

## SIMULTANEOUS DETERMINATION OF NON-STEROIDAL ANTI-INFLAMMATORY DRUGS IN PHARMACEUTICAL FORMULATIONS AND HUMAN SERUM BY REVERSED PHASE HIGH PERFORMANCE LIQUID CHROMATOGRAPHY

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A rapid and sensitive method using high performance liquid chromatography has been developed and validated for the simultaneous determination of non-steroidal anti-inflammatory drugs (NSAIDs) in pharmaceutical formulations and human serum. Six NSAIDs including: naproxen sodium, diclofenac sodium, meloxicam, flurbiprofen, tiaprofenic and mefenamic acid were analyzed simultaneously in presence of ibuprofen as internal standard on Mediterranea C<sub>18</sub> (5 µm, 250 x 0.46 mm) column. Mobile phase comprised of methanol: acetonitrile: H<sub>2</sub>O (60:20:20, v/v; pH 3.35) and pumped at a flow rate of 1 mL min<sup>-1</sup> using 265 nm UV detection. The method was linear over a concentration range of 0.25-50 µg mL<sup>-1</sup> (r<sup>2</sup> = 0.9999).

Keywords: NSAIDs; serum; reversed phase high performance liquid chromatography.

### INTRODUCTION

Non-steroidal anti-inflammatory drugs (NSAIDs) (Figure 1) have analgesics, antipyretics and anti-inflammatory activities and are widely used in the treatment of acute and chronic pain, osteoarthritis, rheumatoid arthritis and related conditions. The pharmacological actions of NSAIDs are related to inhibition of cyclooxygenase (COX), a

key enzyme of prostaglandin biosynthesis at the site of inflammation. The simultaneous measurement of these NSAIDs concentrations in biological samples is required in clinical and toxicological screening, pharmacokinetic studies, as well as in therapeutic monitoring. Furthermore, it is also very important to precisely quantify these NSAIDs in pharmaceutical formulations for quality control.<sup>1</sup>

Until now, a variety of chromatographic methods have been proposed for the determination of NSAIDs in pharmaceutical formulations and biological fluids, by gas chromatography,<sup>2,4</sup> spectrofluorometry,<sup>5</sup> high performance liquid chromatography<sup>6-17</sup> and capillary electrophoresis.<sup>18</sup>

Generally, it is impossible that one patient would be prescribed for more than one kind of NSAIDs at the same time; however simultaneous determination of these drugs is useful in pharmaceutical routine analysis and biological samples.

This paper reports the simultaneous determination of NSAIDs in pharmaceutical formulations and human serum. It is precise, accurate and rapid method.

### EXPERIMENTAL

#### Materials and reagents

Pure tiaprofenic acid was obtained from Aventis Pharma (Pvt) Ltd, meloxicam from AGP (Pvt) Ltd, diclofenac sodium, flurbiprofen, ibuprofen and naproxen sodium from Pharmevo (Pvt) Ltd. Dosages form of Tiaprofenic acid (Surgam® 300 mg) diclofenac sodium (Voltral® 50 mg) flurbiprofen (Synalgo® 100 mg), meloxicam (Melfax® 15 mg) and mefenamic acid (Ponstan® 250 mg) were purchased from local market. Methanol and acetonitrile (analytical grade) were purchased from Tedia, USA. Deionized water was used to prepare a mobile phase.

#### Instrumentation and chromatographic conditions

The development of the method and validation of the work was performed on a liquid chromatographic system consisting of Shimadzu model LC-10AT VP pump with a SPD-10AT VP, variable wavelength UV-Visible detector. Chromatographic system was integrated via Shimadzu model CBM-102 Communication Bus Module

