinone derivatives have been presented a correlation of activity against epimastigotes and DNA binding (reviewed in De Castro 1993). Among a series of *bis*(9-amino-6-chloro-2-methoxyacridines), a bisacridine containing piperazine as central amine showed co-localization with kDNA of epimastigotes (Girault et al. 2000), reinforcing that their activity is associated with DNA interaction. Several 9-thioalkylacridines were active against *T. cruzi* (Bsiri et al. 1996). 9-Amino and 9-thioacridines have been reported to inhibit the enzyme TR (Bonse et al. 2000).

*Nitroimidazoles and nitrofurans derivatives* - Several nitroimidazoles such as megalol (CL 64,855), MK-436 and fexinidazole presented high activity in infected animals (reviewed in De Castro 1993). The current investigation about the use of nitroimidazoles for therapy of African trypanosomiasis, especially megalol (Barrett et al. 2000), renewed the interest in the area of Chagas disease. The coupling of 5-chloro-4-nitro-1-methylimidazole with heterocycles led to the synthesis of two compounds with activity against trypomastigotes (Boechat et al. 2001). More recently Cerecetto et al. (1999) synthesized nitrofurazones (5-nitro-2-furaldehyde semicarbazones) and typhenes (5-nitrothiophene-2-carboxaldehyde) in which N<sup>4</sup>-semicarbazone moiety was replaced by different of amines, aiming to mimic the spermidine part of trypanothione. These compounds presented lower activity against epimastigotes than the parent compound, while some nitrofurazones bearing N<sup>4</sup> other substituents produced complete survival in infected mice (Cerecetto et al. 2000).

*Phenothiazines* - Before the period mentioned in this review several groups investigated the effect of phenothiazines, tricyclic compounds employed clinically as antidepressants (reviewed in De Castro 1993). More recently it was reported that this class inhibited the enzyme TR. Promethazine and thioridazine assayed *in vivo* decreased parasitemia levels and mortality (Paglini-Oliva et al. 1998). When mice infected with low inoculum of *T. cruzi* and treated with thioridazine were checked 135 days post-infection, the heart histology and density of cardiac  $\beta$ -receptors were similar to those of uninfected, untreated controls (Rivarola et al. 1999), suggesting to the authors that this drug could prevent the evolution to the chronic phase of the infection.

*Metal chelating agents and metallic complexes* - Several chelating agents and derivatives were active against epimastigotes; being proposed that they act by interfering with the essential metabolism of iron, copper, or zinc (Rodrigues et al. 1995). Several Fe<sup>3+</sup> chelating agents showed activity against epimastigotes, that could be decrease in the presence of iron (Jones et al. 1993, Singh et al. 1997), and tetraethyl derivative of aminothiols was active against trypomastigotes (Deharo et al. 2000). The metal chelator sodium diethylamine-N-carbodithioate showed, in relation to Bz a similar activity against epimastigotes and intracellular amastigotes but lower activity against trypomastigotes (Lane et al. 1996). Another chelator, 1,10-phenanthroline inhibited epimastigote proliferation and led to electron-dense deposits in the kinetoplast, mitochondrion, and endoplasmic reticulum, identified as containing predominantly calcium and suggesting to the authors the involvement of disruption of calcium homeostasis in the trypanocidal activity (Lane et al. 1998).

Among several osmium(III) complexed with carbamates and different metals complexed with 1,2,4]triazolo [1,5a]pyrimidines some compounds were active against epimastigotes (Castilla et al. 1996, Luque et al. 2000).

*Propene-1-amine derivatives* - A series of 3-(4'-bromo-[1,1'-biphenyl]-4-yl)-3-(4-X-phenyl)-N,N-dimethyl-2-propen-1-amine derivatives were active against the three forms of *T. cruzi* (De Conti et al. 1996, Oliveira et al. 1999). The two compounds - the unsubstituted (X = H) and the bromine (X = Br) analogs - with highest activity against bloodstream forms, and lowest toxicity to mammalian cells, were assayed *in vivo*. The bromo-derivative displayed a strong suppressive effect on the parasitemia, and led to the survival of all the treated mice, whereas its unsubstituted analog was ineffective under the same conditions (Pereira et al. 1998).

*Aminoquinoline derivatives* - Quinolines have been assayed as potential drugs for Chagas disease since the 50s (reviewed in Brener 1979). Against epimastigotes the action of primaquine involves the formation of free radicals and this drug presented synergistic effect with ketoconazole decreasing the parasitemia of experimentally infected mice (reviewed in De Castro 1993). More recently, dipeptide derivatives of this aminoquinoline were synthesized as prodrugs, and inhibited the infection of LLC-MK2 cells with *T. cruzi* (Chung et al. 1997). Among 77 primaquine analogues, one of them reduced the parasitemia in mice in levels 14 and 4 times higher than Nif and primaquine, respectively (Kinnamon et al. 1996). In the same model, among 40 8-aminoquinolines, non-related to primaquine, 6 were more active than Nif, and for one of them, the activity was 13 times higher than this standard drug (Kinnamon et al. 1997).

*Dinitroanilines* - These compounds are microtubule-disrupting herbicides and one of the most studied is trifluralin. This compound was active against epimastigotes, trypomastigotes, and axenic and intracellular amastigotes. In trypomastigotes, trifluralin led to alterations at the surface analyzed by transmission and scanning electron microscopy, with membrane waving not associated with subpellicular microtubules. Treated epimastigotes showed alterations of shape, and some parasites presented two or three flagella and kinetoplasts, suggesting interruption of the cytokinesis process. Trifluralin also inhibited endocytosis in epimastigotes, monitored by complexes of gold with bovine serum albumine (Dantas et al. 1998, Dantas

2000). This dinitroaniline presented also in vivo effect in mice model (Zaidenberg et al. 1999) and inhibited the differentiation of epimastigotes to trypomastigotes (Bogitsh et al. 1999).

Recently we reviewed the effect of dinitroanilines on pathogenic protozoa and observed that *Leishmania* spp. and *Trypanosoma brucei* were more susceptible to trifluralin than *T. cruzi* and also compared sequences of tubulins of susceptible organisms (plants and trypanosomatids) and resistant (mammals) (Traub-Cseko et al. 2001).

*Lysophospholipid analogs* - These compounds are under clinical studies for cancer chemotherapy. The most studied derivatives are the alkylglycerophosphocholine edelfosine, a thioether substituted phosphatidylcholine analog ilmofosine and the alkylphosphocholine hexadecylphosphocholine (mitelfosine) (reviewed in Wieder et al. 1999). Clinical trials with mitelfosine for the treatment of Indian visceral leishmaniasis gave high percentage of cure including in cases resistance antimony therapy (Sundar et al. 2000). These LPAs were active against epimastigotes, intracellular amastigotes and trypomastigotes and led to damage of the flagellar membrane of epimastigote, and edelfosine inhibited the in vitro metacyclogenesis process (Santa-Rita et al. 2000). Previous in vivo experiments showed that LPAs had only a suppressive effect on the parasitaemia of *T. cruzi*-infected mice (Croft et al. 1996). It was found that LPAs are potent inhibitors of phosphatidylcholine synthesis in epimastigotes, and that ergosterol and its 24-ethyl analog were replaced by its  $\Delta^{22}$ -saturated analogs, indicating inhibition of sterol C-22 desaturase (Lira et al. 2001).

#### Drugs derived from natural sources

Natural products account for about half of the pharmaceuticals in use today, but there has been a shift away from their use with the increasing predominance of molecular approaches to drug discovery (Clark 1996). However, in the last decades occurred a movement named by some as "back to Nature", with the revival of phytotherapy, needing a multidisciplinary group with expertise in botany, chemistry, biochemistry, molecular and cellular biology, and pharmacology. This trend was accentuated by the success obtained, for example, by taxol in cancer chemotherapy and artemisinin for malaria. A complementary approach is the development of synthetic analogs of the lead natural compound, aiming the improvement of properties such as pharmacokinetics, compatibilities and stability. Such intensification in the search of drugs from natural sources was also observed in the area of trypanosomiasis and leishmaniasis. In the literature several recent reports deal with the investigation of trypanocidal activity of a wide variety of crude natural extracts, especially vegetal ones, compounds isolated and semi-synthetic analogs.

*Alkaloids* - A great number of alkaloids have been assayed against *T. cruzi*. The activity of a series of alkaloids against epimastigotes was associated to the inhibition of cell respiration, and the most active compound was apomorphine (Morello et al. 1994). Using epimastigotes of Nif-resistant, the activity of  $\beta$ -carboline alkaloids was also associated with respiratory chain (Rivas et al. 1999). Several glycoalkaloids were tested against epimastigotes, bloodstream and metacyclic trypomastigotes, and aconitine and a-solamargine showed higher activity than ketoconazole (Chataing et al. 1998). Another group studied was bisbenzylisoquinoline alkaloids and daphnoline

and cepharanthine were active against the parasite and inhibited enzyme TR (Fournet et al. 1998). In acute infections, daphnoline led to a significant decrease in parasitemia and increase in cure rate in comparison with Bz-treated mice, and in chronic infections, in 70% of the treated mice no parasite was detected (Fournet et al. 2000). Five new bisbenzylisoquinoline derivatives were isolated from the stem bark of *Guatteria boliviana* and funiferine, antioquine and guatteboline were active against trypomastigotes (Mahiou et al. 2000). The alkaloid anti-microtubule agents vinblastine and vincristine obtained from *Vinca rosea* interfere with the proliferation of epimastigotes, inhibiting both nuclear division and cytokinesis, leading to giant cells with multiple nuclei, kinetoplasts and flagella (Grellier et al. 1999).

*Taxoids* - Taxol obtained from the bark of *Taxus brevifolia* is an anti-microtubule drug. This compound and synthetic derivatives are employed in cancer chemotherapy. Taxol was active against epimastigotes and trypomastigotes leading to significant alterations in the morphology of the parasites. Taxol also inhibited the endocytosis of proteins by epimastigotes (Dantas 2000).

*Stilbenoids and derivatives* - Among natural dihydrostilbenoid isonotholaenic acid and several simple derivatives some compounds showed activity similar to Bz against epimastigotes and others were more active than crystal violet against trypomastigotes (Olmo et al. 2001).

*Snake venom* - Venom from *Cerastes cerastes* and *Naja haje* inhibited the proliferation of epimastigotes of *T. cruzi* at levels similar to Bz, and that of *C. cerastes* was also active against trypomastigotes (Fernandez-Gomez et al. 1994).

*Gangliosides* Treatment with gangliosides of mice during acute infection promoted survival and clearance of parasites from the bloodstream and organs, and it was suggested that the effect of gangliosides could be due to interference of parasite penetration into the host cells due to inhibition of phospholipase A<sub>2</sub> (Lujan et al. 1993), but since the compounds had no direct effect on the parasite, Bronia et al. (1999) consider that the in vivo effect could be due to modulation of the host immune system.

*Juvenile hormone inhibitors and analogs* - The juvenile hormone fenoxycarb and methoprene inhibited the proliferation of epimastigotes (Stoka et al. 1995). Fenoxycarb and analogs were synthesized, and assays with epimastigotes suggested that an allyl ether moiety bonded at the polar extreme is important for the trypanocidal effect (Cinque et al. 1998). Some analogs were also active against bloodstream forms and reduced the parasitemia and mortality levels in relation to untreated controls (Fichera et al. 1995). 4-Phenoxyphenoxyethyl thiocyanate was very active against epimastigote and accumulation of low molecular weight metabolites from mevalonate to squalene was observed (Szajnman et al. 2000), leading the authors to suggest that the effect is associated with interference in the synthesis of ergosterol. Also sulfur-containing derivatives showed high activity against epimastigotes and amastigotes of *T. cruzi* (Rodriguez et al. 2000).

