

Neuropathological and neurochemical abnormalities in bipolar disorder

Benício Noronha Frey,^{a,b} Manoela M Rodrigues da Fonseca,^{a,c}
Rodrigo Machado-Vieira,^{a,d} Jair C Soarese and Flávio Kapczinski^{a,c,f}

Original version accepted in Portuguese

^aLaboratory of Experimental Psychiatry of the Clinical Hospital of Porto Alegre (HCPA)

^bDepartment of Biochemistry – Institute of Basic Health Sciences – Federal University of Rio Grande do Sul (UFRGS)

^cPsychiatric Service of the HCPA

^dMood Disorders Program – Foundation Federal School of Medical Sciences of Porto Alegre – Maternal-Child Hospital Presidente Vargas

^eDivision of Mood and Anxiety Disorders, Department of Psychiatry, University of Texas Health Science Center at San Antonio, TX, USA

^fDepartment of Legal Psychiatry and Medicine of UFRGS

Abstract

Objectives: Postmortem, pharmacological, neuroimaging, and animal model studies have demonstrated a possible association of intracellular signaling mechanisms in the pathophysiology of bipolar disorder. The objective of this paper is to review the findings in neuropathology and cellular biochemistry.

Methods: We performed a MEDLINE research, between 1980-2003, using bipolar disorder, signaling, second messengers, and postmortem as keywords, and cross-references.

Results: Neuropathological studies reported a decrease in neuronal and glial cells, mainly in the prefrontal cortex of bipolar patients. Neurochemical studies reported dysfunction in cAMP, phosphoinositide, Wnt/GSK-3 β , and intracellular Ca⁺⁺ pathways in these patients.

Conclusions: The neuropathological and neurochemical abnormalities demonstrated in BD may be related to the pathophysiology of this disorder and the effects of mood stabilizers. However, further studies are needed to clarify the role of the intracellular signaling cascade in the pathogenesis of this disorder.

Keywords: Bipolar disorder; Second messenger systems; Brain chemistry

Introduction

Bipolar disorder (BD) has been known for decades as a chronic mental disorder, with high relapse rates, most of times incapacitating, supposedly having a neurobiological substrate. Although the understanding of neurobiology has expanded in the last years, little is known about the real pathophysiological mechanisms of BD.¹ Recent genetic investigations have reported results which, although conflicting, seem to demonstrate some association, at least in a percentage of ill subjects. Neuroimaging studies showed a series of structural and functional alterations in determined brain regions of bipolar subjects, such as prefrontal and temporal cortices, cerebellum, basal ganglia and limbic system;³ however, these studies do not allow reaching a more specific cell substrate in these regions. Besides, neuropathological studies (postmortem) showed decrease of glia and neuronal density and plasticity, as well as alterations in intracellular neurochemistry.⁴⁻⁵

One of the models that has been applied to BD is kindling,⁶ taken from the model of epilepsy, in which the repetition of crises would cause a process of neuronal sensitization, leading to a progressive

threshold decrease, with the increase in the recurrence of epileptic crises (manic). Animal model studies suggest that this process may involve a series of alterations in the genic expression and second messengers.⁶ In fact, pharmacological studies have been consistent with these findings, demonstrating the action of antidepressants and mood stabilizers in several intracellular mechanisms which involve the regulation of genic expression and cellular plasticity.⁷ This study aims to review the findings in neuropathology and cell biochemistry which seem to be involved in the pathophysiology of BD. A search on MEDLINE, between 1980 and 2003, was performed, using the keywords: bipolar disorder, signaling, second messengers and postmortem. Cross-references of the selected articles were also used.

