

Floristic and vegetation structure of a granitic grassland in Southern Brazil

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ABSTRACT – (Floristic and vegetation structure of a granitic grassland in Southern Brazil). A floristic and structural survey of a natural grassland community was conducted on Morro do Osso, a granitic hill in Porto Alegre, RS, Brazil. Structural data were surveyed in 39 one square meter plots placed over two major grassland areas. An accidental fire has occurred in one of the areas approximately one year prior to our survey, leading to further analysis of parameters differences between sites. The floristic list contains 282 species, whereas the structural survey has found 161 species. Families with highest accumulated importance values were Poaceae, Asteraceae and Fabaceae. The diversity and evenness indexes were 4.51 nats ind⁻¹ and 0.86, respectively. Cluster analysis denoted two groups coinciding with the areas distinguished by the fire disturbance. A similarity analysis between our data and two other data sets from nearby granitic hills resulted in 28% to 35% similarity, with equivalent species-family distribution and many common dominant species, corroborating the concept of a continuous flora along the South Brazilian granitic hills.

Key words - campos, diversity, fire, relict vegetation, similarity, threatened species

RESUMO – (Florística e estrutura da vegetação de um campo granítico no Sul do Brasil). Um levantamento florístico e estrutural de uma comunidade de campo natural foi conduzido no Morro do Osso, um morro granítico em Porto Alegre, RS, Brasil. Os dados estruturais foram obtidos em 39 quadros de 1 m², dispostos em duas grandes áreas de campo. Uma queimada acidental ocorreu em uma das áreas, aproximadamente um ano antes do levantamento, propiciando a análise de diferenças entre os parâmetros dos dois sítios de amostragem. A lista florística contém 282 espécies e, no levantamento estrutural, foram encontradas 161 espécies. As famílias com maior valor de importância acumulado foram Poaceae, Asteraceae e Fabaceae. Os índices de diversidade e a equabilidade foram 4,51 nats ind⁻¹ e 0,86, respectivamente. A análise de agrupamento identificou dois grupos nítidos, que coincidem com os locais atingidos ou não pelo fogo. Uma análise de similaridade entre os dados obtidos e dois outros conjuntos de dados de morros graníticos vizinhos resultou em 28% a 35% de similaridade, com uma equivalente distribuição de espécies por família e muitas espécies dominantes comuns, corroborando a noção de continuidade da flora ao longo dos morros graníticos no sul do Brasil.

Palavras-chave - campos, diversidade, espécies ameaçadas, fogo, similaridade, vegetação relictual

Introduction

The study of natural grassland communities, despite its well known ecological importance, has been historically neglected (Overbeck *et al.* 2007) or limited to dominant grass species used in livestock production (Uys *et al.* 2004) or grain monocultures. As it is, there is a lack of floristic and structural studies concerning these communities worldwide, which leads, at least partly,

to an underestimation of their richness, diversity and conservational value.

Subtropical grasslands, which are also called *campos* in Brazil (Behling 2002, Focht & Pillar 2003, Overbeck *et al.* 2007), were dominant in southern Brazilian landscape during late Pleistocene, and its present distribution can be interpreted as a remnant of a drier and cooler climate in the region (Behling 2002, Bredenkamp *et al.* 2002). Thus, *campos* configure a relict formation, and the knowledge of its floristic and structural features is vital to understand the processes that lead to modern landscape. Rambo (1954) emphasized both the richness and ecological importance of the southern Brazilian herbaceous flora by indicating that herbaceous species of Asteraceae alone are more numerous than the entire local woody flora.

Despite its ecological and social-economic importance, natural grasslands are a historic target of many anthropogenic disturbances, such as land

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conversion to grain and wood monocultures, overgrazing and unrestricted fire, being often wiped away from the landscape even before being well known by science. Nevertheless, fire disturbances not always have hazardous consequences for plant communities, such as depletion of soil nutrients and local plant extinction, and there is growing evidence of the importance of both natural and human-driven fires for the maintenance of many natural grassland communities (Quadros & Pillar 2001, Uys *et al.* 2004, Overbeck *et al.* 2005, Overbeck *et al.* 2006, Behling *et al.* 2007, Müller *et al.* 2007). Moreover, the acknowledgement of fire disturbance as a crucial environmental factor to the very existence of subtropical grasslands is increasing (Quadros & Pillar 2001, Behling 2002, Behling *et al.* 2004, Overbeck *et al.* 2005, Müller *et al.* 2007).

Granitic grassland formations in southern Brazil includes an area beside the city of Porto Alegre, Rio Grande do Sul, represented by a series of granitic hills known as Morro do Osso, Morro Santana, Morro da Polícia, Morro São Pedro, among others. Although recent floristic and structural surveys concerning southern Brazilian granitic grassland formations are scarce, a historical analysis of the Porto Alegre flora can be found in Rambo (1954), who displayed an extensive species list, albeit outdated nowadays. The most extensive approach was presented by Aguiar *et al.* (1986) concerning a preliminary floristic survey in ten granitic hills in Porto Alegre region. Boldrini *et al.* (1998) surveyed floristic and ecological aspects of a grassland community at Morro da Polícia and Overbeck *et al.* (2006) presented a local species list of Morro Santana, linking floristic composition of burned grassland to environmental factors, such as soil properties and distance from the forest border.

Working in a southern Brazilian granitic grassland, our objectives were: (1) to present floristic and structural data of the local herbaceous flora; (2) to explore the effects of an accidental fire event on floristic and structural patterns and (3) to compare plant diversity of similar hill's formation in the context of grassland vegetation dynamic.

Material and methods

Study area – The study was carried out on Parque Natural Morro do Osso (30°07' S, 51°14' W; 143 m a.s.l.), an area inserted into a chain of granitic hills that surrounds the city of Porto Alegre, Rio Grande do Sul, Brazil. This geological formation, known by the presence of several rare and endemic plant species (Rambo 1954, Aguiar *et al.* 1986, Boldrini *et al.* 1998, Brack *et al.* 1998), configures an intersection

point among four different structural provinces that shape the south Brazilian landscape: the Crystalline Shield, the Peripheral Depression, the bottom of the Seashore Plain and the Lagoon System (Menegat *et al.* 1998). Likewise, this granitic chain is historically thought to be responsible for notorious local plant endemism, acting like a refuge area during ocean transgression times (Rambo 1954). The study site is a Conservation Unit since 1994, covering 127 ha of natural area (Brack *et al.* 1998).

Natural vegetation at the site comprises a mosaic of grasslands and forests (Atlantic Rainforest), the first occurring mostly on hilltops and north or northwest slopes and the latter mostly along river courses and south or southeast slopes (Rambo 1954, 1956, Müller *et al.* 2007). Grasslands cover around 40% of the study area and fire, either natural or caused by neighboring human settlements, is a common disturbance to the ecosystem at spaced but non-fixed intervals. There are two major grassland patches in the study site, one is west-oriented and the other is east-oriented, being separated by a relatively large forest/woodland area (figure 1). There is a record of significant burning on the east grassland area in 2005, one year prior to the survey presented in this paper, confirmed by the first collected specimens at the site, which had clear signals of recent burning.