*Propolis* - It is a natural resin produced by honey bees has been used in folk medicine, since it displays strong anti-microbial activity, associated mainly with flavonoids and derivatives of hydroxycinnamic acid (reviewed in De Castro 2001). Ethanolic extract prepared from a North American commercial sample were active in vitro against epimastigotes, trypomastigotes and intracellular

amastigotes (Higashi & De Castro 1994). However, in vivo experiments using different propolis formulations, showed no effect on the course of acute infection (De Castro & Higashi 1995). A Bulgarian sample has been assayed against trypomastigotes, and the ethanolic and acetic extracts showed activity, respectively, similar and two times higher than crystal violet. The chemical composition of both extracts showed a high content of flavonoids, with differences in relative individual concentrations (De Castro et al. 2001). Recently Brazilian propolis became a subject of increasing scientific and commercial interest, and due to significant differences between the composition samples from tropical and temperate zones (reviewed in Bankova et al. 2000), we isolated from a sample collected in the State of Paraná (Brazil) four derivatives of hydroxycinnamic acid than when assayed against trypomastigotes showed activity below that of crystal violet (Marcucci et al. 2001).

*Naphthoquinones and synthetic derivatives* - Quinones are present in different families of plants and in some of them have been used in folk medicine for the treatment of diseases, especially cancer. Among bioactive natural naphthoquinones, we found lapachol, isolated from the heartwood of *Tabebuia* ("ipês") and also  $\alpha$ - and  $\beta$ -lapachone obtained as contaminants in the process of lapachol isolation. Recently, lapachol derivatives were assayed against trypomastigotes and the triacetoxyl derivative of reduced lapachol showed relevant trypanocidal activity (Santos et al. 2001). Due to the increasing number of reports about the activity of  $\beta$ -lapachone against a wide variety of tumor cells, by inhibition of topoisomerases and induction of apoptosis, Dubin et al. (2001) pointed out the potential of clinical applications of this naphthoquinone. Previous studies with  $\beta$ -lapachone showed activity against epimastigotes that was associated to generation of free radicals and inhibition of nucleic acids and protein synthesis (reviewed in De Moura et al. 2001). In a study of naphthoquinones isolated from *Tabebuia*, we synthesized 50 heterocyclic derivatives. The overall analysis of the structures indicated the tendency of trypanocidal activity in compounds with an imidazole or oxazole ring linked to a naphthopyrane structure (Pinto et al. 1997, Neves-Pinto et al. 2000). Among these compounds two naphthimidazoles showed activities against trypomastigotes was 14.5 and 34.8 times higher than the standard crystal violet and both have an aromatic moiety linked to the imidazole ring (reviewed in De Moura et al. 2001).

Other groups of quinones isolated from natural products were also assayed against *T. cruzi*: (a) the trihydroxylated anthraquinone purpurin, obtained from the roots of *Rubia tinctorum* (Rubiaceae), showed an activity against trypomastigotes 1.5 times higher than crystal violet (De Castro et al. 1994); (b) among 1,4-naphthoquinones isolated from *Calceolaria sessilis*, 2,3,3-trimethyl-2,3-dihydronaphtho[2,3-*b*]furan-4,9-quinone showed against epimastigote and tumor lineages a high cytotoxicity and a temporary increase of oxygen consumption (Morello et al. 1995); (c) the polyprenylated benzoquinone 7-epiclusianone, isolated from *Rheedia gardneriana* (Clusiaceae) was active *in vitro* against trypomastigote, but showed no effect on experimentally infected (Alves et al. 1999).

*Crude plant extracts and components* - (i) The sesquiterpene lactone dehydroleucodine isolated from *Artemisia douglasiana* was active against epimastigotes, that presented pycnotic nucleus and a decreased number of

ribosomes (Bregio et al. 2000); (ii) cryptofolione isolated from the fruits of *Cryptocarya alba* was active against trypomastigotes, however showed moderate effect for both amastigotes and macrophages, indicating little selectivity for *T. cruzi* (Schmeda-Hirschmann et al. 2001); (iii) from the seeds of *Annona glauca* (Annonaceae) nine acetogenins were isolated, and among them glaucanisin, squamocin, annonacin A and annonacin showed activity against trypomastigotes (Waechter et al. 1998); (iv) megalomicin is a macrolide antibiotic produced by *Micromonospora megalomicea*, that inhibits vesicular transport in Golgi apparatus, resulting in the undersialylation of cellular proteins in mammalian cells. This compound was active against epimastigotes and intracellular amastigotes at concentrations below those that interfere with the mammalian organelle (Bonay et al. 1998); (v) from the leaves of *Zanthoxylum naranjillo* (Rutaceae), the lignan, methylpluviatolide was highly effective *in vitro* and *in vivo* (Bastos et al. 1999); (vi) *Terpenes*: (a) among diterpenoids isolated from *Azorella compacta* (Llaretia) azorellanol and mulin-11,13-dien-20-oic acid were active against intracellular amastigotes and the cytotoxicity to mammalian cells was lower than that of Nif (Neira et al. 1998); (b) from the aerial parts of *Wedelia paludosa* (Asteraceae), the isolated diterpenes ent-kaur-16-en-19-oic acid, ent-kaur-9(11),16(17)-dien-19-oic acid and 3  $\alpha$ -angeloiloxo-ent-kaur-16-en-19-oic acid showed activity against trypomastigotes (Batista et al. 1999); (c) in a study in Bolivia, 14 plants used in folk medicine to treat cutaneous leishmaniasis, extracts from 53 medicinal plants used for other diseases and different parts from 43 plants were screened and some of these material showed activity against epimastigotes of different strains of *T. cruzi* (Fournet et al. 1994); (d) extracts used in popular medicine in Guatemala from several plants such as *Neurolaena lobata* (Asteraceae), *Tridax procumbens* (Asteraceae), *Petiveria alliacea* (Phytolaccaceae) and *Byrsonima crassifolia* (Malpighiaceae) showed high activity against trypomastigotes. *N. lobata* and *Solanum americanum* showed *in vitro* and *in vivo* trypanocidal activity (Caceres et al. 1998). In a subsequent work, extracts, fractions and isolated sesquiterpene lactones and germacranolides from *N. lobata* showed high activity against epimastigotes (Berger et al. 2001); (e) among 79 extracts from plants of the families Asteraceae, Araceae, Moraceae, Solanaceae, Rhamnaceae, Zingiberaceae, Leguminosae and Sapotaceae, nine of them were active against epimastigotes (Muelas-Serrano et al. 2000); (f) from 32 crude plant extracts of nine species of Rutaceae eight of them showed significant activity trypomastigote. Fractionation of the active extracts provided 25 fractions, and the two fractions more active were obtained from the leaves of *Almeidea coerulea* and from *Conchocarpus inopinatus* (Mafezoli et al. 2000).

### Prophylactic drugs

Although recent advances in vector control in the Southern Cone countries, by initiative of the Pan American Health Organization (PAHO) and World Health Organization (WHO), have decreased the incidence of new infections (Schofield & Dias 1999), we are still challenged by two critical problems: the treatment of chronic cases of the disease and the high level of acute cases in some Latin American countries, such as Bolivia and Mexico, where the incidence of infection in some regions reaches levels

above 80% of the population (Medrano-Mercado et al. 1996). In endemic areas the transfusional transmission of Chagas disease, due mainly to urbanization and migration processes, represents a great threat (Wendel & Dias 1992). For example, in 1990 in the USA, 7 million persons emigrated from countries in which Chagas disease is endemic (Schmunis, 1994, 2000) a preoccupying fact for the health authorities of non-endemic countries.

There is a recommendation from the WHO (1984) for the use of crystal violet in hemotherapeutic centers in endemic areas to eliminate the parasite in the blood used for transfusion (Nussenzweig et al. 1953). This dye presents no substantial side effect, although there are reports about blood micro-agglutination and potential mutagenicity (Thomas & McPhee 1984). Its main disadvantage is the bluish color that it confers to blood and tissues, which is not well accepted by the population. The development of trypanocidal drugs also involves the search for alternatives for crystal violet, when the assays were performed with trypomastigotes in the presence of blood. These studies are included in the above item, but it must be kept in mind that when looking for an alternative to crystal violet its is fundamental to perform the assays with bloodstream forms in the presence of blood and at 4°C. The presence of blood is essential, since several trypanocidal compounds that are inactivated by blood components, such as gossypol (Rovai et al. 1990) and naphthoquinones and their heterocyclic derivatives (Lopes et al. 1978, Pinto et al. 1997). Another problem associated with drugs for prophylaxis of blood is, besides eventual toxicity, the poor solubility in aqueous medium and binding to plasma protein.

#### CONCLUDING REMARKS

Nowadays infectious diseases are still an important cause of mortality and morbidity and the rising incidence of emerging or re-emerging diseases can be explained, at least partly, by the deterioration of health care systems and diverse socio-economic and ecological disorders. The cost of investments and the lack of market potential and market security in developing countries have dampened interest in developing drugs for tropical diseases. Among the 1061 new drugs developed from 1975 to 1994, less than 2.7% concern tropical diseases (Trouiller et al. 2000).

In relation to Chagas disease, great advances are being made in parts of South America to control the transmission by insect vectors or blood transfusion, but more effective chemotherapy is needed for the millions who are already infected. The demonstration of parasite in chronic patients indicates its importance in the maintenance of Chagas disease (reviewed in Tarleton 2001) and open the discussion about the etiologic treatment during this phase (OPAS/OMS 1998), reinforcing the need of finding more efficient and less toxic drugs.

In relation to experimental studies, a dichotomy has arisen between a rational and an empirical approach for drug development (Croft 1994). However, both are important routes to achieve a potential drug and sometimes are even used in conjunction. For example, having in hands a potential parasite target, the finding of an inhibitor could be searched by empirical tests or by modeling and synthesis that could interfere with the special regions of the target, such as the active center of an enzyme. The advent of genomics, rapid DNA sequencing, bioinformatics, combinatorial chemistry and automated high-throughput screenings had strength the interaction among groups with dif-

ferent expertise in order to find compounds with efficacy, including for immunosuppressed patients, without or at least with low toxicity, favorable pharmacokinetics and permeability properties and low cost of production.

Most of the studies with designed drugs against potential targets in *T. cruzi* involves inhibition tests using purified enzymes such as TR, CP and HGPRT, and it is possible that a compound discarded due to a negative result in this assay could be effective against the intact parasite. The continuous research of TR as a target could lead to an inhibitor that is a trypanocidal agent itself or that compromising the redox defenses of the parasite act in synergy with redox-damage drugs. However since the identification of trypanothione and although several groups are involved in finding inhibitors of TR, no drug emerged as effective and with low toxicity in experimentally infected animals.

After the introduction of Nif and Bz, among the extensive list of classes of compounds with in vitro and in vivo activity against *T. cruzi*, with exception of allopurinol, itraconazole and fluconazole, none was submitted to clinical assays. This is due in some cases, to the absence of strong indication of their curative effect, to their potential toxic and/or teratogenic effect (usually assayed only in vitro). This analysis emphasizes the need of a better choice of experimental models and standardization of protocols. In *in vitro* tests, it is of fundamental importance the use of trypomastigotes in the presence of blood both at 4°C (blood prophylaxis) and at 37°C (treatment) and comparison with crystal violet, and the effect against intracellular amastigotes using a suitable host cell and comparison with Nif or Bz. Still today several drugs are screened only against epimastigotes, although several authors have already reported different sensibilities to drugs among different forms of the parasite. In relation to in vivo tests a standardization of both acute and chronic phase models and of parameters to monitor the cure are of utmost importance. Also toxicity and mutagenicity studies in animals are needed. Otherwise we will continue to employ a lot of effort in experimental studies without achieving a drug that could be submitted to clinical trials.