Neuropathology of BD

Structural neuroimaging studies demonstrated significant alterations in brain volume, suggesting neuronal atrophy and/or loss, in at least a percentage of subjects with BD. Several studies reported a significant decrease in the gray matter in prefrontal⁸⁻⁹ and

temporal¹⁰⁻¹¹ cortices, besides increase in lateral ventriculi.¹² Independent studies which studied specific regions of the temporal cortex showed also decrease.¹³⁻¹⁴ and increase¹⁵⁻¹⁶ in amygdale volume of bipolar subjects. A consistent finding on BD studies is a higher frequency of hyperintensities of subcortical white matter.¹⁷ These diffuse lesions may signify interruption in the circuits involved in mood regulation. Although less consistently, alterations in basal ganglia³ and cerebellum¹⁸ were also noted among bipolar patients. As a whole, these findings point to a possible dysfunction in the cortical-limbic circuit as an autonomic substrate of BD. Functional neuroimaging studies provide additional evidence of alterations in the metabolism of glucose and decrease in regional blood flow and cellular energetic phosphates in cortical and subcortical regions in BD.¹⁹ However, the resolution of the current neuroimaging techniques is limited to millimeters, being, therefore, fundamental postmortem studies which allow a direct study with the cellular and molecular resolution.

In one of the neuropathological studies with a larger sample of bipolar subjects (n=18), Öngür et al²⁰ reported a significant decrease (41.2%) in the density of glial cells in the subgenual prefrontal cortex, Brodmann's 24th area, among bipolar and unipolar subjects with positive family history for mood disorders. However, this study did not assess if this decrease occurred specifically for astrocytes, oligodendrocytes or microglia. Recently, Rajkowska et al²¹ have not found significant differences in the density and size of glial cells in the dorsolateral prefrontal cortex (DLPFC – Brodmann's 9th area) of bipolar subjects. However, a more detailed laminar analysis showed a significant decrease (19%) of the glial density in the sublayer IIIc of DLPFC. This study found also a decrease in the number of average-size cells in glial layers III and V, accompanied by an increase in the number of cells with very large nuclei.²¹ Two independent studies provided additional data consistent with glial alterations in mood disorder. Cotter et al²² reported a decrease in the glial density on layer VI of Brodmann's area 24 among unipolar and schizophrenic subjects but not among bipolar ones (most of them were using mood stabilizers, which seem to have a neurotrophic/neuroprotector effect). Using the same cohort of patients, Uranova et al²³ investigated the area 10 of the dorsal prefrontal cortex and showed a significant decrease in the glial density on layer VI of bipolar and schizophrenic subjects, and only among unipolar ones with positive family history for 'severe mental disorder'. As a whole these findings point to the hypothesis that at least one subgroup of bipolar and unipolar subjects, mainly those with positive family history, show some deficit in the glial density in multiple sites of the prefrontal cortex, what may affect their connection with other brain regions.

In the limbic system, Benes et al²⁴ reported a significant decrease of non-pyramidal neurons in the hippocampal region CA2 among bipolar and schizophrenic subjects. More recently, the same group has replicated the same finding, although in layer II of the anterior cortex of the cingulate.²⁵ These findings suggest a possible association with the decrease in the GABAergic inhibition (non-pyramidal neurons) in the pathophysiology of BD.⁴ In the brainstem it was demonstrated a bilateral increase in the number of pigmented neurons on the locus ceruleus of bipolar subjects compared to unipolar ones.²⁶ These neurons are one of the main sources of noradrenaline (NA) in the CNS.³⁴ In this sense, a postmortem study

by Young et al²⁷ reported an increase in NA turnover in the cortex of bipolar subjects.

Glial cells regulate the energetic homeostasis of the CNS, by means of glucose reuptake and phosphorylation during neuronal activity. Besides, they participate in the development, maintenance and remodeling of the synaptic connections, through the release of trophic factors and the regulation of synaptic glutamate concentration.²⁸ Thus, the findings of reduction in the glial density may result in the decrease in the number of functional synapses in BD. Consistently with this hypothesis of synaptic dysfunction, two post-mortem studies which assessed the hippocampal region evidenced decrease in the mRNA expression of synaptic proteins²⁹ and in apical dendritic spines of pyramidal cells³⁰ in the subicular subregion of BD subjects.