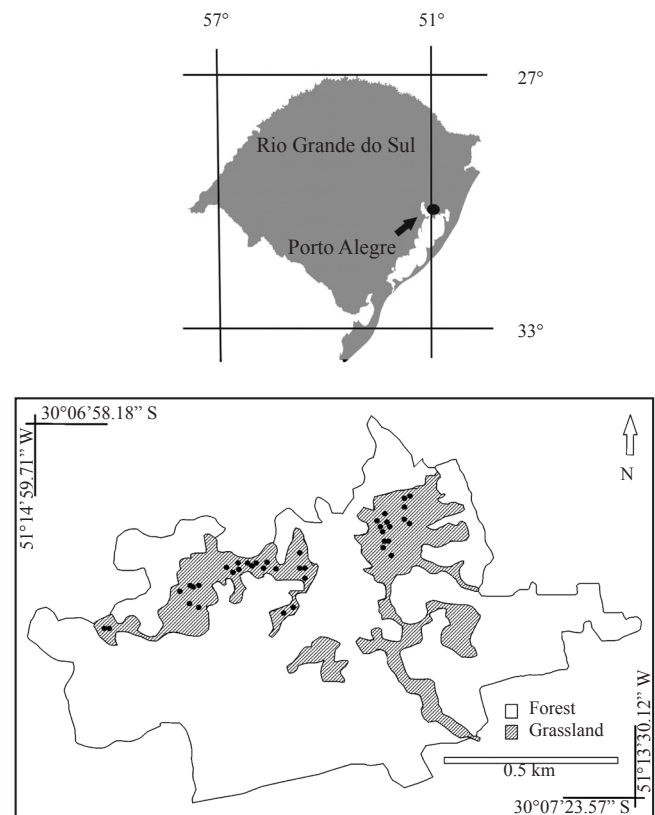


Figure 1. Spatial location of the study site Parque Natural Morro do Osso (30°07' S, 51°14' W) and the Porto Alegre city, Rio Grande do Sul State, Brazil. Black dots on grassland areas represent (not in scale) the 39 sampling units.

Climate at the study site is subtropical humid, with no dry season (Köppen's Cfa). The average annual temperature is 19.5 °C and the mean annual precipitation is 1348 mm (Nimer 1990). The soil is granite-originated, composed by post-tectonic granitoids dated from 550 m.y.a. and known as Santana granite (Sestren-Bastos 2006).

Data collection and analysis – Data for the structural survey were collected in 39 one square meter plots randomly placed in two major grassland areas (figure 1). Other grassland-covered areas that can be seen in figure 1 comprise tracks, shrublands and surroundings of Park management's buildings, thus not included in the sampling area. In each plot, species cover was registered, applying the Daubenmire scale (Mueller-Dombois & Ellenberg 1974). The survey took place in January 2006, although prior to that, other expeditions were made to search floristic records. Plant identification was done using specific bibliography and contacting specialists when necessary. Family circumscription follows APWeb (2003). A full species list for the Morro do Osso's grasslands is presented based on our own information and previous publications (Brack *et al.* 1998, Brack *et al.* 2001).

For each species, absolute and relative frequency (AF and RF, respectively), absolute and relative cover (AC and RC, respectively) and importance value (IV) were calculated. Shannon-Wiener diversity index (H') and the Pielou evenness (E) (Pielou 1969, Magurran 1988) were also obtained. Open soil and litter were evaluated in each plot unit, but omitted in the community indexes calculation.

The raw data matrix obtained for the grassland community (species plus open soil and litter per 39 sampling plots) was submitted to ordination and cluster analysis on Multiv software

(Pillar & Orlóci 1993, 1996, Pillar 2006). Both methods were submitted to the bootstrap analysis to verify cluster groups sharpness and ordination axis stability (Pillar 1999a, b). The ordination method was the principal coordinate analysis (PCoA) and, in the cluster analysis, incremental sum of squares was used as the clustering criterion, both based on chord distance resemblance measure between plots (Podani 2000). Variables (species, litter and open soil cover) with axes correlation values greater than 0.45 were included in the ordination scatter diagram. The statistically supported groups obtained from this analysis were then submitted to separate phytosociological analysis with the previously mentioned parameters. Vegetation height, richness, open soil and litter were submitted to variance analysis comparing groups of plots by randomization test with 10,000 iterations (Pillar & Orlóci 1996, Manly 1997).

The species list obtained in the structural survey was submitted to a similarity analysis (Jaccard similarity index) based on structural data from two neighboring granitic hills: Morro da Polícia (Boldrini *et al.* 1998) and Morro Santana (Overbeck *et al.* 2006). Only fully identified taxa (specific level) were included in this analysis. A nomenclatural revision was also done on the species lists in order to avoid synonyms and standardize data.

Results

Floristic and structural analysis – The floristic list contains 282 species, distributed in 42 families (table 1). Poaceae (74 species), Asteraceae (63 species) and Fabaceae (25 species) were the most representative families in the floristic survey.

Table 1. Floristic list of grassland formations at Morro do Osso. (F = Species collected in field expeditions; R = Species appointed for the study site by Brack *et al.* (1998; 2001); Ph = Species sampled in the structural survey).

Family	Species	
Acanthaceae	<i>Ruellia morongii</i> Britton	Ph
	<i>Stenandrium diphyllum</i> Nees	Ph
Amaranthaceae	<i>Gomphrena globosa</i> L.	R
	<i>Gomphrena graminea</i> Moq.	Ph
	<i>Pfafia tuberosa</i> (Sprengel) Hicken	Ph
Amaryllidaceae	<i>Habranthus gracilifolius</i> Herb.	F
Anacardiaceae	<i>Schinus weinmannifolius</i> Engl.	Ph
Apiaceae	<i>Apium leptophyllum</i> F. Muell. ex Benth.	F
	<i>Eryngium ciliatum</i> Cham. & Schltdl.	Ph
	<i>Eryngium elegans</i> Cham. & Schltdl.	F
	<i>Eryngium eriophorum</i> Cham. & Schltdl.	R
	<i>Eryngium horridum</i> Malme	Ph
	<i>Eryngium pristis</i> Cham. & Schltdl.	Ph
	<i>Eryngium sanguisorba</i> Cham. & Schltdl.	F
	<i>Hydrocotyle umbellata</i> L.	R