After the presentation of potential targets in the parasite and active drugs in experimental models, which are the compounds that could potentially be employed in clinical tests? Among them stand out the nitroimidazoles megalol (CL-64855) (Filardi & Brener 1982) and MK-436 (Andrade et al. 1989), drugs studied before 1993, but that afterwards, at least to our knowledge, no new papers dealing with such promising drugs appeared in the literature. We believe that it could be important to review all the data obtained about megalol and MK-436, and develop new in vivo protocols to monitor their pharmacokinetics and bioavailability properties and also any potential toxic, mutagenic and/or teratogenic effects. It is possible that the discontinuity of such studies was due to the toxicity of the compounds or no interest of pharmaceutical companies in their synthesis. Drugs for Chagas disease are not in the interest of such industries and only the effort and persuasion of researchers in establishing partnerships could change the current scenery. An interesting case is megalol, which although previously shown by the group of Brener as an active drug even against strains of *T. cruzi* resistant to Nif and Bz, was not further investigated (Filardi & Brener 1982). However, several recent works pointed to its usefulness for the treatment of African trypanosomia-

sis (Barrett et al. 2000) and it was communicated in the 4th CostB9 Meeting in Portugal (2001) that a pharmaceutical industry will probably begin to synthesize megazol with this objective.

What must be implemented to achieve new effective and less toxic drugs for the treatment of Chagas disease? With all the knowledge accumulated about the biology and biochemistry of *T. cruzi* a lot of effort must be directed to the understanding of the mechanism of action of selected compounds. Another line that is beginning to be studied is the preparation of different formulations of drugs, which would allow its delivery to the right places. Excellent results have been obtained in the case of formulations of amphotericin for the clinical treatment of leishmaniasis (Sundar et al. 1998) and also experimental Chagas disease (Yardley & Croft 1999). Also in studies with *T. cruzi*, it was already been shown to increase in vitro and in vivo effect of different drugs. For example the use of ethylcyanoacrylate nanoparticles prepared by an emulsion polymerization process together with allopurinol or Nif and surfactants led to higher activity against epimastigotes than the corresponding free compound (Gonzalez-Martin et al. 1998, 2000). Also Molina et al. (2001) incorporated inhibitors of sterol biosynthesis into long-circulating polyethylene glycol-poly lactide nanospheres improving the bioavailability of these poorly soluble compounds and increasing the cure rate in experimental animals.

#### REFERENCES

- Alves TM, Alves R, Romanha AJ, Zani CL, Dos Santos MH, Nagem TJ 1999. Biological activities of 7-epiclusianone. *J Nat Prod* 62: 369-371.
- Andrade AL, Zicker F, de Oliveira RM, Almeida Silva S, Luquetti A, Travassos LR, Almeida IC, De Andrade SS, de Andrade JG, Martelli CM 1996. Randomized trial of efficacy of benzimidazole in treatment of early *Trypanosoma cruzi* infection. *Lancet* 348: 1407-1413.
- Andrade SG, Figueira RF 1977. Estudo experimental sobre a ação terapêutica da droga RO 7 - 1051 na infecção por diferentes cepas do *Trypanosoma cruzi*. *Rev Inst Med Trop São Paulo* 19: 335-341.
- Andrade SG, Andrade ZA, Figueira RM 1977. Estudo experimental sobre a resistência de uma cepa ao Bay 2502. *Rev Inst Med Trop São Paulo* 19: 124-129.
- Andrade SG, Costa Silva R, Santiago CM 1989. Treatment of chronic experimental *Trypanosoma cruzi* infections in mice with MK-436, a 2-substituted 5-nitroimidazole. *Bull WHO* 67: 509-514.
- Andrade SG, Figueira RM, Carvalho ML, Gorini DF 1975. Reaction of the *Trypanosoma cruzi* strain to the experimental therapeutical response to Bay 2502 (results of long term treatment). *Rev Inst Med Trop São Paulo* 17: 380-399.
- Andrade SG, Freitas LAR, Peyrol S, Pimentel AR, Sadigursky M 1991. Experimental chemotherapy of *Trypanosoma cruzi* infection persistence of parasite antigens and positive serology in parasitologically cured mice. *Bull WHO* 69: 191-199.
- Andrade SG, Pimentel AR, de Souza MM, Andrade ZA 2000. Interstitial dendritic cells of the heart harbor *Trypanosoma cruzi* antigens in experimentally infected dogs: importance for the pathogenesis of chagasic myocarditis. *Am J Trop Med Hyg* 63: 64-70.
- Andrade ZA 1999. Immunopathology of Chagas disease. *Mem Inst Oswaldo Cruz* 94 (Suppl I): 71-80.
- Anez N, Carrasco H, Parada H, Crisante G, Rojas A, Fuenmayor C, Gonzalez N, Percoco G, Borges R, Guevara P, Ramirez JL 1999. Myocardial parasite persistence in chronic chagasic patients. *Am J Trop Med Hyg* 60: 726-732.
- Apt W, Aguilera X, Arribada A, Perez C, Miranda C, Sanchez G, Zulantay I, Cortes P, Rodriguez J, Juri D 1998. Treatment of chronic Chagas disease with itraconazole and allopurinol. *Am J Trop Med Hyg* 59: 133-138.
- Araújo MS, Martins-Filho OA, Pereira ME, Brener Z 2000. A combination of benzimidazole and ketoconazole enhances efficacy of chemotherapy of experimental Chagas disease. *J Antimicrob Chemother* 45: 819-824.
- Augustyns K, Amsoms K, Yamani A, Rajan PK, Haemers A 2001. Trypanothione as a target in the design of antitrypanosomal and antileishmanial agents. *Curr Pharm Des* 7: 1117-1141.
- Ávila H, Sigman DS, Cohen LM, Millikan RC, Simpson L 1991. Polymerase chain reaction amplification of *Trypanosoma cruzi* kinetoplast minicircle DNA isolated from whole blood lysates: diagnosis of chronic Chagas disease. *Mol Biochem Parasitol* 48: 211-222.
- Bankova VS, De Castro SL, Marcucci MC 2000. Propolis: recent advances in research on chemistry and plant origin. *Apidologie* 31: 3-15.
- Bastos JK, Albuquerque S, Silva ML 1999. Evaluation of the trypanocidal activity of lignans isolated from the leaves of *Zanthoxylum naranjillo*. *Planta Med* 65: 541-544.
- Batista R, Chiari E, de Oliveira AB 1999. Trypanosomicidal kaurane diterpenes from *Wedelia paludosa*. *Planta Med* 65: 283-284.
- Berger I, Passreiter CM, Caceres A, Kubelka W 2001. Antiprotozoal activity of *Neurolaena lobata*. *Phytother Res* 15: 327-330.
- Bocca-Tourres LC 1969. La enfermedad de Chagas en período agudo y su tratamiento con el Bay 2502. *Bol Chil Parasitol* 24: 24-27.
- Bock M, Gonert R, Haberkorn A 1969. Studies with Bay 2502 on animals. *Bol Chil Parasitol* 24: 13-19.
- Bodley AL, Shapiro TA 1995. Molecular and cytotoxic effects of camptothecin, a topoisomerase I inhibitor, on trypanosomes and *Leishmania*. *Proc Natl Acad Sci USA* 92: 3726-3730.
- Boechat N, Carvalho AS, Fernandez-Ferreira E, Soares RO, Souza AS, Gibaldi D, Bozza M, Pinto AC 2001. Novel nitroimidazoles with trypanocidal and cell growth inhibition activities. *Cytobios* 105: 83-90.
- Bogitsh BJ, Middleton OL, Ribeiro-Rodrigues R 1999. Effects of the antitubulin drug trifluralin on the proliferation and metacyclogenesis of *Trypanosoma cruzi* epimastigotes. *Parasitol Res* 85: 475-480.
- Bonay P, Duran-Chica I, Fresno M, Alarcon B, Alcina A 1998. Antiparasitic effects of the intra-Golgi transport inhibitor megalomicin. *Antimicrob Agents Chemother* 42: 2668-2673.
- Bonnet B, Soulez D, Davioud-Charvet E, Landry V, Horvath D, Sergheraert C 1997. New spermine and spermidine derivatives as potent inhibitors of *Trypanosoma cruzi* trypanothione reductase. *Bioorg Med Chem* 5: 1249-1256.
- Bonse S, Richards JM, Ross SA, Lowe G, Krauth-Siegel RL 2000. (2,2':6',2"-Terpyridine)platinum(II) complexes are irreversible inhibitors of *Trypanosoma cruzi* trypanothione reductase but not of human glutathione reductase. *J Med Chem* 43: 4812-4821.
- Brener Z 1961. Atividade terapêutica do 5-nitrofuraldéido-semicarbazona (nitrofurazona) em esquemas de duração prolongada na infecção experimental pelo *Trypanosoma cruzi*. *Rev Inst Med Trop São Paulo* 3: 43-49.
- Brener Z 1968. Terapêutica experimental da doença de Chagas. In JR Cançado, *Doença de Chagas*. Belo Horizonte, Imprensa Oficial de Minas Gerais, Minas Gerais, p. 510-516.
- Brener Z 1975. Chemotherapy of *Trypanosoma cruzi* infection. *Adv Pharmacol Chemother* 13: 1-44.
- Brener Z 1979. Present status of chemotherapy and chemoprophylaxis of human trypanosomiasis in the Western hemisphere. *PharmacTherap* 7: 71-90.