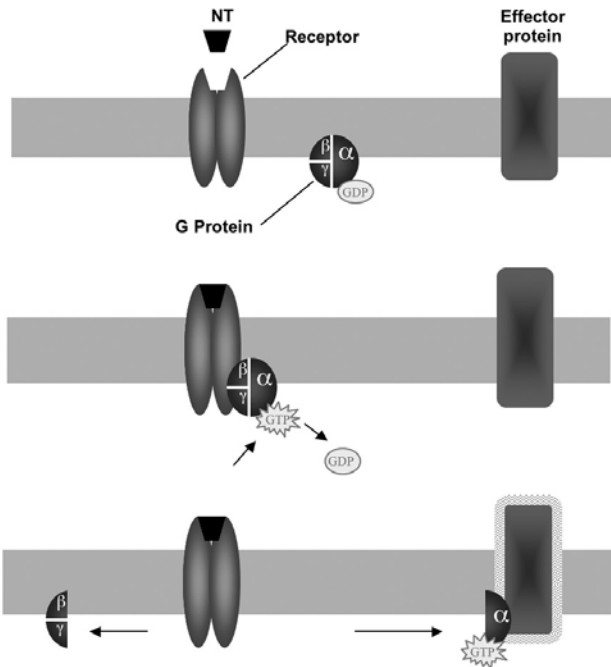
Therefore, the advance of molecular biochemistry applied to post-mortem studies point to a dysfunction of the complex intracellular mechanisms, which involve second messengers systems, regulation of the genic expression and synthesis of trophic factors (neuroplasticity), as associated with the pathophysiology of BD.^{1,6-7}

Intracellular signaling systems

G Proteins

The mechanism which involves the transmission of the information from the synapsis up to the cell nucleus is mediated by an intermediate process called second messengers, such as the cyclic adenosine monophosphate (cAMP) and phosphatidylinositol (PIP₂) pathways. This process involves three stages: 1) the neurotransmitter binds to the membrane receptor; 2) the activation of proteins which use guanosine triphosphate (GTP) as a cofactor, called G proteins; and 3) the activation of effector systems (through second messengers – see figure 1). G proteins have three subunits (α, β and γ), which are closely bound to the internal face of the plasmatic membrane, being activated by the binding of the neurotransmitter to its specific receptor. Multiple CNS receptor systems are modulated by G proteins, including noradrenergic, serotonergic, dopaminergic, cholinergic and histaminergic receptors, among others. G proteins may have either stimulatory or inhibitory effect on effector proteins, being therefore classified as G_s (stimulatory protein) and G_i (inhibitory). In this way, receptor-activated G proteins modulate ion flows, through the regulation of the activity of ionic channels, and control the activity of several effector enzymes. The interest in the study of G proteins in BD has arisen from the findings about the regulatory action of lithium on several G protein subtypes in animal models.³¹⁻³²

Young et al³³⁻³⁴ were the first to report an increase in G_s levels in frontal, temporal and occipital cortices of BD subjects. This finding was replicated in one study which demonstrated an increase in agonist-stimulated G_s activity.³⁵ However, these results do not discard the possibility of being effects of pharmacological treatments or of samples with small sizes. In one study, which investigated a larger sample, Dowlatsahi et al³⁶ did not find differences in G_s levels in bipolar subjects regarding the control group. Nevertheless, they evidenced a significant increase in G_s among patients who were not using lithium. As a result, the use of the drug might have been responsible for the failure to detect the difference between the total of patients and the control group.



NT = Neurotransmitter

Figure 1 - G-protein intracellular signaling pathway

Peripheral blood studies have also confirmed these findings and widened the understanding of the relationship between the functioning of G proteins and mood states. Schreiber et al³⁷ were the first to evidence an increase in the activity of G protein in mononuclear leucocytes of manic patients. Other two studies observed increase in mononuclear G_s levels of non-medicated depressed bipolar patients,³⁸⁻³⁹ whereas other study found increased levels in manic and decreased levels in depressive state.⁴⁰ Besides, this increase in G_s expression was also demonstrated in platelets,⁴¹⁻⁴² but not in lymphoblasts.⁴³ These studies suggest that mood state and cell type may influence in the finding of increased G_s in the peripheral blood of bipolar subjects. Taken as a whole, these findings suggest a possible association of the functioning of G proteins in the pathophysiology of DB. However, it has still not been determined if BD is associated with direct dysfunction in the activity of G proteins or these findings represent a secondary manifestation of a dysfunction in other pathways.