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Family	Species	
Apocynaceae	<i>Asclepias mellodora</i> A. St.-Hil.	Ph
	<i>Macrosiphonia longiflora</i> (Desf.) Müll. Arg.	F
	<i>Mandevilla coccinea</i> (Hook. & Arn.) Woodson	Ph
	<i>Oxypetalum arnottianum</i> H. Buek	R
	<i>Oxypetalum</i> sp. 1	Ph
	<i>Oxypetalum</i> sp. 2	F
Asteraceae	<i>Achyrocline satureioides</i> (Lam.) DC.	R
	<i>Acmella bellidioides</i> (Smith) R.K. Jansen	Ph
	<i>Aspilia montevidensis</i> (Spreng.) Kuntze	Ph
	<i>Baccharis articulata</i> (Lam.) Pers.	Ph
	<i>Baccharis cultrata</i> Baker	Ph
	<i>Baccharis cylindrica</i> (Less.) DC.	Ph
	<i>Baccharis dracunculifolia</i> DC.	Ph
	<i>Baccharis ochracea</i> Spreng.	Ph
	<i>Baccharis pseudomyriocephala</i> I. L. Teodoro	Ph
	<i>Baccharis riograndensis</i> I. L. Teodoro & J. Vidal	Ph
	<i>Baccharis sessiliflora</i> Vahl	Ph
	<i>Baccharis trimera</i> (Less.) DC.	Ph
	<i>Calea stenophylla</i> Baker	Ph
	<i>Calea uniflora</i> Less. var. <i>discoidea</i> Baker	F
	<i>Calea uniflora</i> Less. var. <i>uniflora</i>	Ph
	<i>Chaptalia integerrima</i> (Vell.) Burkart	Ph
	<i>Chaptalia piloselloides</i> (Vahl) Baker	Ph
	<i>Chaptalia runcinata</i> Kunth	Ph
	<i>Chevreulia acuminata</i> Less.	Ph
	<i>Chevreulia sarmentosa</i> (Pers.) S. F. Blake	Ph
	<i>Conyza chilensis</i> Spreng.	Ph
	<i>Criscia stricta</i> (Spreng.) Katinas	F
	<i>Eupatorium ascendens</i> Sch. Bip. ex Baker	Ph
	<i>Eupatorium congestum</i> Hook. & Arn.	Ph
	<i>Eupatorium laetevirens</i> Hook. & Arn.	Ph
	<i>Eupatorium ligulaefolium</i> Hook. & Arn.	Ph
	<i>Eupatorium squarrulosum</i> Hook. & Arn.	Ph
	<i>Eupatorium subhastatum</i> Hook. & Arn.	Ph
	<i>Eupatorium tanacetifolium</i> Gillies ex Hook. & Arn.	F
	<i>Eupatorium tremulum</i> Hook. & Arn.	R
	<i>Gamochoaeta filaginea</i> (DC.) Cabrera	Ph
	<i>Gochnatia cordata</i> Less.	R
	<i>Gochnatia orbiculata</i> (Malme) Cabrera	R
	<i>Heterothalamus psiadioides</i> Less.	Ph
	<i>Hypochaeris glabra</i> L.	Ph
	<i>Isostigma peucedanifolium</i> Less.	F
	<i>Lucilia acutifolia</i> (Poir.) Cass.	Ph
<i>Lucilia nitens</i> Less.	Ph	
<i>Noticastrum decumbens</i> (Baker) Cuatrec.	Ph	
<i>Orthopappus angustifolius</i> (Sw.) Gleason	Ph	
<i>Porophyllum lanceolatum</i> DC.	F	
<i>Porophyllum obscurum</i> (Spreng.) DC.	Ph	
<i>Pterocaulon alopecuroides</i> (Lam.) DC.	Ph	

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Family	Species	
	<i>Pterocaulon angustifolium</i> DC.	Ph
	<i>Pterocaulon polypterum</i> (DC.) Cabrera	Ph
	<i>Schlechtendalia luzulifolia</i> Less.	F
	<i>Senecio cisplatinus</i> Cabrera	F
	<i>Senecio heterotrichius</i> DC.	F
	<i>Senecio leptolobus</i> DC.	R
	<i>Stenachaenium campestre</i> Baker	Ph
	<i>Stenachaenium macrocephalum</i> Griseb.	R
	<i>Stenachaenium megapotamicum</i> Baker	R
	<i>Stenachaenium riedelii</i> Baker	F
	<i>Tagetes minuta</i> L.	R
	<i>Vernonia brevifolia</i> Less.	Ph
	<i>Vernonia flexuosa</i> Sims	Ph
	<i>Vernonia megapotamica</i> Spreng.	Ph
	<i>Vernonia nudiflora</i> Less.	Ph
	<i>Vernonia scorpioides</i> (Lam.) Pers.	Ph
	<i>Vernonia sellowii</i> Less.	Ph
	<i>Vernonia squarrosa</i> (D. Don) Less.	F
	<i>Viguiera immarginata</i> (DC.) Herter	Ph
	<i>Viguiera nudicaulis</i> (Pers.) Baker	Ph
Boraginaceae	<i>Cordia verbenacea</i> DC.	F
	<i>Moritzia ciliata</i> (Cham.) DC. ex Meisn.	F
Bromeliaceae	<i>Dyckia choristaminea</i> Mez	R
	<i>Dyckia leptostachya</i> Baker	R
Cactaceae	<i>Frailea gracillima</i> (Lem.) Britton & Rose	R
	<i>Parodia ottonis</i> (Lehm.) N. P. Taylor	Ph
Campanulaceae	<i>Wahlenbergia linarioides</i> (Lam.) A.DC.	F
Cistaceae	<i>Helianthemum brasiliensis</i> (Lam.) Pers.	Ph
Commelinaceae	<i>Commelina erecta</i> L.	Ph
Convolvulaceae	<i>Convolvulus crenatifolius</i> Ruiz & Pav.	F
	<i>Dichondra microcalyx</i> (Hallier f.) Fabris	R
	<i>Dichondra sericea</i> Sw.	Ph
	<i>Dichondra</i> sp.	F
	<i>Evolvulus sericeus</i> Sw.	Ph
Cyperaceae	<i>Bulbostylis capillaris</i> (L.) C. B. Clarke	Ph
	<i>Bulbostylis consanguinea</i> Nees	Ph
	<i>Bulbostylis juncoides</i> (Vahl) Kük. ex Osten	F
	<i>Bulbostylis sphaerocephala</i> (Boeck.) C. B. Clarke	Ph
	<i>Cyperus aggregatus</i> (Willd.) Endl.	Ph
	<i>Cyperus reflexus</i> Vahl var. <i>fraternus</i> (Kunth) Kuntze	F
	<i>Cyperus reflexus</i> Vahl var. <i>reflexus</i>	F
	<i>Rhynchospora gollmeri</i> Boeck.	F
	<i>Rhynchospora rugosa</i> (Vahl) Gale	Ph
	<i>Rhynchospora setigera</i> Griseb.	Ph
	<i>Scleria</i> cf. <i>balansae</i> Maury	F
Euphorbiaceae	<i>Croton glandulosus</i> L.	Ph
	<i>Croton</i> sp.	Ph
	<i>Euphorbia selloi</i> (Klotzsch & Garcke) Boiss.	Ph