- Brener Z 1984. Recent advances in chemotherapy of Chagas disease. *Mem Inst Oswaldo Cruz* 79 (Suppl): 149-155.
- Brener Z 2000. Terapêutica experimental na doença de Chagas. In Z Brener, Z Andrade, M Barral-Netto (eds), *Trypanosoma cruzi e Doença de Chagas*, 2ª ed., Guanabara Koogan, Rio de Janeiro, p. 379-388.
- Brener Z, Andrade Z, M Barral-Netto 2000. *Trypanosoma cruzi e doença de Chagas*, 2ª ed., Guanabara Koogan, Rio de Janeiro, 431 pp.
- Brener Z, Tafuri WL, Maria TA 1969. An electron microscope study on *Trypanosoma cruzi* intracellular forms in mice treated with active nitrofurazone compound. *Rev Inst Med Trop São Paulo* 11: 245-249.
- Brener Z, Cançado JR, Galvão LM, Da Luz ZM, Filardi LS, Pereira ME, Santos LM, Cançado CB 1993. An experimental and clinical assay with ketoconazole in the treatment of Chagas disease. *Mem Inst Oswaldo Cruz* 88: 149-153.
- Brengio SD, Belmonte SA, Guerreiro E, Giordano OS, Pietrobon EO, Sosa MA 2000. The sesquiterpene lactone dehydroleucodine (DhL) affects the growth of cultured epimastigotes of *Trypanosoma cruzi*. *J Parasitol* 86: 407-412.
- Bressi JC, Verlinde CL, Aronov AM, Shaw ML, Shin SS, Nguyen LN, Suresh S, Buckner FS, Van Voorhis WC, Kuntz ID, Hol WG, Gelb MH 2001. Adenosine analogues as selective inhibitors of glyceraldehyde-3-phosphate dehydrogenase of trypanosomatidae via structure-based drug design. *J Med Chem* 44: 2080-2093.
- Britto C, Cardoso MA, Vanni CMM, Haslocher-Moreno A, Xavier SS, Wincker P 1995. Polymerase chain reaction detection of *Trypanosoma cruzi* in human blood samples as a tool for diagnosis and treatment evaluation. *Parasitology* 110: 241-247.
- Britto C, Silveira C, Cardoso MA, Marques P, Luquetti A, Macedo V, Fernandes O 2001. Parasite persistence in treated chagasic patients revealed by xenodiagnosis and polymerase chain reaction. *Mem Inst Oswaldo Cruz* 96: 823-826.
- Bronia DH, Pereira BM, Lujan HD, Fretes RE, Fernandez A, Paglini PA 1999. Ganglioside treatment of acute *Trypanosoma cruzi* infection in mice promotes long-term survival and parasitological cure. *Ann Trop Med Parasitol* 93: 341-350.
- Bsiri N, Johnson C, Kayirere M, Galy AM, Galy JP, Barbe J, Osuna A, Mesa-Valle MC, Castilla Calvente JJ, Rodriguez-Cabezas MN 1996. Trypanocidal structure-activity relationship in 9-thioalkylacridines. *Ann Pharm Fr* 54: 27-33.
- Buckner FS, Griffin JH, Wilson AJ, Van Voorhis WC 2001. Potent anti-*Trypanosoma cruzi* activities of oxidosqualene cyclase inhibitors. *Antimicrob Agents Chemother* 45: 1210-1215.
- Buckner FS, Wilson AJ, White TC, Van Voorhis WC 1998. Induction of resistance to azole drugs in *Trypanosoma cruzi*. *Antimicrob Agents Chemother* 42: 3245-3250.
- Caceres A, Lopez B, Gonzalez S, Berger I, Tada I, Maki J 1998. Plants used in Guatemala for the treatment of protozoal infections. I. Screening of activity to bacteria, fungi and American trypanosomes of 13 native plants. *J Ethnopharmacol* 62: 195-202.
- Camargo ME 1966. Fluorescent antibody test for serodiagnosis of American Trypanosomiasis. Technical modification employing preserved culture forms of *Trypanosoma cruzi* in a slide test. *Rev Inst Med Trop São Paulo* 8: 227-234.
- Cançado JR 1963. Aspectos clínicos na padronização dos métodos de avaliação terapêutica na doença de Chagas. *Rev Goiana Med* 9 (Supl.): 212-232.
- Cançado JR 1968. Tratamento da doença de Chagas. In JR Cançado, *Doença de Chagas*, Imprensa Oficial de Minas Gerais, Minas Gerais, p. 517-540.
- Cançado JR 1981. Standardization of protocols for chemotherapy of Chagas disease (Working paper for discussion) Workshop on Standardization of Protocols for the Chemotherapy of Chagas Disease UNDP/World Bank/TDR. PAHO, Washington, November, 22-26.
- Cançado JR 1985. Tratamento específico. In JR Cançado, M Chuster (eds), *Cardiopatía Chagásica*. Imprensa Oficial de Minas Gerais, Minas Gerais, p. 327-355.
- Cançado JR 1997. Terapêutica específica. In JCP Dias, JR Coura, *Clínica e Terapêutica da Doença de Chagas. Uma Abordagem Prática para o Clínico Geral*. Fiocruz, Rio de Janeiro, p. 323-351.
- Cançado JR 1999. Criteria of Chagas disease cure. *Mem Inst Oswaldo Cruz* 94 (Suppl I): 331-335.
- Cançado JR 2000. Tratamento etiológico da doença de Chagas. In Z Brener, Z Andrade, M Barral-Netto (eds), *Trypanosoma cruzi e Doença de Chagas*, 2ª ed., Guanabara Koogan, Rio de Janeiro, p. 389-405.
- Cançado JR, Brener Z 1979. Terapêutica. In Z Brener, Z Andrade (eds), *Trypanosoma cruzi e Doença de Chagas*, Guanabara Koogan, Rio de Janeiro, p. 362-424.
- Cançado JR, Marra UD, Brener Z 1964. Ensaio terapêutico clínico com a 5-nitro-2-furaldeído-semicarbazona (Nitrofurazona) na forma crônica da doença de Chagas. *Rev Inst Med Trop São Paulo* 6: 12-16.
- Cançado JR, Marra UD, Lopes M, Mourão O, Faria CAF, Álvares JM, Salgado AA 1969. Toxicidade y valor terapêutico del Bay 2502 en la enfermedad de Chagas in tres esquemas posológicos. *Bol Chil Parasitol* 24: 28-32.
- Cançado JR, Duval Marra U, Mourão OG, Álvares JM, Oliveira J, Salgado A 1973. Bases para avaliação do tratamento específico da doença de Chagas humana segundo a parasitemia. *Rev Soc Bras Med Trop* 7: 155-166.
- Cançado JR, Salgado AA, Marra UD, Alvares JM, Machado JR 1975. Clinical therapeutic trial in chronic Chagas disease using nifurtimox in 3 schedules of long duration. *Rev Inst Med Trop São Paulo* 17: 111-127.
- Cançado JR, Salgado AA, Batista SM, Chiari C 1976. Segundo ensaio terapêutico com nifurtimox na doença de Chagas. *Rev Goiana Med* 22: 203-233.
- Castilla JJ, Mesa-Valle CM, Sanchez-Moreno M, Arnedo T, Rosales MJ, Mascaro C, Craciunescu D, Osuna A 1996. *In vitro* activity and biochemical effectiveness of new organometallic complexes of osmium(III) against *Leishmania donovani* and *Trypanosoma cruzi*. *Arznein Forsch* 46: 990-996.
- Cazzullo JJ, Stoka V, Turk V 2001. The major cysteine proteinase of *Trypanosoma cruzi*: a valid target for chemotherapy of Chagas disease. *Curr Pharm Des* 7: 1143-1156.
- Cerecetto H, Di Maio R, Gonzalez M, Risso M, Saenz P, Seoane G, Denicola A, Peluffo G, Quijano C, Olea-Azar C 1999. 1,2,5-Oxadiazole N-oxide derivatives and related compounds as potential antitrypanosomal drugs: structure-activity relationships. *J Med Chem* 42: 1941-1950.
- Cerecetto H, Di Maio R, Gonzalez M, Risso M, Sagrera G, Seoane G, Denicola A, Peluffo G, Quijano C, Stoppani AO, Paulino M, Olea-Azar C, Basombrio MA 2000. Synthesis and antitrypanosomal evaluation of E-isomers of 5-nitro-2-furaldehyde and 5-nitrothiophene-2-carboxaldehyde semicarbazone derivatives. Structure-activity relationships. *Eur J Med Chem* 35: 343-350.
- Cerisola JA, Rabionvich A, Alvarez M, Corleto CA, Prumeda J 1972. Enfermedad de Chagas y la transfusión de sangre. *Bol Of Sanit Panam* 73: 203-221.
- Cerisola JA, Silva NN, Prata A, Schenone H, Rohwedder R 1977. Evaluación mediante xenodiagnóstico de la efectividad del nifurtimox en la infección chagásica crónica humana. *Bol Chil Parasitol* 32: 51-62.
- Chagas C, Chagas E 1935. *Manual de Doenças Tropicais e Infectuosas*, Vol. I, Editora Freitas Bastos, Rio de Janeiro, 189 pp.
- Chan C, Yin H, Garforth J, McKie JH, Jaouhari R, Speers P, Douglas KT, Rock PJ, Yardley V, Croft SL, Fairlamb AH 1998. Phenothiazine inhibitors of trypanothione reductase as potential antitrypanosomal and antileishmanial drugs. *J*



- Med Chem* 41: 148-156.
- Chataing B, Concepcion JL, Lobaton R, Usubillaga A 1998. Inhibition of *Trypanosoma cruzi* growth *in vitro* by *Solanum alkaloids*: a comparison with ketoconazole. *Planta Med* 64: 31-36.
- Chiari E, Brener Z 1966. Contribuição ao diagnóstico parasitológico da doença de Chagas na fase crônica. *Rev Inst Med Trop São Paulo* 8: 134-138.
- Chiari E, Dias JCP, Lana M, Chiari CA 1989. Hemocultures for the parasitological diagnosis of human chronic Chagas disease. *Rev Soc Bras Med Trop* 22: 19-23.
- Chiari E, Oliveira AB, Prado MA, Alves RJ, Galvão LM, Araújo FG 1996. Potential use of WR6026 as prophylaxis against transfusion-transmitted American trypanosomiasis. *Antimicrob Agents Chemother* 40: 613-615.
- Chowdhury SF, Di Lucrezia R, Guerrero RH, Brun R, Goodman J, Ruiz-Perez LM, Pacanowska DG, Gilbert IH 2001. Novel inhibitors of leishmanial dihydrofolate reductase. *Bioorg Med Chem Lett* 11: 977-980.
- Chung MC, Gonçalves MF, Colli W, Ferreira EI, Miranda MT 1997. Synthesis and *in vitro* evaluation of potential antichagasic dipeptide prodrugs of primaquine. *J Pharm Sci* 86: 1127-1131.
- Cinque GM, Szajnman SH, Zhong L, Docampo R, Schwartzapel AJ, Rodriguez JB, Gros EG 1998. Structure-activity relationship of new growth inhibitors of *Trypanosoma cruzi*. *J Med Chem* 41: 1540-1554.
- Clark AM 1996. Natural products as a resource for new drugs. *Pharmaceutical Res* 13:1133-1141.
- Costa Silva R, Santiago CM, Pontes AL, Andrade SG 1990. Isoenzymatic pattern of *Trypanosoma cruzi* Y strain after specific chemotherapy. *Mem Inst Oswaldo Cruz* 84: 81-86.
- Coura JR 1996. Perspectivas actuales del tratamiento específico de la enfermedad de Chagas. *Bol Chil Parasitol* 51: 69-75.
- Coura JR, Silva JR 1961. Aspectos atuais do tratamento da doença de Chagas. *Rev Bras Med* 51: 283-290.
- Coura JR, Ferreira LF, Saad EA, Mortel RE, Silva JR 1961. Tentativa terapêutica com a nitrofurazona (Furacin) na forma crônica da doença de Chagas. *O Hospital* 60: 425-429.
- Coura JR, Ferreira LF, Silva JR 1962. Experiências com nitrofurazona na fase crônica da doença de Chagas. *O Hospital* 62: 957-964.
- Coura JR, Brindeiro PJ, Ferreira I 1978. Benznidazole in the treatment of Chagas disease. Current Chemotherapy. *Proc 10 th Int Cong Chemotherapy* 1: 161-162.
- Coura JR, de Abreu LL, Willcox HP, Petana W 1991. Evaluation of the xenodiagnosis of chronic Chagas patients infected ten years or over in an area where transmission has been interrupted-Iguatama and Pains, West Minas Gerais State, Brazil. *Mem Inst Oswaldo Cruz* 86: 395-398.
- Coura JR, de Abreu LL, Willcox HP, Petana W 1997. Comparative controlled study on the use of benznidazole, nifurtimox and placebo, in the chronic form of Chagas disease, in a field area with interrupted transmission. I. Preliminary evaluation. *Rev Soc Bras Med Trop* 30: 139-144.
- Croft SL 1994. A rationale for antiparasite drug discovery. *Parasitol Today* 10: 385-386.
- Croft SL, Snowdon D, Yardley V 1996. The activities of four anticancer alkyllysophospholipids against *Leishmania donovani*, *Trypanosoma cruzi* and *Trypanosoma brucei*. *J Antimicrob Chemother* 38: 1041-1047.
- Dantas AP 2000. *Efeito de agentes anti-microtúbulos e de derivados de naftoquinonas sobre Trypanosoma cruzi*, MSc Thesis, Instituto Oswaldo Cruz-Fiocruz, Rio de Janeiro, xiv + 204 pp.
- Dantas AP, Ortigão M, Traub-Czebo Y, Polaquevitch PFC, Barbosa HS, De Castro SL 1998. Studies on the effect of dinitroaniline herbicides in *Trypanosoma cruzi*. *Mem Inst Oswaldo Cruz* 93 (suppl II): 311
- De Castro SL 1993. The challenge of Chagas disease chemotherapy: an update of drugs assayed against *Trypanosoma cruzi*. *Acta Trop* 53: 83-98.
- De Castro 2001. Propolis: Biological and pharmacological activities. Therapeutic uses of this bee-product. *Ann Rev Biol Sci*, in press.
- De Castro SL, Higashi KO 1995. Effect of different formulations of propolis on mice infected with *Trypanosoma cruzi*. *J Ethnopharmacol* 46: 55-59.
- De Castro SL, Pinto MCFR, Pinto AV 1994. Screening of natural and synthetic drugs against *Trypanosoma cruzi*: establishing a structure/activity relationship. *Microbios* 78: 83-90.
- De Castro SL, Dantas AP, Salomão K, Prytyk E, Pereira AS, Aquino Neto FR, Bankova VS 2001. Trypanocidal activity and chemical composition of Bulgarian samples of propolis. *Proc. 4th COST-B9 Cong Antiprotozoal Chemother*, Lisbon, Portugal.
- De Conti R, Rita RM, de Souza EM, Melo PS, Haun M, De Castro SL, Durán N 1996. *In vitro* trypanocidal activities of a novel series of N, N-dimethyl-2-propen-1-amine derivative. *Microbios* 85: 83-87.
- De Moura KCG, Emery FS, Neves-Pinto C, Pinto MCFR, Dantas AP, Salomão K, De Castro SL, Pinto AV 2001. Synthesis and trypanocidal activity of naphthoquinones isolated from *Tabebuia* and heterocyclic derivatives: a review from an interdisciplinary study. *J Braz Chem Soc* 12: 325-338.
- Deharo E, Loyevsky M, John C, Balanza E, Ruiz G, Munoz V, Gordeuk VR 2000. Aminothiol multidentate chelators against Chagas disease. *Exp Parasitol* 94: 198-200.
- Di Maio R, Cerecetto H, Seoane G, Ochoa C, Aran VJ, Perez E, Gomez Barrio A, Muelas S 1999. Synthesis and antichagasic properties of new 1,2,6-thiadiazin-3, 5-dione 1,1-dioxides and related compounds. *Arznein Forsch* 49: 759-763.
- Dias JCP 2000. Epidemiologia. In Z Brener, Z Andrade, M Barral-Netto (eds), *Trypanosoma cruzi e Doença de Chagas*, 2ª ed., Guanabara Koogan, Rio de Janeiro, p. 48-74.
- Diaz de Toranzo EG, Castro JA, Franke de Cazzulo BM, Cazzulo JJ 1988. Interaction of benznidazole reactive metabolites with nuclear and kinetoplastic DNA, proteins and lipids from *Trypanosoma cruzi*. *Experientia* 44: 880-881.
- DoCampo R 2001. Recent developments in the chemotherapy of Chagas disease. *Curr Pharm Design* 7: 1157-1164.
- DoCampo R, Moreno SNJ 1986. Free radical metabolism of antiparasitic agents. *Fed Proceed* 45: 2471-2476.
- DoCampo R, Moreno SNJ 2001. Bisphosphonates as chemotherapeutic agents against trypanosomatids and Apicomplexan parasites. *Current Drug Targets-Infectious Disorders* 1: 51-61.
- Dubin M, Fernandez Villamil SH, Stoppani AO 2001. Cytotoxicity of beta-lapachone, a naphthoquinone with possible therapeutic use. *Medicina (B Aires)* 61: 343-350.
- Eakin AE, Guerra A, Focia PJ, Torres-Martinez J, Craig SP 1997. Hypoxanthine phosphoribosyltransferase from *Trypanosoma cruzi* as a target for structure-based inhibitor design: crystallization and inhibition studies with purine analogs. *Antimicrob Agents Chemother* 41: 1686-1692.
- Engel JC, Doyle PS, Palmer J, Bainton DF, McKerrow JH 1998a. Cysteine protease inhibitors after Golgi complex ultrastructure and function in *Trypanosoma cruzi*. *J Cell Sci* 111: 597-606.
- Engel JC, Doyle PS, Hsieh I, McKerrow JH 1998b. Cysteine protease inhibitors cure an experimental *Trypanosoma cruzi* infection. *J Exp Med* 188: 725-734.
- Fairlamb AH 1999. Future prospects for the chemotherapy of Chagas disease. *Medicina (B Aires)* 59: 179-187.
- Fairlamb AH, Cerami A 1992. Metabolism and functions of trypanothione in the kinetoplastida. *Annu Rev Microbiol* 46: 695-729.
- Fernandez-Gomez R, Zerrouk H, Sebti F, Loyens M, Benslimane A, Ouassi MA 1994. Growth inhibition of *Trypanosoma cruzi* and *Leishmania donovani infantum* by different snake venoms: preliminary identification of proteins from *Cerastes*