Cyclic adenosine monophosphate (cAMP) pathway

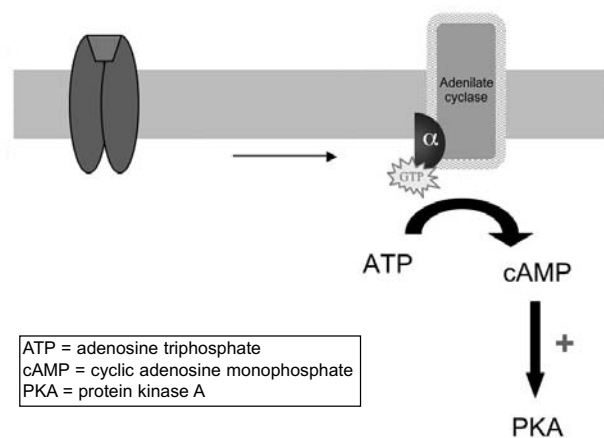
One of the effector proteins regulated by G proteins is adenilate cyclase (AC), an enzyme which catalyzes the formation of cAMP, an important second messenger, from adenosine triphosphate (ATP – see Figure 2). One of the main functions of cAMP is the activation of other enzyme, a cAMP-dependent protein kinase (PKA), which integrates the fast neurotransmission alterations in long-term neurobiological alterations. Several studies demonstrated a significant increase in the activity of basal and activated AC among BD subjects, and these alterations may be associated with the dysfunction of G proteins described above.^{33,40,44-45} Besides, these studies showed a relationship between AC activity and mood treat-

ment or state, with a decrease in the activity of the enzyme among depressed or euthymic patients who relapse after lithium treatment.^{40,44-45} One postmortem study found a decrease in [³H]cAMP binding to PKA in the frontal, temporal, parietal, occipital, thalamic and cerebellum cortices of bipolar subjects, translating an indirect measure of the increase of cAMP activity in these patients.⁴⁶ Two more recent postmortem studies which assessed the frontal and temporal cortices,⁴⁷⁻⁴⁸ and two platelet studies⁴⁹⁻⁵⁰ confirmed an increase in PKA activity in bipolar patients. As a whole, these studies consistently suggest an increase in the activity of the cAMP-PKA pathway in several brain regions of BD subjects.

Studies have also shown that chronic use of lithium decreases AC activation and that this action may be reverted by the increase in GTP concentration, suggesting that the effects of lithium treatment may be attenuated at the level of G- proteins.⁵¹⁻⁵² However, basal lithium has increased the formation of cAMP in rat brains.⁵³ Therefore, it has been suggested that the action of lithium in AC activity is state-dependent: in basal conditions, when the tonic inhibition of the formation of G_s-mediated cAMP is predominant, lithium increases the formation of cAMP; when AC is activated by the receptor-G_s complex, the formation of cAMP is attenuated. This ‘bimodal’ mechanism of action may be one of the explanations of the therapeutic effect of lithium both in depression and mania. Chronic valproate use in clinically-relevant concentration produced a significant increase in b-adrenergic receptors bound to the cAMP pathway on in vitro cells.⁷ Carbamazepine, in turn, shows to inhibit basal⁵⁴ and activated⁵⁵ AC, besides reducing the high levels of cAMP in the liquor of manic patients.⁵⁶