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Family	Species	
Fabaceae	<i>Aeschynomene falcata</i> (Poir.) DC.	Ph
	<i>Aeschynomene virgata</i> Cav.	Ph
	<i>Centrosema virginianum</i> (L.) Benth.	Ph
	<i>Chamaecrista repens</i> (Vogel) H. S. Irwin & Barneby	Ph
	<i>Clitoria nana</i> Benth.	Ph
	<i>Collaea stenophylla</i> (Hook. & Arn.) Benth.	F
	<i>Crotalaria tweediana</i> Benth.	F
	<i>Desmanthus virgatus</i> (L.) Willd.	Ph
	<i>Desmodium incanum</i> DC.	Ph
	<i>Desmodium uncinatum</i> (Jacq.) DC.	R
	<i>Eriosema tacuareboense</i> Arechav.	Ph
	<i>Galactia marginalis</i> Benth.	Ph
	<i>Indigofera asperifolia</i> Bong. ex Benth.	F
	<i>Lathyrus nervosus</i> Boiss.	R
	<i>Lathyrus subulatus</i> Lam.	F
	<i>Lupinus bracteolaris</i> Desr.	R
	<i>Macropodium prostratum</i> (Benth.) Urb.	Ph
	<i>Mimosa dolens</i> Vell.	Ph
	<i>Poiretia tetraphylla</i> (Poir.) Burkart	F
	<i>Rhynchosia corylifolia</i> Mart. ex Benth.	Ph
	<i>Rhynchosia diversifolia</i> Micheli	Ph
	<i>Sellocharis paradoxa</i> Taub.	Ph
	<i>Stylosanthes leiocarpa</i> Vogel	Ph
<i>Stylosanthes montevidensis</i> Vogel	Ph	
<i>Zornia burkartii</i> Vanni	Ph	
Gesneriaceae	<i>Sinningia allagophylla</i> (Mart.) Wiehler	F
Hypericaceae	<i>Hypericum caprifoliatum</i> Cham. & Schltld.	R
Hypoxidaceae	<i>Hypoxis decumbens</i> L.	R
Iridaceae	<i>Cypella herbertii</i> (Lindl.) Herb.	Ph
	<i>Herbertia pulchella</i> Sweet	Ph
	<i>Sisyrinchium avenaceum</i> Klatt	F
	<i>Sisyrinchium megapotamicum</i> Malme	Ph
	<i>Sisyrinchium scariosum</i> I.M. Johnst.	Ph
Lamiaceae	<i>Cunila</i> sp.	F
	<i>Cunila galioides</i> Benth.	Ph
	<i>Glechon ciliata</i> Benth.	Ph
Liliaceae	<i>Clara ophiopogonoides</i> Kunth	F
Linaceae	<i>Cliococca selaginoides</i> (Lam.) C. M. Rogers & Mildner	Ph
Lythraceae	<i>Cuphea carthagenensis</i> (Jacq.) J. F. Macbr.	R
	<i>Cuphea glutinosa</i> Cham. & Schltld.	Ph
	<i>Cuphea thymoides</i> Cham. & Schltld.	F
Malpighiaceae	<i>Galphimia brasiliensis</i> (L.) A. Juss.	Ph
	<i>Janusia guaranitica</i> (A. St.-Hil.) A. Juss.	F
Malvaceae	<i>Abutilon malachroides</i> A. St.-Hil. & Naudin	F
	<i>Krapovickasia macrodon</i> (DC.) Fryxell	Ph
	<i>Krapovickasia urticifolia</i> (A. St.-Hil.) Fryxell	Ph
	<i>Pavonia hastata</i> Cav.	Ph
	<i>Sida rhombifolia</i> L.	R

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Family	Species	
	<i>Waltheria douradinha</i> A. St.-Hil.	Ph
	<i>Waltheria indica</i> L.	F
	<i>Wissadula glechomifolium</i> (A. St.-Hil.) R.E. Fr.	Ph
Melastomataceae	<i>Tibouchina gracilis</i> (Bonpl.) Cogn.	F
	<i>Tibouchina</i> sp.	Ph
Myrtaceae	<i>Campomanesia aurea</i> O. Berg	Ph
	<i>Eugenia dimorpha</i> O. Berg	R
	<i>Psidium formosum</i> (Barb. Rodr.) Burret	F
	<i>Psidium incanum</i> (O. Berg) Burret	F
	<i>Psidium luridum</i> (Spreng.) Burret	Ph
Orchidaceae	<i>Epidendrum fulgens</i> L.	F
	<i>Skeptrostachys congestiflora</i> (Cogn.) Garay	Ph
Oxalidaceae	<i>Oxalis conorrhiza</i> Jacq.	F
Plantaginaceae	<i>Plantago myosuroides</i> Lam.	F
	<i>Plantago paralias</i> Decne.	F
	<i>Plantago</i> sp.	Ph
Poaceae	<i>Agenium villosum</i> (Nees) Pilg.	F
	<i>Andropogon lateralis</i> Nees	Ph
	<i>Andropogon selloanus</i> (Hack.) Hack.	Ph
	<i>Aristida circinalis</i> Lindm.	Ph
	<i>Aristida filifolia</i> (Arechav.) Herter	Ph
	<i>Aristida flaccida</i> Trin. & Rupr.	Ph
	<i>Aristida jubata</i> (Arechav.) Herter	F
	<i>Aristida laevis</i> (Nees) Kunth	Ph
	<i>Aristida spegazzinii</i> Arechav.	Ph
	<i>Axonopus affinis</i> Chase	R
	<i>Axonopus siccus</i> (Nees) Kuhlm.	Ph
	<i>Axonopus suffultus</i> (Mikan ex Trin.) Parodi	Ph
	<i>Briza lamarckiana</i> Nees	Ph
	<i>Briza macrostachya</i> (J. Presl) Steud.	F
	<i>Briza minor</i> L.	R
	<i>Briza subaristata</i> Lam.	Ph
	<i>Briza uniolae</i> (Nees) Nees ex Steud.	Ph
	<i>Bromus auleticus</i> Trin. ex Nees	F
	<i>Bromus catharticus</i> Vahl	F
	<i>Calamagrostis alba</i> (J. Presl) Steud.	F
	<i>Calamagrostis viridiflavescens</i> (Poir.) Steud.	F
	<i>Cortaderia selloana</i> Asch. & Graebn.	R
	<i>Cynodon dactylon</i> (L.) Pers.	R
	<i>Danthonia cirrata</i> Hack. & Arechav.	Ph
	<i>Dichanthelium sabulorum</i> (Lam.) Gould & C. A. Clark	Ph
	<i>Digitaria insularis</i> (L.) Fedde	F
	<i>Eleusine tristachya</i> (Lam.) Lam.	F
	<i>Elyonurus candidus</i> (Trin.) Hack.	F
	<i>Elyonurus</i> sp.	Ph
	<i>Eragrostis lugens</i> Nees	Ph
	<i>Eragrostis neesii</i> Trin.	F
	<i>Eragrostis perennis</i> Döll	F