- cerastes* venom which interact with the parasites. *Toxicon* 32: 875-882.
- Ferreira HO 1961. Forma aguda da doença de Chagas tratada pela nitrofurazona. *Rev Inst Med Trop São Paulo* 3: 287-289.
- Ferreira HO 1962. Fase aguda da doença de Chagas. *O Hospital* 61: 307-311.
- Ferreira HO 1976. Ensaio terapêutico-clínico com benznidazol na doença de Chagas. *Rev Inst Med Trop São Paulo* 18: 357-364.
- Ferreira HO 1990. Tratamento da forma indeterminada da doença de Chagas com nifurtimox e benznidazol. *Rev Soc Bras Med Trop* 23: 209-211.
- Ferreira HO, Prata A, Rassi A 1963. Administração prolongada de nitrofurazona no tratamento da doença de Chagas aguda. *O Hospital* 63: 131-139.
- Ferreira MS, Nishioka SA, Rocha A, Silva AM 1997a. Doença de Chagas e imunossupressão. In JCP Dias, JR Coura (eds), *Clínica e Terapêutica da Doença de Chagas*. Fiocruz, Rio de Janeiro, p. 365-379.
- Ferreira MS, Nishioka SD, Silvestre MT, Borges AS, Nunes-Araujo FR, Rocha A 1997b. Reactivation of Chagas disease in patients with AIDS: report of three new cases and review of the literature. *Clin Infect Dis* 25: 1397-1400.
- Fichera L, Esteva M, Wimmer Z, Rodriguez JB, Gros EG 1995. Effects of juvenile hormone analogues (JHA) on the development of *Trypanosoma cruzi*. *Z Naturforsch* 50C: 578-580.
- Fife Jr. EH, Mushel LH 1959. Fluorescent antibody technique for serodiagnosis of *Trypanosoma cruzi* infection. *Proc Soc Exp Biol* 101: 540-543.
- Filardi LS, Brener Z 1982. Nitroimidazole-thiadiazole derivative with curative action in experimental *Trypanosoma cruzi* infection. *Ann Trop Med Parasitol* 76: 293-297.
- Fournet A, Barrios AA, Munoz V 1994. Leishmanicidal and trypanocidal activities of Bolivian medicinal plants. *J Ethnopharmacol* 41: 19-37.
- Fournet A, Inchausti A, Yaluff G, Rojas De Arias A, Guinaudeau H, Bruneton J, Breidenbach MA, Karplus PA, Faerman CH 1998. Trypanocidal bisbenzylisoquinoline alkaloids are inhibitors of trypanothione reductase. *J Enzym Inhib* 13: 1-9.
- Fournet A, Rojas De Arias A, Ferreira ME, Nakayama H, Torres de Ortiz S, Schinini A, Samudio M, Vera de Bilbao N, Lavault M, Bonte F 2000. Efficacy of the bisbenzylisoquinoline alkaloids in acute and chronic *Trypanosoma cruzi* murine model. *Int J Antimicrob Agents* 13: 189-195.
- Fragata Filho AA, Boianain E, Silva MAD, Correia EB, Borges Filho R, Martins C, Salene V, Batlouni M, Souza E 1995. Validade do tratamento etiológico da fase crônica da doença de Chagas com benznidazol. *Arq Bras Cardiol* 65 (Supl I): 71.
- Fragoso SP, Goldenberg S 1992. Cloning and characterization of the gene encoding *Trypanosoma cruzi* DNA topoisomerase II. *Mol Biochem Parasitol* 55: 127-134.
- Fragoso SP, Mattei D, Hines JC, Ray D, Goldenberg S 1998. Expression and cellular localization of *Trypanosoma cruzi* type II DNA topoisomerase. *Mol Biochem Parasitol* 94: 197-204.
- Freitas JLP, Almeida JO 1949. Nova técnica de fixação de complemento para a moléstia de Chagas. (Reação quantitativa com antígeno geleificado de culturas de *Trypanosoma cruzi*). *O Hospital* 35: 787-800.
- Freymann DM, Wenck MA, Engel JC, Feng J, Focia PJ, Eakin AE, Craig SP 2000. Efficient identification of inhibitors targeting the closed active site conformation of the HPRT from *Trypanosoma cruzi*. *Chem Biol* 7: 957-968.
- Galhardo MCG, Martins, I, Hasslocher-Moreno A, Xavier SS, Coelho JMC, Vasconcellos ACV, Santos RR 1999. Reactivação da infecção por *Trypanosoma cruzi* em paciente com síndrome de imunodeficiência adquirida. *Rev Soc Bras Med Trop* 32: 291-294.
- Galleano RH, Marr JJ, Sosa RR 1990. Therapeutic efficacy of allopurinol in patients with chronic Chagas disease. *Am J Trop Med Hyg* 43: 159-166.
- Girault S, Davioud-Charvet E, Salmon L, Berecibar A, Debreu MA, Sergheraert C 1998. Structure-activity relationships in 2-aminodiphenylsulfides against trypanothione reductase from *Trypanosoma cruzi*. *Bioorg Med Chem Lett* 8: 1175-1180.
- Girault S, Grellier P, Berecibar A, Maes L, Mouray E, Lemiere P, Debreu MA, Davioud-Charvet E, Sergheraert C 2000. Antimalarial, antitrypanosomal, and antileishmanial activities and cytotoxicity of bis(9-amino-6-chloro-2-methoxyacridines): influence of the linker. *J Med Chem* 43: 2646-2654.
- Gonzales-Perdomo M, De Castro SL, Meirelles MN, Goldenberg S 1990. *Trypanosoma cruzi* proliferation and differentiation are blocked by topoisomerase II inhibitors. *Antimicrob Agents Chemother* 34: 1707-1714.
- Gonzalez-Martin G, Figueroa C, Merino I, Osuna A 2000. Allopurinol encapsulated in polycyanoacrylate nanoparticles as potential lysosomotropic carrier: preparation and trypanocidal activity. *Eur J Pharm Biopharm* 49: 137-142.
- Gonzalez-Martin G, Merino I, Rodriguez-Cabezas MN, Torres M, Nunez R, Osuna A, 1998. Characterization and trypanocidal activity of nifurtimox-containing and empty nanoparticles of polyethylcyanoacrylates. *J Pharm Pharmacol* 50: 29-35.
- Grellier P, Sinou V, Garreau-de Loubresse N, Bylen E, Boulard Y, Schrevel J 1999. Selective and reversible effects of *Vinca* alkaloids on *Trypanosoma cruzi* epimastigote forms: blockage of cytokinesis without inhibition of the organelle duplication. *Cell Motil Cytoskeleton* 42: 36-47.
- Guerreiro C, Machado A 1913. Da reação de Bordet e Gengou na moléstia de Carlos Chagas como elemento de diagnóstico. *Brasil Med* 27: 225-226.
- Gutierrez-Correa J, Fairlamb AH, Stoppani AO 2001. *Trypanosoma cruzi* trypanothione reductase is inactivated by peroxidase-generated phenothiazine cationic radicals. *Free Radic Res* 34: 363-378.
- Henderson GB, Ulrich P, Fairlamb AH, Rosemberg I, Pereira M, Sela M, Cerami A 1988. "Subversive" substrates for the enzyme trypanothione disulphide reductase, alternative approach to chemotherapy of Chagas disease. *Proc Natl Acad Sci USA* 85: 5374-5378.
- Higashi KO, De Castro SL 1994. Propolis extracts are effective against *Trypanosoma cruzi* and have an impact on its interaction with host cells. *J Ethnopharmacol* 43: 149-155.
- Higuchi ML 1999. Human chagasic cardiopathy: participation of parasite antigens, subsets lymphocytes, cytokines and microvascular abnormalities. *Mem Inst Oswaldo Cruz* 94 (Suppl I): 263-267.
- Higuchi ML, Brito T, Reis M, Bellotti G, Pereira-Barreto AC, Pileggi F 1993. Correlation between *Trypanosoma cruzi* parasitism and myocardial inflammation in human chronic chagasic myocarditis. Light microscopy and immunohistochemical findings. *Cardiovasc Pathol* 2: 101-106.
- Higuchi ML, Reis M, Aiello VD, Benvenuti LA, Gutierrez PS, Bellotti G, Pileggi F 1997. Human chronic chagasic myocarditis is *Trypanosoma cruzi* antigen and CD8<sup>+</sup> T cell dependent. *Am J Trop Med Hyg* 56: 485-489.
- Hoare CA, Wallace FG 1966. Developmental stages of trypanosomatid flagellates: a new terminology. *Nature* 204: 69-70.
- Hoffmann BK 1972. Toxicological investigations on the tolerability of nifurtimox. *Arzneim Forsch* 22: 1590-1603.
- Ianni BM, Arteaga E, Mady C, Barretto ACP, Pileggi F 1993. Uso de benznidazol em chagásicos na forma indeterminada: Resultados a longo prazo. *Arq Bras Cardiol* 61 (Supl II): 130.
- Jatene AD, Costa R, Jatene MB 1997. Tratamento cirúrgico da cardiopatia chagásica. In JCP Dias, JR Coura (eds), *Clínica e Terapêutica da Doença de Chagas*. Fiocruz, Rio de Janeiro,