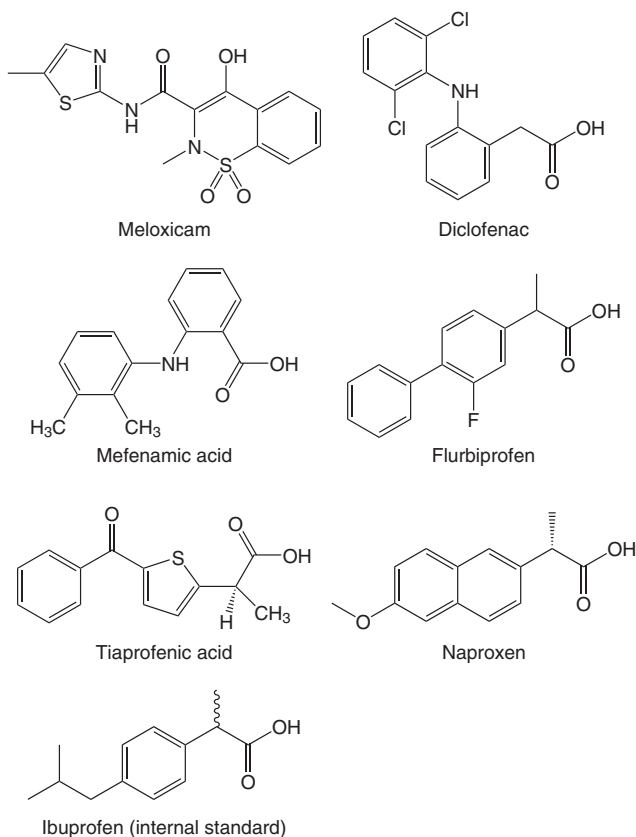


Figure 1. Chemical structures of non-steroidal anti-inflammatory drugs

to a Pentium-4 PC. Analysis was conducted on a Mediterranea C18 (Teknokroma®, Karachi, Pakistan) (5  $\mu\text{m}$ , 250 x 0.46 mm) column, mobile phase consisted of methanol: acetonitrile:  $\text{H}_2\text{O}$  (60:20:20, v/v) and pH was adjusted to 3.35 (apparent pH) with orthophosphoric acid. Mobile phase was filtered daily using 0.45  $\mu\text{m}$  membrane filter (Millipore, Germany) and degassed in an ultrasonic bath. The samples were introduced through a rheodyne injector valve with a 20-mL sample loop. Assays were performed at ambient temperature at a flow rate of 1  $\text{mL min}^{-1}$  and detection at 265 nm.

### Preparation of standard solutions

Standard solution of non-steroidal anti-inflammatory drugs and internal standard (100  $\mu\text{g mL}^{-1}$ ) were prepared by dissolving them in methanol. Internal standard solution was further diluted with methanol to give concentration 50  $\mu\text{g mL}^{-1}$ .

### Preparation of calibration curves

Appropriate dilution of standard solution (100  $\mu\text{g mL}^{-1}$ ) of non-steroidal anti-inflammatory drugs were made using methanol to obtain solution of concentration of 0.25, 1.0, 2.50, 5.0, 12.50, 25.0 and 50.0  $\mu\text{g mL}^{-1}$ . A correlation between peak area and concentration 0.25-50  $\mu\text{g mL}^{-1}$  of non-steroidal anti-inflammatory drugs were established and calibration curves were obtained.

### Limit of detection and quantification (LOD and LOQ)

The limit of detection is the lowest quantity of analyte in any sample which can be detected but cannot be quantified where as the quantitation limit of any analytical procedure is the lowest amount of analyte in a sample which can be quantitatively analyzed with suitable precision and accuracy. The equation used to calculate LOD and LOQ are:

$$\text{LOD} = 3.3\sigma/S \text{ and } \text{LOQ} = 10\sigma/S$$

$\sigma$  = standard deviation of the lowest standard concentration,  $S$  = slope of the standard curve.

### Precision and accuracy

The precision of proposed method was evaluated through intra-day (repeatability) and inter-day (intermediate precision) at concentration of 0.25, 2.50 and 12.50  $\mu\text{g mL}^{-1}$ . Three replicates were made for intra- and inter-day study. The precision is expressed as percentage relative standard deviation (% RSD). To evaluate the accuracy of the proposed method, recovery tests for all analytes were performed by adding known amounts of standard solutions to samples followed by analysis using proposed method. Accuracy was expressed as percentage recovery and determined at three concentration levels (0.25, 2.50 and 12.50  $\mu\text{g mL}^{-1}$ ). Three replicates were made for accuracy study.

### Selectivity

The selectivity of the method was evaluated by analyzing blank drug-free serum samples obtained from healthy volunteer and spiked serum samples. The probable interferences from endogenous substances were assessed by observing the chromatograms of blank and spiked serum samples. Non-steroidal anti-inflammatory drugs pharmaceutical formulations were also analyzed and compared with the standard chromatograms of drugs for excipients interference.