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Family	Species	
	<i>Eragrostis plana</i> Nees	F
	<i>Eragrostis polytricha</i> Nees	Ph
	<i>Eragrostis</i> sp.	Ph
	<i>Eustachys distichophylla</i> (Lag.) Nees	F
	<i>Eustachys uliginosa</i> (Hack.) Herter	F
	<i>Leptocoryphium lanatum</i> (Kunth) Nees	Ph
	<i>Melica brasiliiana</i> Ard.	F
	<i>Melinis repens</i> (Willd.) Zizka	F
	<i>Panicum bergii</i> Arechav.	F
	<i>Panicum chaseae</i> Roseng., B. R. Arrill. & Izag.	Ph
	<i>Panicum olyroides</i> Kunth	F
	<i>Paspalum dilatatum</i> Poir.	F
	<i>Paspalum mandiocanum</i> Trin.	F
	<i>Paspalum notatum</i> Flügge	F
	<i>Paspalum paniculatum</i> L.	F
	<i>Paspalum paucifolium</i> Swallen	F
	<i>Paspalum plicatulum</i> Michx.	Ph
	<i>Paspalum quarinii</i> Morrone & Zuloaga	F
	<i>Paspalum urvillei</i> Steud.	F
	<i>Phalaris angusta</i> Nees ex Trin.	F
	<i>Piptochaetium montevidense</i> (Spreng.) Parodi	Ph
	<i>Piptochaetium ruprechtianum</i> E. Desv.	Ph
	<i>Piptochaetium stipoides</i> (Trin. & Rupr.) Hack.	F
	<i>Saccharum</i> cf. <i>asperum</i> (Nees) Steud.	F
	<i>Schizachyrium imberbe</i> (Hack.) A. Camus	Ph
	<i>Schizachyrium microstachyum</i> (Ham.) Roseng., B. R. Arrill. & Izag.	Ph
	<i>Schizachyrium spicatum</i> (Spreng.) Herter	Ph
	<i>Schizachyrium tenerum</i> Nees	Ph
	<i>Setaria parviflora</i> (Poir.) Kerguélen	Ph
	<i>Setaria vaginata</i> Spreng.	Ph
	<i>Sorghastrum albescens</i> (E. Fourn.) Beetle	F
	<i>Sporobolus camporum</i> Swallen	Ph
	<i>Sporobolus indicus</i> (L.) R. Br.	R
	<i>Steinchisma hians</i> (Elliott) Nash	F
	<i>Stipa</i> cf. <i>filiculmis</i> Del.	F
	<i>Stipa filifolia</i> Nees	Ph
	<i>Stipa megapotamia</i> Spreng. ex Trin.	F
	<i>Stipa melanosperma</i> J. Presl var. <i>melanosperma</i>	F
	<i>Stipa nutans</i> Hack.	F
	<i>Stipa</i> sp.	Ph
	<i>Trachypogon montufarii</i> (Kunth) Nees	Ph
	<i>Urochloa decumbens</i> (Stapf) R. D. Webster	F
Polygalaceae	<i>Monnina cardiocarpa</i> A. St.-Hil.	Ph
	<i>Polygala extraaxillaris</i> Chodat	Ph
Rubiaceae	<i>Diodella apiculata</i> (Willd. ex Roem. & Schult.) Delprete	Ph
	<i>Galianthe fastigiata</i> Griseb.	Ph
	<i>Galium hirtum</i> Lam.	Ph
	<i>Galium richardianum</i> (Gillies ex Hook. & Arn.) Endl. ex Walp.	Ph
	<i>Richardia grandiflora</i> (Cham. & Schltdl.) Steud.	Ph

continue

continuation

Family	Species	
	<i>Richardia humistrata</i> (Cham. & Schltld.) Steud.	Ph
	<i>Spermacoce capitata</i> Ruiz & Pav.	Ph
	<i>Spermacoce verticillata</i> L.	Ph
Scrophulariaceae	<i>Angelonia integerrima</i> Spreng.	Ph
	<i>Buchnera integrifolia</i> Larrañaga	Ph
	<i>Buddleja thyrsoides</i> Lam.	R
	<i>Gerardia communis</i> Cham. & Schltld.	Ph
	<i>Scoparia ericacea</i> Cham. & Schltld.	Ph
Smilacaceae	<i>Smilax</i> cf. <i>campestris</i> Griseb.	F
Solanaceae	<i>Calibrachoa ovalifolia</i> (Miers) Stehmann & Semir	F
	<i>Nicotiana alata</i> Link & Otto	F
	<i>Petunia integrifolia</i> (Hook.) Schinz & Thell.	Ph
	<i>Solanum sisymbriifolium</i> Lam.	F
Symplocaceae	<i>Symplocos uniflora</i> (Pohl) Benth.	F
Turneraceae	<i>Piriqueta selloi</i> Urb.	Ph
Urticaceae	<i>Urtica circularis</i> (Hicken) Sorarú	R
Verbenaceae	<i>Glandularia incisa</i> (Hook.) Tronc.	F
	<i>Glandularia platensis</i> (Spreng.) Schnack & Covas	Ph
	<i>Glandularia subincana</i> Tronc.	Ph
	<i>Lantana montevidensis</i> (Spreng.) Briq.	Ph
	<i>Stachytarpheta cayennensis</i> (Rich.) Vahl	F
	<i>Verbena lindmani</i> Briq.	R
	<i>Verbena pseudojuncea</i> Gay	Ph
Undetermined	Dicot. 1	Ph
	Dicot. 2	Ph

The structural survey carried out has found 161 species, distributed in 30 families. The most diverse families in the structural survey were Asteraceae (45 species), Poaceae (33 species) and Fabaceae (17 species). However, considering the accumulated IV per family, Poaceae had the highest value (33.21%), and five of the top-ten IV-ordered species belong to this family (table 2). The accumulated IV for Asteraceae and Fabaceae was respectively 28.57% and 7.22%. The low value for Fabaceae characterized it as a diverse family with low contribution on frequency or cover in the study site. The species with the highest IV were particularly grasses, the prostrate Rubiaceae *Richardia grandiflora* and some species of Asteraceae (table 2). The diversity and evenness indexes obtained for the community were $H' = 4.51 \text{ nats ind}^{-1}$ and $E = 0.86$.

Results from the similarity analysis among Morro do Osso, Morro da Polícia and Morro Santana indicated similar grassland community composition in the three granitic hills analyzed (table 3). Although the values themselves are not remarkably high (average of 33%

similarity), they are very close to each other (0.03 standard deviation). There are 57 common species among the three surveys. Many of these species are dominant grasses like *Andropogon lateralis*, *Aristida flaccida* and *Trachypogon montufarii*, suggesting similar community structure and landscape on the different study sites. Moreover, *Stipa filifolia*, an endemic grass with distribution restricted to Brazilian granitic hills, was also found in the three surveys. Besides grasses, many forb and sub-shrub species of *Baccharis*, *Chaptalia* and *Eryngium* are common among the areas, as well as some rare species like the cactus *Parodia ottonis*.

Mean species number found per plot (1 m²) was 27.10 (standard deviation = 4.96). Plots with lowest and highest species richness had respectively 15 and 35 species. Mean vegetation height on the sampled area was 28.87 cm (standard deviation 8.62) and open soil and litter were present in all sampling units, the first with 4.07% (standard deviation = 13.11) of mean absolute cover per plot unit and the latter with 0.55% (standard deviation = 0.86).

Table 2. Grassland species sampled in the structural survey, decreasingly ordered by IV. Only the top-20 IV-valued species are shown. (IV = importance value; N = number of sampling units in which the species was accounted; RF = relative frequency; RC = relative cover).

Family	Species	IV (%)	N	RF (%)	RC (%)
Poaceae	<i>Aristida flaccida</i>	3.22	27	2.55	3.88
Poaceae	<i>Andropogon lateralis</i>	2.89	27	2.55	3.23
Rubiaceae	<i>Richardia grandiflora</i>	2.84	31	2.93	2.75
Poaceae	<i>Axonopus suffultus</i>	2.68	26	2.46	2.91
Asteraceae	<i>Vernonia nudiflora</i>	2.36	26	2.46	2.26
Asteraceae	<i>Eupatorium laetevirens</i>	2.25	22	2.08	2.42
Poaceae	<i>Schizachyrium tenerum</i>	2.10	18	1.70	2.50
Asteraceae	<i>Vernonia flexuosa</i>	2.10	23	2.18	2.02
Lamiaceae	<i>Glechon ciliata</i>	2.04	21	1.99	2.10
Poaceae	<i>Panicum chaseae</i>	2.03	19	1.80	2.26
Poaceae	<i>Paspalum plicatulum</i>	1.87	20	1.89	1.86
Apiaceae	<i>Eryngium pritis</i>	1.78	12	1.14	2.42
Asteraceae	<i>Aspilia montevidensis</i>	1.75	20	1.89	1.62
Fabaceae	<i>Galactia marginalis</i>	1.75	20	1.89	1.62
Cyperaceae	<i>Rhynchospora rugosa</i>	1.75	20	1.89	1.62
Asteraceae	<i>Calea uniflora</i> var. <i>uniflora</i>	1.75	19	1.80	1.70
Convolvulaceae	<i>Evolvulus sericeus</i>	1.67	19	1.80	1.53
Poaceae	<i>Trachypogon montufarii</i>	1.63	14	1.32	1.94
Euphorbiaceae	<i>Croton</i> sp. 1	1.62	18	1.70	1.53
Poaceae	<i>Axonopus siccus</i>	1.60	15	1.42	1.78
Total		41.70	–	39.45	43.94

Clustering and ordination analysis – Cluster analysis denoted the existence of two sharp groups among the sampling units (figure 2). Cluster groups coincided with the two major grassland-covered areas that were distinguished by a natural environmental bias caused by fire disturbance due to recent burn on the eastern slope (axis 1 variation; figure 3). The majority of the east plots formed a compact

distribution of points on figure 3, whereas the west plots shaped a more scattered pattern (axis 2 variation). Because of the great coincidence among cluster groups and the set of plots located in the same slope, further analyses were done according to plot location.