- p. 255-265.
- Jones EM, Colley DG, Tostes S, Lopes ER, Venencak-Jones C, Mc Curley TL 1993. Amplification of *Trypanosoma cruzi* DNA sequence from inflammatory lesions in human chagasic cardiomyopathy. *Am J Trop Med Hyg* 48: 348-357.
- Junqueira ACV, Chiari E, Wincker P 1996. Comparison of the polymerase chain reaction with two classical parasitological methods, for diagnosis of Chagas disease in endemic region of North-Eastern Brazil. *Trans R Soc Trop Med Hyg* 90: 129-132.
- Kelser RA 1936. A complement fixation test for Chagas disease employing an artificial culture antigen. *Am J Trop Med Hyg* 16: 405-415.
- Kerschmann RL, Wolfson JS, McHugh GL, Dickersin GR, Hooper DC, Swartz MN 1989. Novobiocin-induced ultrastructural changes and antagonism of DNA synthesis in *Trypanosoma cruzi* amastigotes growing in cell-free medium. *J Protozool* 36: 14-20.
- Kinnamon KE, Poon BT, Hanson WL, Waits VB 1996. Primaquine analogues that are potent anti-*Trypanosoma cruzi* agents in a mouse model. *Ann Trop Med Parasitol* 90: 467-474.
- Kinnamon KE, Poon BT, Hanson WL, Waits VB 1997. Evidence that certain 8-aminoquinolines are potentially effective drugs against Chagas disease. *Ann Trop Med Parasitol* 91: 147-52.
- Krauth-Siegel RL, Enders B, Henderson GB, Fairlamb AH, Schirmer RH 1987. Trypanothione reductase from *Trypanosoma cruzi*. Purification and characterization of the crystalline enzyme. *Eur J Biochem* 164: 123-128.
- Krettli AU, Brener Z 1982. Resistance against *Trypanosoma cruzi* associate to anti-living trypomastigote antibodies. *J Immunol* 128: 2009-2012.
- Krettli AU, Cançado JR, Brener Z 1984. Criterion of cure of human Chagas disease after specific treatment: recent advances. *Mem Inst Oswaldo Cruz* 79 (Suppl): 157-164.
- Lalmanach G, Mayer R, Serveau C, Scharfstein J, Gauthier F 1996. Biotin-labelled peptidyl diazomethane inhibitors derived from the substrate-like sequence of cystatin: targeting of the active site of cruzipain, the major cysteine proteinase of *Trypanosoma cruzi*. *Biochem J* 318: 395-399.
- Lane JE, Ribeiro-Rodrigues R, Suarez CC, Bogitsh BJ, Jones MM, Singh PK, Carter CE 1996. *In vitro* trypanocidal activity of tetraethylthiuram disulfide and sodium diethylamine-N-carbodithioate on *Trypanosoma cruzi*. *Am J Trop Med Hyg* 55: 263-266.
- Lane JE, Bogitsh BJ, Ribeiro-Rodrigues R, Kral MV, Jones MM, Carter CE 1998. Ultrastructural effects of the chelating agent 1,10-phenanthroline on *Trypanosoma cruzi* epimastigotes *in vitro*. *Parasitol Res* 84: 399-402.
- Lauria-Pires L, Castro CN, Emanuel A, Prata A 1988. Ineficácia do allopurinol em pacientes na fase aguda da doença de Chagas. *Rev Soc Med Trop* 21: 79
- Lazzari JO, Freilij H 1998. Tratamiento de la enfermedad de Chagas crónica en Argentina. *Rev Patol Trop* 27 (Supl): 11-16.
- Li Z, Fennie MW, Ganem B, Hancock MT, Kobaslija M, Rattendi D, Bacchi CJ, O'Sullivan MC 2001. Polyamines with N-(3-phenylpropyl) substituents are effective competitive inhibitors of trypanothione reductase and trypanocidal agents. *Bioorg Med Chem Lett* 11: 251-254.
- Lira R, Contreras LM, Rita RM, Urbina JA 2001. Mechanism of action of anti-proliferative lysophospholipid analogues against the protozoan parasite *Trypanosoma cruzi*: potentiation of *in vitro* activity by the sterol biosynthesis inhibitor ketoconazole. *J Antimicrob Chemother* 47: 537-546.
- Lopes JN, Cruz FS, DoCampo R, Vasconcellos ME, Sampaio MCR, Pinto AV, Gilbert B 1978. *In vitro* and *in vivo* evaluation of the toxicity of 1,4-naphthoquinone and 1,2-naphthoquinone derivatives against *Trypanosoma cruzi*. *Ann Trop Med Parasitol* 72: 523-531.
- Lujan HD, Paglini P, Fretes R, Fernandez A, Fidelio GD, Bronia DH 1993. Effect of gangliosides on *Trypanosoma cruzi* infection in mice. *Life Sci* 53: PL69-73.
- Luque F, Fernandez-Ramos C, Entrala E, Rosales MJ, Navarro JA, Romero MA, Salas JM, Sanchez-Moreno M 2000. *In vitro* evaluation of newly synthesized [1,2,4]triazolo[1,5a]pyrimidine derivatives against *Trypanosoma cruzi*, *Leishmania donovani* and *Phytomonas staheli*. *Comp Biochem Physiol C Toxicol Pharmacol* 126: 39-44.
- Luquetti 1997. Etiological treatment for Chagas disease. The National Health Foundation of Brazil. *Parasitol Today* 13: 127-128.
- Luz ZMP, Coutinho MG, Cançado JR, Krettli AU 1994. Hemocultura: técnica sensível na detecção do *Trypanosoma cruzi* em pacientes chagásicos crônicos. *Rev Soc Bras Med Trop* 27: 143-148.
- Macêdo VO 1997. Forma indeterminada da doença de Chagas. In JCP Dias, JR Coura (eds), *Clínica e Terapêutica da Doença de Chagas. Uma Abordagem Prática para o Clínico Geral*. Fiocruz, Rio de Janeiro, p. 383-409.
- Macêdo VO, Silveira CA 1987. Perspectivas da terapêutica específica na doença de Chagas. Experiências na forma indeterminada. *Rev Soc Bras Med Trop* 20 (Supl II): M24-M26.
- Mafezoli J, Vieira PC, Fernandes JB, Da Silva MF, De Albuquerque S 2000. *In vitro* activity of Rutaceae species against the trypomastigote form of *Trypanosoma cruzi*. *J Ethnopharmacol* 73: 335-340.
- Mahiou V, Roblot F, Fournet A, Hocquemiller R 2000. Bisbenzylisoquinoline alkaloids from *Guatteria boliviana* (Annonaceae). *Phytochemistry* 54: 709-716.
- Marcucci MC, Ferreres F, García-Viguera C, Bankova VS, De Castro SL, Dantas AP, Valente PHM, Paulino N 2001. Phenolic compounds from Brazilian propolis with pharmacological activities. *J Ethnopharmacol* 74: 105-112.
- Marr JJ 1991. Purine analogs as chemotherapeutic agents in leishmaniasis and American trypanosomiasis. *J Lab Clin Med* 118: 111-119.
- Martin MB, Grimley JS, Lewis JC, Heath HT, 3rd, Bailey BN, Kendrick H, Yardley V, Caldera A, Lira R, Urbina JA, Moreno SN, Docampo R, Croft SL, Oldfield E 2001. Bisphosphonates inhibit the growth of *Trypanosoma brucei*, *Trypanosoma cruzi*, *Leishmania donovani*, *Toxoplasma gondii*, and *Plasmodium falciparum*: A potential route to chemotherapy. *J Med Chem* 44: 909-916.
- Maya JD, Morello A, Repetto Y, Tellez R, Rodriguez A, Zelada U, Puebla P, Caballero E, Medarde M, Nunez-Vergara LJ, Squella JA, Bonta M, Bollo S, San Feliciano A 2000. Effects of 3-chloro-phenyl-1,4-dihydropyridine derivatives on *Trypanosoma cruzi* epimastigotes. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 125: 103-109.
- Mayer M, Rocha Lima H 1912. Zur Entwicklung von *Schizotrypanum cruzi* in Saengatieren. *Arch Schiffs u Tropen Hyg* 16: 90-94.
- Mayer M, Rocha Lima H 1914. Zum Verhalten von *Schizotrypanum cruzi* in Warmblutern und Arthropoden. *Arch Schiffs u Tropen-Hyg* 5: 101-136.
- Mazza S, Cossio R, Zucardi E 1937. Primer caso agudo de enfermedad de Chagas, comprobado em Tucuman y su tratamiento com Bayer 7602. *Mis Estudios Patolog Reg Argentina Publ* 70 (Univ Buenos Aires) (MEPRA) 32: 3-18.
- Mazza S, Basso G, Basso R 1942. Ensayos terapêuticos del producto 9736 (As) Bayer y de su acción comparada en el 7602 (Ac) Bayer en la enfermedad de Chagas. *Mis Estudios Patolog Reg Argentina Publ* 70 (Univ Buenos Aires) (MEPRA) 61: 1-76.
- McKerrow JH 1999. Development of cysteine protease inhibitors as chemotherapy for parasitic diseases: Insights on safety, target validation, and mechanism of action. *Int J Parasitol* 29: 833-837.
- Medrano NM, Luz MRMP, Cabello P, Tapia GT, Van Leuven F, Araújo-Jorge TC 1996. Acute Chagas disease: Plasma levels of alpha-2-macroglobulin and C-reactive protein in children