Phosphatidylinositol (PIP₂) pathway

Several neurotransmission systems use the phophatidylinositol pathway (Table 1) through the activation of G proteins. In this pathway, G protein activation stimulates the phospholipase C effector protein (PLC), which hydrolyzes a membrane phospholipide, called phosphatidylinositol (PIP₂), forming two important second messengers: diacilglycerol (DAG) and inositol triphosphate (IP₃). IP₃ has a specific receptor situated in the smooth endoplasmatic reticulum which releases Ca⁺² stocks whenever activated. DAG, in turn,



ATP = adenosine triphosphate
cAMP = cyclic adenosine monophosphate
PKA = protein kinase A

Figure 2 - cAMP pathway

Table 1 – Regulatory effects of neurotransmitters in the intracellular signaling

G-protein bound receptors	cAMP pathway	PIP ₂ pathway
Acetylcholine		
Muscarinic	–	+
GABA	+	
Glutamate – metabotropic		
Type I		+
Type II	–	
Type III	–	
Dopamine		
D ₁	+	
D ₂	–	
Noradrenaline		
α ₁		+
α ₂	–	
β ₁	+	
β ₂	+	
Serotonin		
5-HT ₁	+	
5-HT ₂		+
Histamine		
H ₁		+
H ₂	+	

cAMP = cyclic adenosine monophosphate
 PIP₂ = phosphatidylinositol
 GABA = Gamma-aminobutyric acid

has the function of activating protein kinase C (PKC – Figure 3). Several postmortem and peripheral cells studies have demonstrated alterations of this pathway in subjects with BD. One post-mortem study evidenced increased G protein and PLC activity in the occipital cortex of bipolar subjects, without alterations in frontal and temporal regions.⁵⁷ Other study observed a decrease in G-protein activity linked to the PIP₂ pathway, also in the occipital region.⁵⁸ The authors suggested that these controversial findings may stem from a cellular adaptive process or from the chronic use of lithium. In order to maintain the transmission efficiency of this pathway, the cell needs to keep an adequate supply of inositol for the re-synthesis of PIP₂. As inositol crosses weakly the blood brain barrier, its supply is provided by dephosphorylation of IP₃, through catalization by inositol monophosphatase (IMPase). Shimon et al⁵⁹ compared brains of bipolar, suicide subjects and controls and showed a significant decrease in free inositol in the frontal cortex of bipolar and suicide subjects when compared to the control group, but did not find alterations in IMPase activity on that region. There were no alterations in occipital cortex or cerebellum. Two platelet studies reported increase in PIP₂ levels among non-medicated bipolar subjects, both in their manic⁶⁰ and depressive⁶¹ phase, whereas other studies found significantly reduced PIP₂ levels in platelets of bipolar subjects after lithium treatment.⁶²⁻⁶⁴

These findings support pharmacological studies which demonstrated that lithium in therapeutical concentrations (K_i=0.8mM) is a potent inhibitor of IMPase.⁶⁵ Therefore, this regulatory action of lithium on the PIP₂ pathway may be one of the mood regulating mechanisms of this drug. More recently, one study using magnetic resonance spectroscopy, an exam capable of measuring in vivo brain neurochemical substances, showed that lithium significantly decreased inositol in depressed bipolar subjects' right frontal cortex.⁶⁶ However, although this effect of lithium occurred within 5-7 days, mood improvement has only occurred 3-4 weeks after use of the medication,⁶⁶ suggesting that this initial effect of lithium modulates a series of posterior cascading events, such as the regulation of genic expression and neuronal plasticity, needed to obtain a significant clinical response.