Species plotted on figure 3 are related to differences in floristic and structural composition between the two

Table 3. Jaccard similarity indexes between our study (Morro do Osso) and two others granitic hills from Porto Alegre region. Data for Morro Santana and Morro da Policia obtained respectively from Overbeck *et al.* (2006) and Boldrini *et al.* (1998). “Sampled Species” values represent taxa identified to the specific level.

	Morro do Osso	Morro Santana	Morro da Policia
Morro do Osso	1		
Morro Santana	0.35897	1	
Morro da Policia	0.33884	0.28736	1
Sampled species	152e	165	171
Exclusive species	43	64	71
Sampling method	39 × 1 m ² quadrats	48 × 0.75 m ² quadrats	2829 points

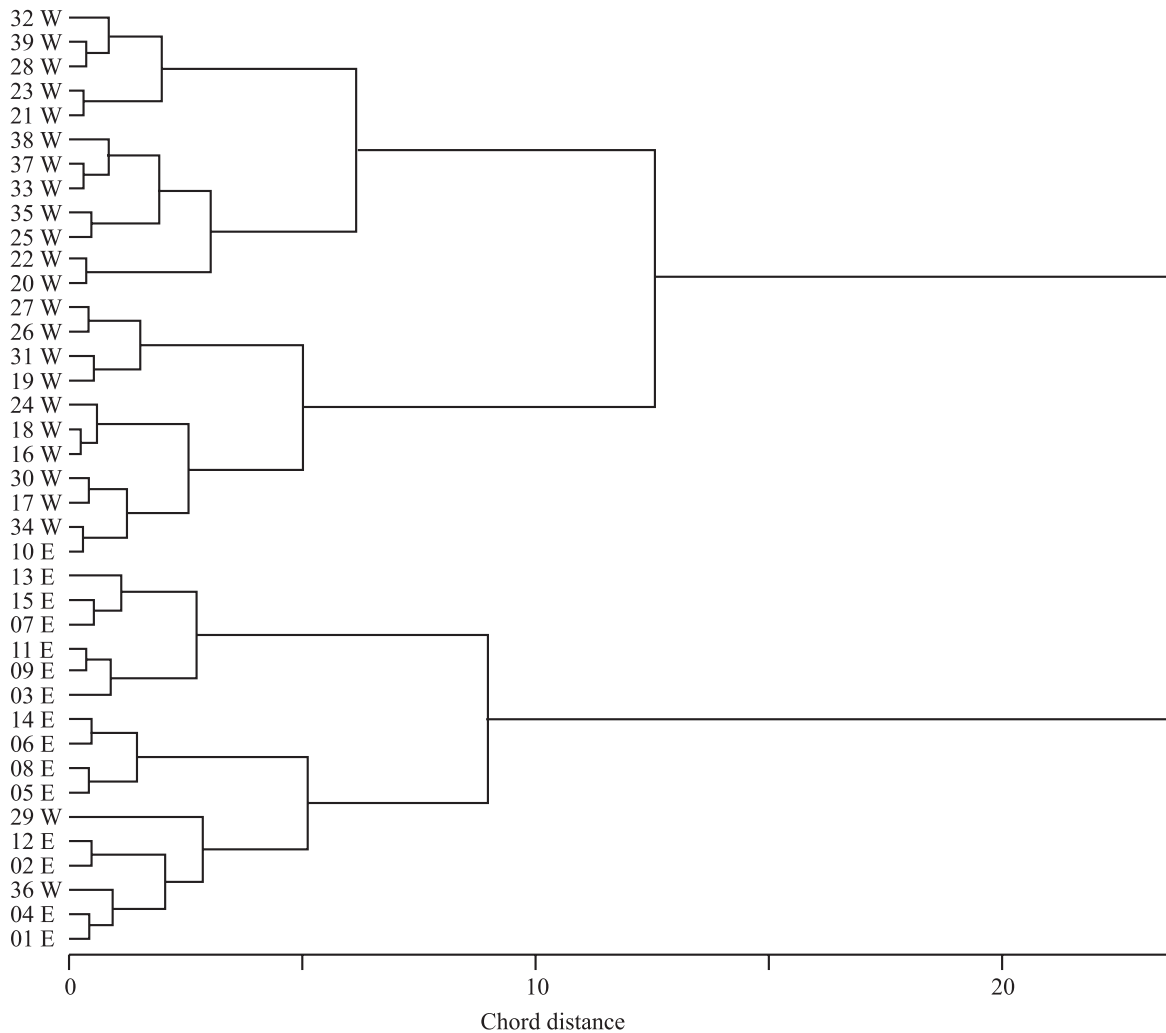


Figure 2. Cluster analysis diagram obtained for the 39 sampling plots described by cover value of species, litter and open soil variables. Only two groups are statistically supported by bootstrap analysis ($P > 0.1$). Incremental sum of squares (horizontal axis) was used as the clustering criterion and chord distance as the dissimilarity measure (E1-E16 = plots in the east grassland area; W17-W39 = plots in the west grassland area).

major grassland areas. These differences can be better observed by species performances (figure 4) in each plot group. East plots presented a total of 111 species, being 32 exclusive of this area, whereas for the west plots 129 species were surveyed in total, being 50 exclusive species. Even with these given floristic peculiarities, there are no statistical differences in richness between the areas (east = 27.75 species per plot; west = 26.65).

Grass species such as *Schizachyrium tenerum*, *Trachypogon montufarii*, *Axonopus siccus* and *Aristida filifolia* had their performance decreased in the east area (compare table 2 and figure 4), whereas *Schizachyrium microstachyum*, *Eragrostis polytricha* (grasses), *Eryngium horridum*, *Pterocaulon angustifolium* and *Vernonia flexuosa* (forbs) increased their performance. *Andropogon*

lateralis, *Aristida flaccida*, *Axonopus suffultus* (grasses), *Vernonia nudiflora* (sub-shrub), *Eupatorium laetevirens* and *Aspilia montevidensis* (forbs) were dominant species that maintained their relative cover proportion in both areas. The first two grasses, however, had proportionally more cover in east plots (figure 4).

There was no difference considering vegetation height in both grassland areas (east plots: 26.68 cm; west plots: 28.69 cm). In addition, open soil mean cover had no statistical difference between the areas, albeit the slight difference between east and west plots (mean of 1.62 and 1.26, respectively) and the visual difference observed in the field. However, litter presented statistical difference between the areas (mean of 1.06 and 1.39 for east and west plots, respectively; $P = 0.03$).