- under 13 years in a high endemic area of Bolivia. *J Trop Pediatr* 42: 68-74.
- Molina J, Martins-Filho O, Brener Z, Romanha AJ, Loeberberg D, Urbina JA 2000. Activities of the triazole derivative SCH 56592 (posaconazole) against drug-resistant strains of the protozoan parasite *Trypanosoma (Schizotrypanum) cruzi* in immunocompetent and immunosuppressed murine hosts. *Antimicrob Agents Chemother* 44: 150-155.
- Molina J, Urbina J, Gref R, Brener Z, Rodrigues Junior JM 2001. Cure of experimental Chagas disease by the bis-triazole DO870 incorporated into "stealth" polyethyleneglycol-poly lactide nanospheres. *J Antimicrob Chemother* 47: 101-104.
- Montalvetti A, Bailey BN, Martin MB, Severin GW, Oldfield E, DoCampo R 2001. Bisphosphonates are potent inhibitors of *Trypanosoma cruzi* farnesyl pyrophosphate synthase. *J Biol Chem* 276: 33930-33937.
- Morello A, Lipchenca I, Cassels BK, Speisky H, Aldunate J, Repetto Y 1994. Trypanocidal effect of boldine and related alkaloids upon several strains of *Trypanosoma cruzi*. *Comp Biochem Physiol Pharmacol Toxicol Endocrinol* 107: 367-371.
- Morello A, Pavani M, Garbarino JA, Chamy MC, Frey C, Mancilla J, Guerrero A, Repetto Y, Ferreira J 1995. Effects and mode of action of 1,4-naphthoquinones isolated from *Calceolaria sessilis* on tumoral cells and *Trypanosoma* parasites. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 112: 119-128.
- Moser DR, Kirchoff LV, Donelson JE 1989. Detection of *Trypanosoma cruzi* by DNA amplification using the polymerase chain reaction. *J Clin Microbiol* 27: 1477-1482.
- Muelas S, Di Maio R, Cerecetto H, Seoane G, Ochoa C, Escario JA, Gomez-Barrio A 2001. New thiadiazine derivatives with activity against *Trypanosoma cruzi* amastigotes. *Folia Parasitol (Praha)* 48: 105-108.
- Muelas-Serrano S, Nogal JJ, Martinez-Diaz RA, Escario JA, Martinez-Fernandez AR, Gomez-Barrio A 2000. *In vitro* screening of American plant extracts on *Trypanosoma cruzi* and *Trichomonas vaginalis*. *J Ethnopharmacol* 71: 101-107.
- Nakajima-Shimada J, Hirota Y, Aoki T 1996. Inhibition of *Trypanosoma cruzi* growth in mammalian cells by purine and pyrimidine analogs. *Antimicrob Agents Chemother*. 40: 2455-2458.
- Neal RA, Miles RA 1970. Indirect hemagglutination test for Chagas disease, with a simple method for survey work. *Rev Inst Med Trop São Paulo* 12: 325-332.
- Neira I, Poblete L, Porcille P, Silva P, Araya J, Borquez J, Morales G, Loyola LA, Sagua H 1998. Activity of Dipernoids isolated from *Azorella compacta* (Llaretia) on *Trypanosoma cruzi* amastigotes. *Bol Chil Parasitol* 53: 9-13.
- Neves-Pinto C, Dantas AP, De Moura KCG, Emery FS, Polequevitch PF, Pinto MCFR, De Castro SL, Pinto AV 2000. Chemical reactivity studies with naphthoquinones from *Tabebuia* with anti-trypanosomal efficacy. *Arzneim Forsch* 50: 1120-1128.
- Nunez-Vergara LJ, Squella JA, Bollo-Dragnic S, Morello A, Repetto Y, Aldunate J, Letelier ME 1997. Nitro aryl 1,4-dihydropyridine derivatives: effects on *Trypanosoma cruzi*. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 118: 105-111.
- Nussenzweig V, Sonntag R, Biancalana A, Freitas JLP, Amato Neto V, Kloetzel J 1953. Ação da violeta de genciana sobre o *T. cruzi in vitro*: sua importância na esterilização do sangue destinado à transfusão. *Rev Paul Med* 42: 57-58.
- Ochoa C, Perez E, Perez R, Suarez M, Ochoa E, Rodriguez H, Gomez Barrio A, Muelas S, Nogal JJ, Martinez RA 1999. Synthesis and antiprotozoan properties of new 3,5-disubstituted-tetrahydro-2H-1,3,5-thiadiazine-2-thione derivatives. *Arzneim Forsch* 49: 764-769.
- Oliveira DA, Fernandes AM, De Conti R, Rodriguez JA, Haun M, Souza-Brito AR, De Castro SL, Durán N 1999. Evaluation of *in vitro* toxicity of N,N-dimethyl-2-propen-1-amine isomers. *Pharmazie* 54: 847-850.
- Olmo E, Armas MG, Lopez-Perez JL, Ruiz G, Vargas F, Gimenez A, Deharo E, Feliciano AS 2001. Anti-*Trypanosoma* activity of some natural stilbenoids and synthetic related heterocyclic compounds. *Bioorg Med Chem Lett* 11: 755-757.
- OPAS/OMS 1998. Tratamiento Etiológico de la Enfermedad de Chagas. Conclusiones de una consulta técnica. OPC/HCP/HCT/140/99, 32 pp. (published in *Rev Patol Trop* 28: 247-279, 1999)
- Packchanian A 1952. Chemotherapy of experimental Chagas disease with nitrofurans compounds. *J Parasitol* 38: 30-40.
- Packchanian A 1957. Chemotherapy of experimental Chagas disease with nitrofurans compounds. *Antibiotics & Chemotherapy* 7: 13-23.
- Paglini-Oliva P, Fernandez AR, Fretes R, Peslman A 1998. Structural, ultrastructural studies and evolution of *Trypanosoma cruzi*-infected mice treated with thioridazine. *Exp Mol Pathol* 65: 78-86.
- Pereira DG, De Castro SL, Durán N 1998. Activity of N,N-dimethyl-1-2-propen-1-amine derivatives in mice experimentally infected with *Trypanosoma cruzi*. *Acta Trop* 69: 205-211.
- Pifano FC 1941. La enfermedad de Chagas en el Estado Jaracuy, Venezuela. *Caracas Medico* 8: 1103-1166.
- Pineda JP, Luquetti A, Castro C 1998. Comparison between classical and artificial xenodiagnosis in chronic Chagas disease. *Rev Soc Bras Med Trop* 31: 473-480.
- Pinto AV, Neves Pinto C, Pinto MCFR, Santa-Rita RM, Pezzella C, De Castro SL 1997. Trypanocidal activity of synthetic heterocyclic derivatives from active quinones from *Tabebuia* sp. *Arzneim-Forsch* 4: 74-79.
- Polak A, Richle R 1978. Mode of action of 2-nitroimidazole derivative benznidazole. *Ann Trop Med Parasitol* 72: 228-232.
- Prata A 1963. Estado atual da terapêutica da doença de Chagas. Revisão dos medicamentos até hoje utilizados. *Rev Goiana Medicina* 9 (Supl.): 109-124.
- Prata A, Macedo V, Santos I, Cerisola JA, Silva N 1975. Tratamento da doença de Chagas pelo nifurtimox (Bayer 2505). *Rev Soc Brasil Med Trop* 6: 297-308.
- Rassi A, Ferreira HO 1971. Tentativas de tratamento específico da fase aguda da doença de Chagas com nitrofuranos em esquemas de duração prolongada. *Rev Soc Bras Med Trop* 5: 235-262.
- Rassi A, Luquetti AO 1992. Therapy of Chagas disease. In S Wendel, Z Brener, E Camargo, A Rassi (eds), *Chagas Disease (American Trypanosomiasis): its Impact on Transfusion and Clinical Medicine*. ISBT, São Paulo, p. 237-247.
- Rassi A, Amato Neto V, De Siqueira AF, Ferrioli Filho F, Amato VS, Rassi Jr A 1999. Protective effect of benznidazole against parasite reactivation in patients chronically infected with *Trypanosoma cruzi* and treated with corticoids for associated diseases. *Rev Soc Bras Med Trop* 32: 475-482.
- Reche P, Arrebola R, Santi DV, Gonzalez-Pacanowska D, Ruiz-Perez LM 1996. Expression and characterization of the *Trypanosoma cruzi* dihydrofolate reductase domain. *Mol Biochem Parasitol* 76: 175-185.
- Rezende JM 1959. Forma digestiva da moléstia de Chagas. *Rev Goiana Med* 5: 193-220.
- Rezende JM, Lauar KM, Oliveira AR 1960. Aspectos clínicos e radiológicos da aperistalsis do esôfago. *Rev Bras Gastroenterol* 12: 247-261.
- Richle R 1973. Chemotherapy of experimental acute Chagas disease in mice: beneficial effect of Ro-71051 on parasitemia and tissue parasitism. *Le Progres Medical* 101: 282.
- Rivarola HW, Fernandez AR, Enders JE, Fretes R, Gea S, Suligoy M, Palma JA, Paglini-Oliva P 1999. Thioridazine treatment modifies the evolution of *Trypanosoma cruzi* infection in mice. *Ann Trop Med Parasitol* 93: 695-702.
- Rivas P, Cassels BK, Morello A, Repetto Y 1999. Effects of