PKC is an important enzyme in the PIP₂ pathway, acting on the regulation of the neuronal excitability, release of neurotransmitters, genic expression and synaptic plasticity.¹ One postmortem study showed significant increase in PKC activity in the frontal cortex of bipolar patients,⁶⁷ a finding also demonstrated on platelet studies.⁶⁸⁻⁶⁹ Soares et al⁶² did not find differences in PKC levels on platelets of euthymic subjects treated with lithium. Actually, lithium also showed effects in the inhibition of PKC activity in animal studies.⁷⁰⁻⁷¹ Therefore, the findings of increase in PKC activity on BD

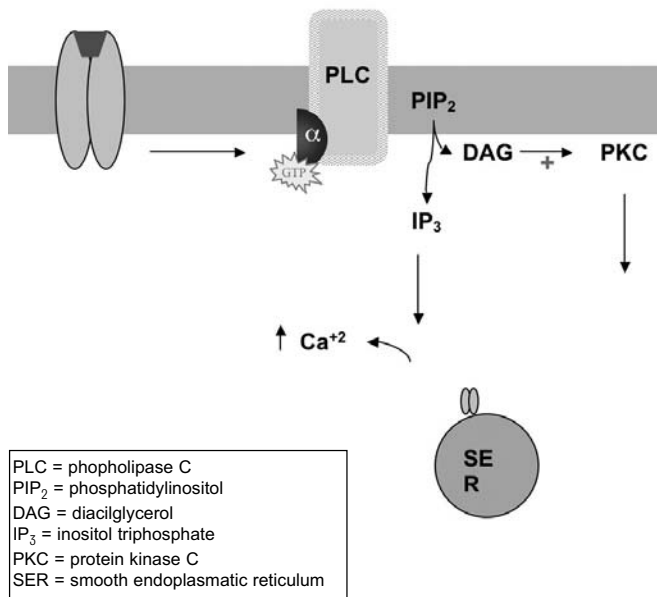


Figure 3 - Phosphatidylinositol

and its decrease with lithium may be clinically relevant. In this sense, Bebchuk et al⁷² have recently published one study showing possible antimanic effects of tamoxifene, a PKC antiestrogenic-inhibitor.

Wnt (wingless)/Glycogen synthase kinase 3β (GSK3-β) pathway

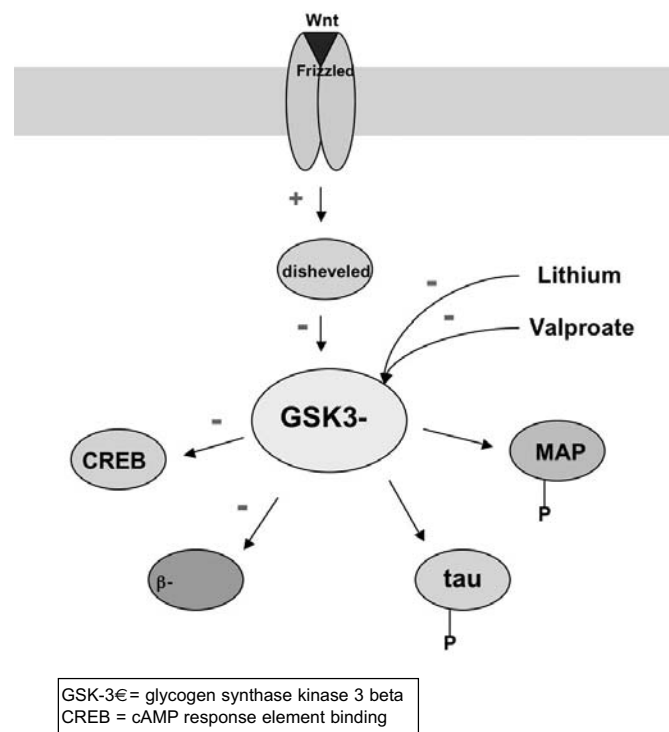
Wnt proteins bind to G-protein binding membrane receptors (frizzled), activating the disheveled protein kinase, which inhibits the glycogen synthase kinase 3β (GSK3-β) activity (Figure 4). The interest in the study of the role of GSK3-β in BD has arisen from observations that lithium and, more recently, valproate decrease the activity of this protein in therapeutical concentrations.⁷³ GSK3-β is able to phosphorylate an extensive range of metabolic, signaling and structural proteins, besides genic transcription factors.⁷⁴ Among these activities, stands out the modulation of proteins associated with cytoskeleton microtubules, such as tau, MAP-1B and MAP-2, and the regulation of programmed cell death (apoptosis).⁷⁵ The phosphorylation of tau and MAP-1B by GSK3-β is associated with the loss or destabilization of microtubules' conformation^{75,76} and the use of lithium showed to decrease the phosphorylation of tau in human neuron culture.⁷⁷ GSK3-β is directly associated with the increase in neuronal apoptosis,⁷⁴ decreasing the activities of proteins which promote the neuronal survival, such as cAMP response element binding protein (CREB) and the heat shock factor-1 (HSF-1).⁷⁸⁻⁷⁹ Besides, lithium, valproate and lamotrigine protected SH-SY5Y cells from apoptosis facilitated by GSK3-β.⁸⁰