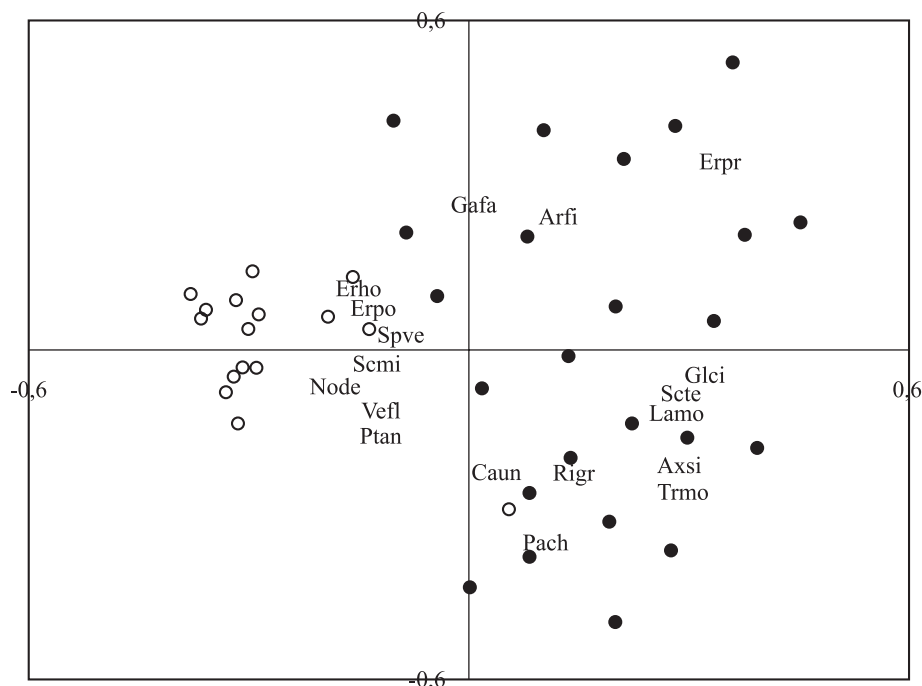


Figure 3. Ordination diagram by PCoA of the 39 sampling plots on the cover value of species, litter and open soil. Plot labels are identified according to main aspect (east; west) of grassland area. Chord distance was used as dissimilarity measure. The ordination axes sum 20% of total variation. Variables correlated more than 0.45 with the ordination axes were plotted in the diagram (Arfi = *Aristida filifolia*; Axsi = *Axonopus siccus*; Caun = *Calea uniflora*; Erpo = *Eragrostis polytricha*; Erho = *Eryngium horridum*; Erpr = *Eryngium pristic*; Gafa = *Galianthe fastigiata*; Glei = *Glechon ciliata*; Lamo = *Lantana montevidensis*; Node = *Noticastrum decumbens*; Pach = *Panicum chaseae*; Ptan = *Pterocaulon angustifolium*; Rigr = *Richardia grandiflora*; Scmi = *Schizachyrium microstachyum*; Scte = *Schizachyrium tenerum*; Spve = *Spermacoce verticillata*; Trmo = *Trachypogon montufarii*; Vefl = *Vernonia flexuosa*; empty circles = east; full circles = west).

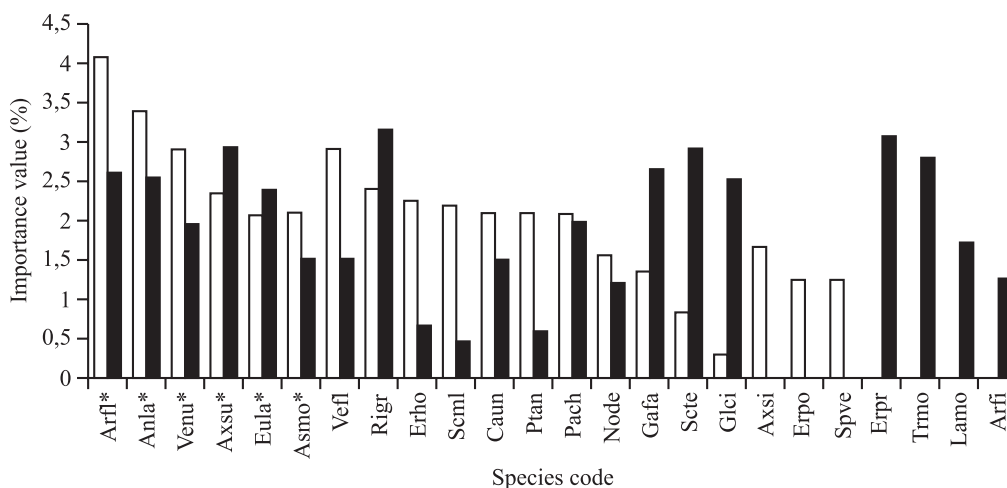


Figure 4. Importance value of typical species of plots representing east (recent burned site) and west grassland areas (species shown in figure 3) and the first five species (*) which are well represented in both areas (Arfl = *Aristida flaccida*; Anla = *Andropogon lateralis*; Venu = *Vernonia nudiflora*; Axsu = *Axonopus suffultus*; Eula = *Eupatorium laetevirens*; Asmo = *Aspilia montevidensis*; Vefl = *Vernonia flexuosa*; Erho = *Eryngium horridum*; Scmi = *Schizachyrium microstachyum*; Pach = *Panicum chaseae*; Rigr = *Richardia grandiflora*; Caun = *Calea uniflora*; Ptan = *Pterocaulon angustifolium*; Gafa = *Galianthe fastigiata*; Scte = *Schizachyrium tenerum*; Glei = *Glechon ciliata*; Axsi = *Axonopus siccus*; Erpo = *Eragrostis polytricha*; Spve = *Spermacoce verticillata*; Node = *Noticastrum decumbens*; Erpr = *Eryngium pristic*; Trmo = *Trachypogon montufarii*; Lamo = *Lantana montevidensis*; Arfi = *Aristida filifolia*; empty bars = east; full bars = west).

Discussion

Diversity in grassland formations presented in studies carried out in Rio Grande do Sul State are quite variable, depending on the method and the region surveyed. In a flat region in the middle of the State, different efforts have found less species when compared to the present study. Focht & Pillar (2003) have found 148 grassland species with vegetation patterns related to relief position and other factors, such as soil moisture, on a study located approximately 35 km west from ours. On the other hand, granitic grassland formations tend to present high species richness, as showed by Boldrini *et al.* (1998) in a study based on point method on Morro da Polícia, where 189 species were surveyed. In another similar hill (Morro Santana), 165 species were recorded by plot survey in a grassland area that burns in intervals of three to five years (Overbeck *et al.* 2006).

The survey accomplished at Morro da Polícia by Boldrini *et al.* (1998) showed more exclusive species (71) and more overall species (171 fully identified species), probably due to a more exhausting sampling process (18 transects with 2829 points). The survey at Morro Santana (Overbeck *et al.* 2006) was carried out in 48 plots of 0.75 square meters, a sampling process similar to the one we have implemented at Morro do Osso, and resulted in 165 species (64 exclusive) identified to the specific level. Our survey, comprising the smaller total area among the three hills, showed 43 exclusive species. We consider that, among the exclusive species found in our survey, very few are actually exclusive, and their apparent exclusiveness could be simply related to different collection efforts, as the three compared areas are spatially and floristically close to each other.