- some  $\beta$ -carboline alkaloids on intact *Trypanosoma cruzi* epimastigotes. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 122: 27-31.
- Rodrigues RR, Lane JE, Carter CE, Bogitsh BJ, Singh PK, Zimmerman LJ, Molenda JJ, Jones MM 1995. Chelating agent inhibition of *Trypanosoma cruzi* epimastigotes *in vitro*. *J Inorg Biochem* 60: 277-288.
- Rodriguez JB 2001. Specific molecular targets to control tropical diseases. *Curr Pharm Des* 7: 1105-1116.
- Rodriguez JB, Gros EG 1995. Recent developments in the control of *Trypanosoma cruzi*, the causative agents of Chagas disease. *Curr Med Chem* 2: 723-742.
- Rodriguez JB, DoCampo R, Gros EG 2000. Sulphur-containing derivatives structurally related to fenoxycarb are potent growth inhibitors against the intracellular form of *Trypanosoma cruzi*. *Int J Antimicrob Agents* 13: 215-218.
- Roush WR, Hernandez AA, McKerrow JH, Selzer PM, Hansell E, Engel JC 2000. Design, synthesis and evaluation of D-homophenylalanylepoxysuccinate inhibitors of the trypanosomal cysteine protease cruzain. *Tetrahedron* 56: 9747-9762.
- Rovai LE, Aoki A, Gerez de Burgos NM, Blanco A 1990. Effect of gossypol on trypomastigotes and amastigotes of *Trypanosoma cruzi*. *J Protozool* 37: 280-286.
- Rubio M, Donoso F 1969. Enfermedad de Chagas en niños y tratamiento con Bay 2502. *Bol Chil Parasitol* 24: 43-48.
- Ryley JF, McGregor S, Wilson RG 1988. Activity of ICI 195,739 - a novel orally active bistriazole - in rodent models of fungal and protozoal infection. *Ann N Y Acad Sci* 310: 328.
- Salmon-Chemin L, Buisine E, Yardley V, Kohler S, Debreu MA, Landry V, Sergheraert C, Croft SL, Krauth-Siegel RL, Davioud-Charvet E 2001. 2- and 3-substituted 1,4-naphthoquinone derivatives as subversive substrates of trypanothione reductase and lipoamide dehydrogenase from *Trypanosoma cruzi*: synthesis and correlation between redox cycling activities and *in vitro* cytotoxicity. *J Med Chem* 44: 548-565.
- Santa-Rita RM, Barbosa HS, Meirelles MNL, De Castro SL 2000. Effect of alkyllysophospholipids on the proliferation and differentiation of *Trypanosoma cruzi*. *Acta Trop* 75: 219-228.
- Santos AF, Ferraz PA, de Abreu FC, Chiari E, Goulart MO, Sant'Ana AE 2001. Molluscicidal and trypanocidal activities of lapachol derivatives. *Planta Med* 67: 92-3.
- Schenone H, Concha L, Aranda R, Rojas A, Alfaro E 1969. Experiencia terapéutica con el Bayer 2502 en la infección chagásica crónica del adulto. Importancia del uso adecuado del xenodiagnóstico. *Bol Chil Parasitol* 24: 66-69.
- Schenone H, Concha L, Aranda R, Rojas A, Alfaro E, Knierin E, Rojo M 1975. Atividade quimioterápica de um derivado nitroimidazolacetamida na infecção chagásica crônica. *Bol Chil Parasitol* 30: 91-93.
- Schenone H, Concha L, Aranda R, Rojas A, Knierin F, Rojo M 1972. Treatment of chronic Chagas infection with Lampit. *Bol Chil Parasitol* 27: 11-14.
- Schenone H, Rojas A, Alfaro E, Concha L, Aranda R 1981. Estudio longitudinal de la persistencia de la acción terapéutica del nifurtimox y del benznidazol en pacientes con infección chagásica crónica. *Bol Chil Parasitol* 36: 59-62.
- Schofield CJ, Dias JCP 1999. The Southern Cone Initiative against Chagas disease. *Adv Parasitol* 42: 1-27.
- Schmeda-Hirschmann G, Astudillo L, Bastida J, Codina C, Rojas de Arias A, Ferreira ME, Inchausti A, Yaluff G 2001. Cryptofolione derivatives from *Cryptocarya alba* fruits. *J Pharm Pharmacol* 53: 563-567.
- Schmunis GA 1994. American trypanosomiasis as a public health problem. In PAHO, *Chagas Disease and the Nervous System*. Scientific Publ n° 547, p. 3-29.
- Schmunis GA 2000. A tripanosomíase americana e seu impacto na saúde pública das américas. In Z Brener, Z Andrade, M Barral-Netto (eds), *Trypanosoma cruzi e Doença de Chagas*, 2ª ed., Guanabara Koogan, Rio de Janeiro, p. 1-15.
- Silva NN, Kuhn G, Santos JFC, Von Eye G, Chaer JAB 1974. Eficácia e tolerância do nitrofurfurilidene na fase crônica da moléstia de Chagas. *Rev Soc Bras Med Trop* 88: 325-334.
- Silveira JF 1992. *Trypanosoma cruzi* recombinant antigens for serodiagnosis. In S Wendel, Z Brener, E Camargo, A Rassi (eds), *Chagas Disease (American Trypanosomiasis): its Impact on Transfusion and Clinical Medicine*. ISBT, São Paulo, p. 207-218.
- Silveira JF, Umezawa ES, Luquetti AO 2001. Chagas disease: Recombinant *Trypanosoma cruzi* antigens for serological diagnosis. *Trends in Parasitology* 17: 286-291.
- Singh PK, Jones MM, Lane JE, Nessel A, Zimmerman LJ, Ribeiro-Rodrigues R, Richter A, Stenger MR, Carter CE 1997. Synthesis and *in vitro* trypanocidal activity of some novel iron chelating agents. *Arzneim Forsch* 47: 311-315.
- Solari A, Saavedra H, Sepulveda C, Oddó D, Acuña G, Labarca J, Muñoz S, Cuny G, Brengues C, Veas F, Bryan RT 1993. Successful treatment of *Trypanosoma cruzi* encephalitis in a patient with hemophilia and AIDS. *Clin Inf Dis* 16: 255-259.
- Sosa Estani S, Segura EL, Ruiz AM, Velazquez E, Porcel BM, Yampotis C 1998. Efficacy of chemotherapy with benznidazole in children in the indeterminate phase of Chagas disease. *Am J Trop Med Hyg* 59: 526-529.
- Souza DH, Garratt RC, Araújo AP, Guimarães BG, Jesus WD, Michels PA, Hannaert V, Oliva G 1998. *Trypanosoma cruzi* glycosomal glyceraldehyde-3-phosphate dehydrogenase: structure, catalytic mechanism and targeted inhibitor design. *FEBS Lett* 424: 131-135.
- Stoka V, Nycander M, Lenarcic B, Labriola C, Cazzulo JJ, Bjork I, Turk V 1995. Inhibition of cruzipain, the major cysteine proteinase of the protozoan parasite, *Trypanosoma cruzi*, by proteinase inhibitors of the cystatin superfamily. *FEBS Lett* 370: 101-104.
- Stoppani AOM 1999. The chemotherapy of Chagas disease. *Medicina (B Aires)* 59: 147-165.
- Storino R, Galleano R, Sosa R 1994. Tratamiento antiparasitario específico. In R Storino, J Milei (eds), *Enfermedad de Chagas*, Mosby Doyma, Argentina, p. 557-568.
- Sturm NR, Degraeve W, Morel CM, Simpson L 1989. Sensitive detection by amplification of kinetoplast minicircle DNA sequences: use in diagnosis of Chagas disease. *Mol Biochem Parasitol* 33: 205-214.
- Sundar S, Goyal AK, More DK, Singh MK, Murray HW 1998. Treatment of antimony-unresponsive Indian visceral leishmaniasis with ultra-short courses of amphotericin-B-lipid complex. *Ann Trop Med Parasitol* 92: 755-764.
- Sundar S, Makharia A, More DK, Agrawal G, Voss A, Fischer C, Bachmann P, Murray HW 2000. Short-course of oral miltefosine for treatment of visceral leishmaniasis. *Clin Infect Dis* 31: 1110-1113.
- Szajnman SH, Yan W, Bailey BN, DoCampo R, Elhalem E, Rodriguez JB 2000. Design and synthesis of aryloxyethyl thiocyanate derivatives as potent inhibitors of *Trypanosoma cruzi* proliferation. *J Med Chem* 43: 1826-1840.
- Tarleton RL 2001. Parasite persistence in the aetiology of Chagas disease. *Int J Parasitol* 31: 550-554.
- Teixeira AR, Cordoba JC, Souto Maior IC, Solorzano E 1990. Chagas disease: lymphoma growth in rabbits treated with benznidazole. *Am J Trop Med Hyg* 43: 146-158.
- Teixeira AR, Calixto MA, Teixeira ML 1994. Chagas disease: carcinogenic activity of the antitrypanosomal nitroarenes in mice. *Mutat Res* 305: 189-196.
- Thomas SM, McPhee DG 1984. Crystal violet: a direct-acting frameshift mutagen whose mutagenicity is enhanced by mammalian metabolism. *Mutation Res* 140: 165-167.
- Tomazela DM, Pupo MT, Passador EA, da Silva MF, Vieira PC, Fernandes JB, Fo ER, Oliva G, Pirani JR 2000. Pyranochalcones and a flavone from *Neoraputia magnifica* and their *Trypanosoma cruzi* glycosomal glyceraldehyde-3-phosphate dehydrogenase-inhibitory activities. *Phytochemistry* 55: 643-651.

- Tomimori-Yamashita J, Deps PD, Almeida DR, Enokihara MM, De Seixas MT, Freymuller E 1997. Cutaneous manifestation of Chagas disease after heart transplantation: successful treatment with allopurinol. *Br J Dermatol* 37: 626-630.
- Traub-Cseko YM, Ramalho-Ortigao JM, Dantas AP, De Castro SL, Barbosa HS, Downing KH 2001. Dinitroaniline herbicides against protozoan parasites: the case of *Trypanosoma cruzi*. *Trends Parasitol* 17: 136-141.
- Trouiller P, Rey JL, Bouscharain P 2000. Pharmaceutical development concerning diseases predominating in tropical regions: The concept of indigent drugs. *Ann Pharm Fr* 58: 43-46.
- Ullman B, Carter D 1997. Molecular and biochemical studies on the hypoxanthine-guanine phosphoribosyltransferases of the pathogenic haemoflagellates. *Int J Parasitol* 27: 203-213.
- Umezawa ES, Nascimento MS, Kesper Jr, Coura JR, Borges-Pereira J, Junqueira ACV, Camargo ME 1996. Immunoblot assay using excreted-secreted antigens of *Trypanosoma cruzi* in serodiagnosis of congenital, acute and chronic Chagas disease. *J Clin Microbiol* 37: 1554-1560.
- Umezawa ES, Nascimento MS, Stolf AMS 2001. Enzyme-linked immunosorbent assay with *Trypanosoma cruzi* excreted-secreted antigens (TESA-ELISA) for serodiagnosis of acute and chronic Chagas' disease. *Diag Microbiol Infect Disease* 39: 169-176.
- Urbina JA 1999. Chemotherapy of Chagas disease: the how and the why. *J Mol Med* 77: 332-338.
- Urbina JA, Payares G, Molina J, Sanoja C, Liendo A, Lazard K, Piras MM, Piras R, Perez N, Wincker P, Ryley JF 1996. Cure of short- and long-term experimental Chagas disease using D0870. *Science* 273: 969-971.
- Urbina JA, Moreno B, Vierkotter S, Oldfield E, Payares G, Sanoja C, Bailey BN, Yan W, Scott DA, Moreno SN, DoCampo R 1999. *Trypanosoma cruzi* contains major pyrophosphate stores, and its growth *in vitro* and *in vivo* is blocked by pyrophosphate analogs. *J Biol Chem* 274: 33609-33615.
- Urbina JA, Lira R, Visbal G, Bartroli J 2000. *In vitro* antiproliferative effects and mechanism of action of the new triazole derivative UR-9825 against the protozoan parasite *Trypanosoma (Schizotrypanum) cruzi*. *Antimicrob Agents Chemother* 44: 2498-2502.
- Viotti R, Vigliano C, Armenti H, Segura E 1994. Treatment of chronic Chagas disease with benzimidazole: clinical and serologic evolution of patients with long-term follow-up. *Am Heart J* 127: 151-162.
- Voller A, Draper C, Bidwell DE, Bartlett A 1975. A micro-plate enzyme-linked immunosorbent assay (ELISA) for Chagas disease. *The Lancet* 1: 426-429.
- Waechter AI, Yaluff G, Inchausti A, Rojas de Arias A, Hocquemiller R, Cavé A, Fournet A 1998. Leishmanicidal and trypanocidal activities of acetogenins isolated from *Ammonia glauca*. *Phytotherapy Res* 12: 541-544.
- Wang CC 1997. Validating targets for antiparasite chemotherapy. *Parasitology* 114 (Suppl): S31-44.
- Wendel S, Dias JCP 1992. Transfusion transmitted Chagas disease. In S Wendel, Z Brener, E Camargo, A Rassi (eds), *Chagas Disease (American trypanosomiasis): its Impact on Transfusion and Clinical Medicine*. ISBT, São Paulo, p. 103-134.
- WHO-World Health Organization 1984. Meeting on the development of trypanocidal compounds for the sterilization of blood UNDP/WB/TDR, Geneva.
- WHO-World Health Organization 1997. Chagas disease. Thirteenth Programme Report UNDP/TDR, Geneva.
- Wieder T, Reutter W, Orfanos CE, Geilen CC 1999. Mechanisms of action of phospholipid analogs as anticancer compounds. *Prog Lipid Res* 38: 249-259.
- Winker P, Britto C, Borges Pereira, Cardoso MA, Coleman W, Morel CM 1994. Use of simplified polymerase chain reaction procedures to detect, *Trypanosoma cruzi* in blood samples from chronic chagasic patients in rural area. *Am J Trop Med Hyg* 51: 771-777.
- Yardley V, Croft SL 1999. *In vitro* and *in vivo* activity of amphotericin B-lipid formulations against experimental *Trypanosoma cruzi* infections. *Am J Trop Med Hyg* 61: 193-197.
- Yong V, Schmitz V, Vannier-Santo MA, de Lima AP, Lalmanach G, Juliano L, Gauthier F, Scharfstein J 2000. Altered expression of cruzipain and a cathepsin B-like target in a *Trypanosoma cruzi* cell line displaying resistance to synthetic inhibitors of cysteine-proteinases. *Mol Biochem Parasitol* 109: 47-59.
- Zaidenberg A, Tournier H, Schinella G, Marín G, Buschiazzo H 1999. Effects of trifluralin on *Trypanosoma cruzi* *in vitro* and *in vivo*. *Pharmacol Toxicol* 84: 98-100.
- Zuccotto F, Brun R, Gonzalez Pacanowska D, Ruiz Perez LM, Gilbert IH 1999. The structure-based design and synthesis of selective inhibitors of *Trypanosoma cruzi* dihydrofolate reductase. *Bioorg Med Chem Lett* 9: 1463-1468.
- Zuccotto F, Martin AC, Laskowski RA, Thornton JM, Gilbert IH 1998. Dihydrofolate reductase: a potential drug target in trypanosomes and leishmania. *J Comput Aided Mol Des* 12: 241-257.