### Dosage analysis

The contents of 20 tablets of tiaprofenic acid (Surgam® 300 mg) diclofenac sodium (Voltral® 50 mg) flurbiprofen (Synalgo® 100 mg), meloxicam (Melfax® 15 mg) and mefenamic acid (Ponstan® 250 mg) were weighed and finely grounded. A mass of powder equivalent to the average mass of a tablet was transferred to volumetric flasks with methanol. Solutions were sonicated in ultrasonic bath for 10 min and then diluted to volume with methanol. These primary stock solutions were filtered through Whatman filter paper and the filtrates were further diluted suitably to prepare a secondary stock solution. Aliquots of the secondary stock solution were diluted to a concentration of 0.25, 2.50 and 12.50  $\mu\text{g mL}^{-1}$  and the samples were analyzed using above mentioned chromatographic conditions.

### Serum drug analysis

The recoveries of non-steroidal anti-inflammatory drugs from pooled human serum were determined by the stated chromatographic conditions. Multiple blood samples (10 mL) of 15 healthy non-smoker volunteers (age ranging from 21-25 years), not taking any other medications were collected in glass tubes. 10 mL of acetonitrile was then added to each serum samples to precipitate the proteins. After 15 min of centrifugation at 3000 rpm, all the upper layers from the serum samples were put together to receive a representative pool of human serum samples. This pool of serum samples was mixed with NSAIDs and internal standard in 1:1:1 ratio, filtered and analyzed.<sup>19,20</sup>

## RESULTS AND DISCUSSION

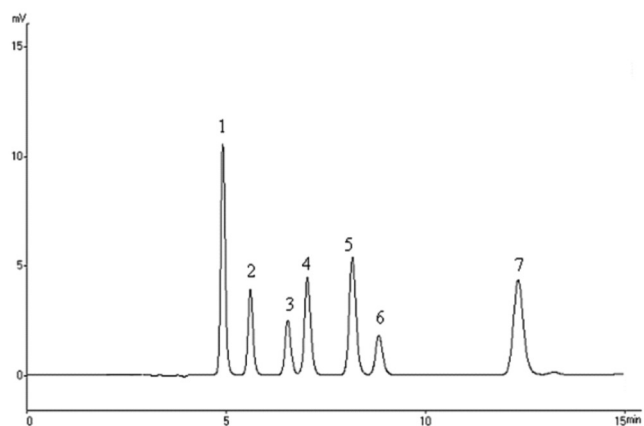
Isocratic-mode of HPLC with UV detection was developed for the determination of non-steroidal anti-inflammatory drugs in pharmaceutical formulations and in human serum. Serum deproteinization for the determination of drugs is commonly accepted as the simplest methods as compared liquid-liquid extraction or solid-phase extraction from plasma samples and usually have high reproducibility.

### Optimization of chromatographic conditions

In order to achieve a suitable separation of analytes from endogenous compounds of serum matrix, the isocratic elution mode was chosen. The method was optimized by variation in pH of the mobile phase and the percentage and nature of the organic modifier (acetonitrile or methanol).

Firstly, an attempt was made regarding the composition of the mobile phase and pH. Proportions of water higher than 20% gave rise to chromatographic signals which suffered major frontings. Higher percentage of methanol than 60% results in long retention time and 20% of acetonitrile was found suitable in mobile phase. When percentage of acetonitrile was increased in mobile phase peaks were emerged. Chromatographic peaks obtained at pH 3.35 were sharp (Figure 2) than those observed at pH 4 and also absorbance of endogenous components of serum was lower at pH 3.35 (Figure 3a and 3b). So mobile phase consisting of methanol:acetonitrile: $\text{H}_2\text{O}$  (60:20:20, v/v) and pH 3.35 was optimal for the separation of non-steroidal anti-inflammatory drugs and it provides high sensitivity.

The system suitability was performed by studying the parameters which include column efficiency or theoretical plates (N), resolution (R), peak asymmetry or tailing factor ( $A_s$ ) and capacity factor (k). The low value of peak asymmetry or tailing factor and high values of column efficiency or theoretical plates, resolution and capacity factor indicated the suitability and proper selection of mobile phase. These parameters are presented in Table 1.