Considering a broad study of Porto Alegre's hills region, 522 grassland species were pointed out in a survey on ten granitic hills, not including Morro do Osso (Aguiar *et al.* 1986). The occurrence of many concomitant species among the lists of these studies and ours indicates a rather continuous flora along the granitic hills. In addition, despite the species richness differences estimated by the similarity indexes for Morro Santana, Morro da Polícia and the present study, there is a very similar species composition among these areas, with an equivalent species-family distribution and many common dominant species. This point may corroborate Rambo's idea that these granitic hills were refuge areas during ocean transgression times (Rambo 1954), so that the extant grassland vegetation configures a remnant of different climate conditions (Behling 2002, Bredenkamp *et al.* 2002).

Presence of endemic and rare species in the study site is considerably high. Rambo (1954), in a classic paper concerning the southern Brazilian flora, emphasized plant endemism to be outstanding in Porto Alegre area, mostly comprising herbaceous species and genera. Although the endemic species list this author presents have greatly diminished at present due to growing collection effort and subsequent increase in the distribution area of many southern Brazilian species, some species deserve attention concerning their particular occurrence on granitic hills, such as *Moritzia ciliata* and *Stipa filifolia*. Other species, such as *Dyckia choristaminea*, have their distribution area limited to Porto Alegre region, whereas *Baccharis riograndensis*, *Sellocharis paradoxa* and *Eugenia dimorpha* have a broader distribution in the State of Rio Grande do Sul. Besides these, *Heterothalamus psiadioides* and *Dyckia leptostachya* have their distribution limited to the southernmost Brazilian state, whereas *Eryngium horridum* and *E. ciliatum* have the state as their northern distributional limit, also occurring on northern Uruguay and Argentina.

Eleven species presented in the floristic list of this study are categorized as extinction-threatened by official governmental list (Sema 2003). Among them, eight are labeled as vulnerable and three as endangered. None of the endangered species (*Dyckia choristaminea*, *Frailea gracillima* and *Schlechtendalia luzulifolia*) and only four of the vulnerable ones (*Gomphrena graminea*, *Mandevilla coccinea*, *Parodia ottonis* and *Waltheria douradinha*) were accounted for in the structural survey, suggesting scarcity in the area. The other vulnerable species are *Gochnatia cordata*, *G. orbiculata*, *Moritzia ciliata* and *Stenachaenium macrocephalum*. Only one individual of *M. ciliata* was sampled, and very few were seen in subsequent expeditions. This evidence of possible population decrease is concerning, as this species is considered, farther than threatened, rare in Brazilian granitic grasslands. The scarcity of *S. luzulifolia* seen on the field is also concerning, as the distribution of this species has Porto Alegre city region as its northern limit (Mondin & Baptista 1996).

The absolute values of diversity and evenness indexes obtained for the presented community were supposedly higher (4.51 nats ind⁻¹ and 0.86, respectively) than for Morro da Polícia (4.01 nats ind⁻¹ and 0.76; Boldrini *et al.* 1998). Considering that species richness was lower in our area (161 for our study and 189 for Boldrini *et al.* 1998), the evenness in species performance at Morro do Osso survey should explain the higher Shannon diversity. Another important point is the recent burn undergone in

part of the sampling area, which may have contributed to the high diversity index obtained for the community. Overbeck *et al.* (2005, 2006), working in Morro Santana, have shown that fire disturbance clearly leads to a short term increase in species richness and diversity (Denslow 1985, Harrison *et al.* 2003), but does not significantly affect overall community species composition. The maintenance of composition, alongside the short-term diversity peak and the regularly break of high competitors tussock species dominance, could explain the diversity and evenness values for the surveyed community, since fire in local area is a long term disturbance event. Forb species may be outcompeted by tussock grass dominance (Lattera *et al.* 2003, Overbeck *et al.* 2005). This may be first perceived only at above-ground vegetation, since many species can survive underground as bud-bank or reserve organs waiting for better opportunities, but forb submission may occur if disturbance intervals become long enough to suppress those organs (Rodríguez *et al.* 2003).

Although cluster and ordination analysis showed a distinct pattern in the studied grassland community, patterns among each clustering group were not uniform. East plots showed more similarity among themselves both in cluster and ordination analysis, suggesting a structural and floristic pattern, probably due to the survival of common fire-resilient species (Müller *et al.* 2007) and similar post-fire species recruitment (Pillar & Quadros 1997, Uys *et al.* 2004). On the other hand, west plots do not share a common trait like recent fire and, moreover, are distributed over a larger and more variable environment. Therefore, the pattern drawn by these plots along the second axis on the ordination diagram showed less spatial cohesion, probably illustrating other environmental differences among sampling plots.

Most floristic changes in disturbed areas are a consequence of covering area reduction of dominant species that play a key role on new species recruitment or changes in species performance and consequently in any community diversity modification (Connell 1978, Huston 1979, Olf & Ritchie 1998). Although the fire-driven dominance reduction that occurred in part of our study area enabled the sprouting of many exclusive species (32) in the east plots, west grassland plots had much more exclusive species (50), possibly due to the removal of fire-vulnerable species in the first and to the higher number of plot units on the western grassland area (23 west plots; 16 east plots). *Eryngium pristis*, *Lantana montevidensis*, *Trachypogon montufarii* and *Stipa filifolia* are examples of high performance species present exclusively on the west plot units. Overbeck

et al. (2005) pointed out that areas with an intermediate time without fire were the most rich of a comparison considering recently (one year) and long time (more than five years) burned places. As it is, the east overall grassland area might reach its species richness peak within few years after this study and our west group plots may be at an intermediate time without influence of fire, presenting more concentration of species per area.

Although fire had no effect on mean vegetation height of sampling units, it did affect floristic and structural composition between the two major areas analyzed. Open soil cover did not differ as well, but litter cover was higher in west plots, clearly as a consequence of burn effect on east grassland area.

Species composition and performance depicted differences between east and west areas. *Eryngium horridum*, *Eragrostis polytricha*, *Spermacoce verticillata*, *Noticastrum decumbens*, *Schizachyrium microstachyum* and *Pterocaulon angustifolium*, which were better represented on our recently burned area, are typically present in early successional stages and altered areas (Eggers & Porto 1994, Heringer & Jacques 2002). Quadros & Pillar (2001), working on a south Brazilian grassland, have also focused on floristic changes after fire and grazing disturbance. They have discussed that vegetation showed greater resilience to fire treatments, albeit remarkably differences were seen in initial observations.

The similarity we have found among the granitic hills, as well as the presence of rare and endemic species among them, denotes a historical continuous flora (at least on grasslands communities) along these formations, corroborating the shared natural history of the area. Besides that, the great diversity observed reflects the ecological importance of this site, as an example of great plant diversity of grassland formations. Nevertheless, further research including more granitic hills and a biogeographical approach are considered to be necessary in order to understand the Porto Alegre's granitic hill chain flora as a whole.

Fire disturbance, with the basic effect of diminishing dominance of some species, thus allowing the rising of others, was partly responsible for the differences in species composition although differences in overall diversity were not recordable. The accidental fire held at the study site showed more effect on performance of species, which is temporary, than on their presence or exclusion of the area. Considering the whole area, this contributes to the high diversity level found for the whole community.

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