GSK3-β activity may be modulated by a series of intracellular signaling cascades. More specifically, the phosphorylation of GSK3-β by PKA, PKC and Akt decreases, while intracellular Ca⁺⁺ may increase its activity.⁷³⁻⁷⁴ Therefore, it has been suggested that the neuroprotecting effects of neurotrophins (NGF, BDNF) and lithium may stem, at least partially, from the inhibition of GSK3-β through the PI3K/Akt pathway.⁸¹ Two postmortem studies did not find alte-

rations in GSK3-β levels on prefrontal cortex of bipolar patients.⁸²⁻⁸³ However, even though there is no direct evidence of abnormalities in the Wnt/GSK3-β pathway on BD, robust evidence highlights the importance of the regulation of this pathway in the treatment of this disorder.^{73,80}

Intracellular Calcium (Ca⁺⁺)

The variation in intracellular calcium levels (Ca⁺⁺) modulates the synaptic plasticity, cell survival and death. In fact, Ca⁺⁺ signaling interacts with several other signaling cascades, including the cAMP and PIP₂ pathways.⁸⁴ Besides, calcium may interact with other regulatory proteins, such as calmoduline, forming complexes which modulate the activity of other important enzymes including calcium-calmodulin dependent protein kinases (CaMKs). Ca⁺⁺-activated PKC decreased PKA activity in fibroblasts,⁸⁵ whereas cAMP caused desensitization of IP₃ receptors and decreased Ca⁺⁺ inflow, reducing therefore Ca⁺⁺ levels.⁸⁶ Dubovsky et al⁸⁷ were the first to describe an increase in Ca⁺⁺ levels in leucocytes and platelets of manic non-medicated bipolar and non-medicated depressed subjects. Besides, using stimuli which increase Ca⁺⁺ concentrations, it was noted a significantly increased response among non-medicated manic bipolar subjects compared to euthymic patients, who were using mood stabilizers.⁸⁸⁻⁸⁹ These studies suggest that the variations in mood state may be related to alterations in Ca⁺⁺ levels and that these alterations may be reverted with the remission of crises,⁸⁷ although other studies have not replicated these findings.⁹⁰⁻⁹¹ Emagoreishi et al⁹² have recently showed that bipolar individuals with high basal Ca⁺⁺ levels had a lower production of cAMP after stimulation of β-adrenergic recep-



GSK3-β = glycogen synthase kinase 3 beta
CREB = cAMP response element binding

Figure 4 - Wnt/GSK3-β pathway

tors with isoproterenol and higher basal activity of AC bound to G protein, corroborating the findings of increased G-protein activity described before and suggesting that alterations in one pathway may decompensate or induce adaptive alterations in other signaling pathways.