**Figure 2.** Chromatogram of non-steroidal anti-inflammatory drugs in the presence of ibuprofen as internal standard: (1) tiaprofenic acid, (2) meloxicam, (3) naproxen sodium, (4) flurbiprofen, (5) diclofenac sodium, (6) ibuprofen, (7) mefenamic acid

**Table 1.** Chromatographic data for separation of NSAIDs on a Mediterranea C<sub>18</sub> column under the optimum separation conditions

Analytes	K <sup>a</sup>	As <sup>b</sup>	N <sup>c</sup>	Rs <sup>d</sup>
Tiaprofenic	1.17	1.19	9088	10.15
Meloxicam	1.47	1.14	9308	3.17
Naproxen	1.89	1.17	9136	3.73
Flurbiprofen	2.11	1.12	9789	1.77
Diclofenac	2.61	1.13	10699	3.78
Mefenamic	4.45	1.13	11364	8.79

<sup>a</sup>capacity factor; <sup>b</sup>peak asymmetry; <sup>c</sup>number of theoretical plates, <sup>d</sup>resolution

Method development and validation was carried out according to ICH validation guidelines.<sup>21</sup>

### Linearity and sensitivity

The linearity of the method was evaluated by injecting six replicate of different concentrations. Linearity was observed in the concentration range of 0.25-50 µg mL<sup>-1</sup>. For the determination of linearity, standard calibration curve was used. The statistical results of the linearity, LOD and LOQ values are given in Table 2.

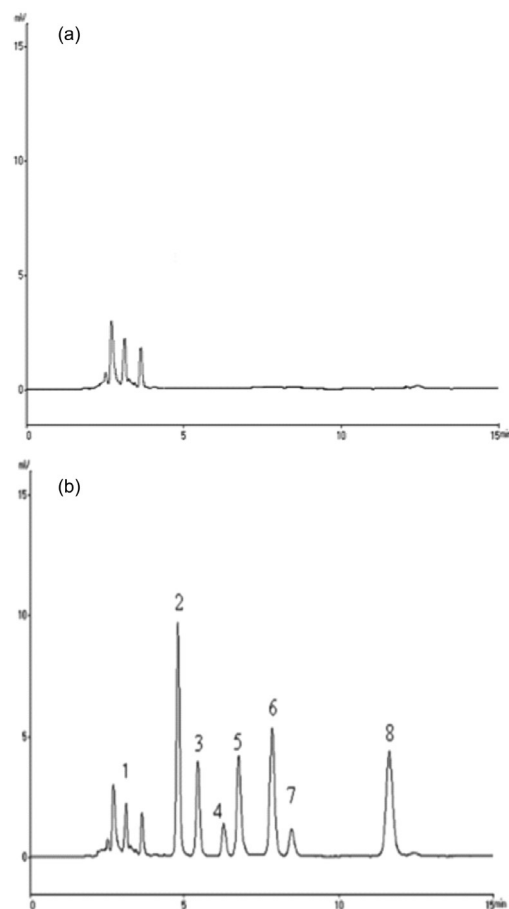
### Precision and accuracy

The precision was expressed as percentage relative standard deviation (% RSD) and it ranged in 0.02-2.76% and 0.01-1.54% for

**Table 2.** Calibration curves and limit of detection and quantification of non-steroidal anti-inflammatory drugs in pharmaceutical formulations

Analytes	Concentration range (µg mL <sup>-1</sup> )	Regression equation	r <sup>2</sup> <sup>a</sup>	LOD <sup>b</sup>	LOQ <sup>c</sup>
				(µg mL <sup>-1</sup> )	
Tiaprofenic	0.25-50	y = 1635.00x + 943.58	0.9995	0.004	0.013
Meloxicam	0.5-50	y = 681.90x + 380.18	0.9997	0.005	0.015
Naproxen	1.0-50	y = 510.37x + 354.13	0.9996	0.007	0.023
Flurbiprofen	0.5-50	y = 966.21x + 535.10	0.9996	0.005	0.016
Diclofenac	0.25-50	y = 1286.40x + 479.23	0.9998	0.003	0.009
Mefenamic	0.5-50	y = 1518.20x + 854.37	0.9995	0.002	0.007

<sup>a</sup>Correlation of coefficient, <sup>b</sup>limit of detection (S/N=3), <sup>c</sup>limit of quantification (S/N=10)



**Figure 3.** Chromatograms of: (a) drug-free human serum; (b) non-steroidal anti-inflammatory drugs spiked in human serum: (1) serum peaks, (2) tiaprofenic acid, (3) meloxicam, (4) naproxen sodium, (5) flurbiprofen, (6) diclofenac sodium, (7) ibuprofen, (8) mefenamic acid

intra- and inter-day respectively in pharmaceutical formulations while in serum it was less than 3% (Table 3). Accuracy was expressed as percentage recovery and it was determined at three concentration levels. Three replicates were made for accuracy study, intra- and inter-day percentage recoveries from pharmaceutical formulations ranged in 98-102% and 97-102%, respectively. Recoveries from human serum ranged in 97-101% (Table 4).

### Selectivity

The chromatograms obtained from blank, spiked serum samples and pharmaceutical formulations were identical with that obtained chromatogram from standard solution of NSAIDs. There was no peak

**Table 3.** Accuracy and precision of non-steroidal anti-inflammatory drugs in pharmaceutical formulations

<sup>a</sup> Conc. added ( $\mu\text{g mL}^{-1}$ )	Found ( $\mu\text{g mL}^{-1}$ )	Intra-day		Found ( $\mu\text{g mL}^{-1}$ )	Inter-day	
		RSD% <sup>b</sup>	% Rec. <sup>c</sup>		RSD%	% Rec.
Tiaprofenic						
0.25	0.26	0.94	100	0.25	1.00	98
2.50	2.49	1.97	100	2.50	3.86	100
12.50	12.50	1.26	100	12.51	0.11	100
Diclofenac sodium						
0.25	0.25	1.23	101	0.25	0.99	99
2.50	2.56	0.65	102	2.50	0.18	100
12.50	12.46	1.25	100	12.49	0.05	100
Flurbiprofen						
0.50	0.49	1.75	98	0.51	0.77	102
2.50	2.51	0.34	100	2.50	0.29	100
12.0	12.5	1.28	100	12.51	0.06	100
Mefenamic acid						
0.25	0.24	1.23	97	0.25	1.20	98
2.50	2.49	0.13	100	2.50	0.10	100
12.50	12.49	1.53	100	12.50	0.01	100
Meloxicam						
0.50	0.52	0.14	102	0.49	1.54	97
2.50	2.51	0.52	100	2.49	0.61	100
12.50	12.48	2.76	100	12.48	0.04	100
Naproxen						
1.00	0.98	0.79	98	1.00	1.00	99
2.50	2.56	3.22	102	2.49	5.10	100
12.50	12.49	0.21	100	12.49	1.12	100

<sup>a</sup>Concentration, <sup>b</sup>relative standard deviation, <sup>c</sup>% recovery

**Table 4.** Accuracy and precision of non-steroidal anti-inflammatory drugs in human serum

Conc. added ( $\mu\text{g mL}^{-1}$ )	Found ( $\mu\text{g mL}^{-1}$ )	% recovery	Precision	
			Intra-day RSD%	Inter-day RSD%
Tiaprofenic				
0.25	0.24	97	1.02	0.95
2.50	2.47	99	0.97	1.23
12.50	12.44	100	2.01	1.65
Diclofenac				
0.25	0.26	98	1.14	2.11
2.50	2.48	99	2.70	2.10
12.50	12.49	100	1.05	0.98
Flurbiprofen				
0.50	0.50	98	1.02	0.85
2.50	2.52	101	0.46	1.01
12.50	11.48	100	1.23	3.86
Mefenamic				
0.25	0.25	102	0.52	0.76
2.50	2.48	99	0.20	0.89
12.50	12.49	100	2.76	1.24
Meloxicam				
0.50	0.49	98	0.39	0.41
2.50	2.49	100	1.02	1.12
12.50	12.49	100	0.15	0.89
Naproxen				
1.00	1.06	101	0.75	0.65
2.50	2.47	101	1.96	1.26
12.50	12.50	100	1.23	0.92

observed when the analysis of placebo solution of NSAIDs were done. In addition, peak purity index of all analytes were investigated and found to be >0.999. Thus method demonstrated good resolution and found to be free of interferences.

### Ruggedness

Method ruggedness was evaluated by two analysts performing assay on separate lots of non-steroidal anti-inflammatory drugs. Each analyst prepared samples in triplicate and used separate instruments, reagents, diluent and mobile phase solutions. Relative standard deviation ( $n = 3$ ) of for all of the samples for each lot was less than 2.0% indicating acceptable robustness. The method did not show any notable deviation in results from acceptable limits.

### CONCLUSIONS

The proposed method using reversed phase high performance liquid chromatography was found to be suitable for simultaneous determination of non-steroidal anti-inflammatory drugs in pharmaceutical formulations and human serum. The detector response was found to be linear over a wide concentration range. Results are accurate and precise and are confirmed by the statistical parameters. The precision and accuracy of the method are well within the limits required for bioanalytical assays. The lower limit of quantification permitted the use of the method for pharmacokinetic studies.

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