Regulation of gene expression and neuroplasticity

The activity of second messenger's pathways has as an important final target the modulation of a family of proteins which act as gene transcription factors. These proteins bind to specific DNA sites and regulate the expression of a wide range of genes capable of regulating cellular functions such as proliferation and apoptosis (Figure 5). One transcription factor that has been studied in BD is CREB, a protein situated in the cell nucleus, usually in its inactive form, being activated by several protein kinases, such as PKA, MAPK or CaMK.⁹⁶ After being activated, CREB promotes the production of mRNA when binding to a specific site in the promoting regions of target genes, with the formation of proteins which may permanently alter specific brain regions' structure or function. Dowlatshahi et al³⁶ did not find significant alterations in CREB levels in bipolar subjects in one postmortem study; however, they found decreased protein levels among individuals who committed suicide and those treated with anticonvulsants at their death. Pharmacological studies which examined the effects of lithium in CREB activity found conflicting results,^{68,78} whereas carbamazepine showed to decrease protein phosphorylation in gliomes.⁵⁴ Besides, the chronic use of ECT and antidepressants increased the expression of BDNF and its receptor TrkB through the increase in CREB activity,⁹³⁻⁹⁴ and GSK3- ϵ decreased BDNF-induced CREB phosphorylation.⁹⁵

Activator protein-1 (AP-1) is a family of gene transcription factors, which include Fos, Jun and ATF, also called immediate early genes, which regulate the genes involved in cellular growth, proliferation and death. The increase in GSK3- ϵ activity in cell cultures decreased significantly AP-1 activity,⁹⁶ whereas the use of lithium showed to revert this action.⁹⁷ Other transcription factor that may be modulated by the Wnt/GSK3- ϵ pathway is b-catenine. The activa-

tion of b-catenine leads to its binding to Tcf/Lef transcription factors, regulating the expression of genes which modulate the circadian rhythm, cellular adherence and development,⁹⁸ being its activity also decreased by GSK3- ϵ .

As reported, recent studies have shown that lithium, valproate and carbamazepine may exert therapeutical effects by means of the regulation of the gene expression through transcription factors. Lithium acts on the several second messengers cascading levels, as well as on protein kinases. Besides, there is evidence of the action of lithium in one of the main brain PKC substrates, MARCKS protein (miristoylated alanine-rich C kinase substrate), which is also a protein related to the regulation of neuroplastic events, altering the conformation of the cytoskeleton through actine filaments. Four-week lithium treatment reduced dramatically the expression of MARCKS protein in hippocampal cells.⁹⁹ Further studies showed that this action of lithium occurred through the regulation of the protein's genic transcription¹⁰⁰ and that it was dependent on inositol concentration and on activation of the receptor linked to the PIP₂ cascade.¹⁰¹ Other study showed that valproate is also capable of reducing MARCKS expression in hippocampal cells, by means of mechanisms which differed from those of lithium,¹⁰² supporting clinical observations on the synergetic therapeutical effect of these two drugs in mood regulation.

Conclusions

Postmortem studies and neuroimaging findings have consistently revealed a significant decrease of volume of determined CNS regions, accompanied by loss or atrophy of neurons, mainly glial cells. Nevertheless, it has not been determined yet if these findings represent early alterations in neuronal migration, cellular losses stemming from the development of the disease proper, biochemical alterations which accompany mood crises or the action of the several medications used. On the other hand, the possible role of cell death/atrophy in the progressive functioning decline found in many patients remains undeciphred. Studies comparing treated patients with those who have never received a medication may help to understand the effects of psychotropics in the cellular morphology and functioning.

Increasing evidence points to the association of intracellular mechanisms, involving the second messengers system as part of the neurobiological alterations of BD. It has been demonstrated an increase in the activity of G proteins and cAMP and PIP₂ pathways, which, by means of the regulation of ADN synthesis, modify the proteins involved in synaptic plasticity, neurogenesis and conformation of the cytoskeleton. However, it is still uncertain if these alterations reflect an increase in the subject's vulnerability (as a result of genetic factors/early life events), effects of established treatments or the disease's central etiological process. Early prospective studies, assessing the alterations of genic expression during the course and treatment of the disorder are a promising research field. Besides, the enhancement of animal models, such as the use of mutant mice, is needed to test if the alterations in determined intracellular signaling cascades suffice to promote behavioral alterations and if the blocking of these pathways are able to inhibit the action of psychopharmacs.

Lastly, pharmacological studies have revealed the action of the main mood stabilizers in several of these intracellular mechanisms. It is possible, therefore, that the acute effects of these

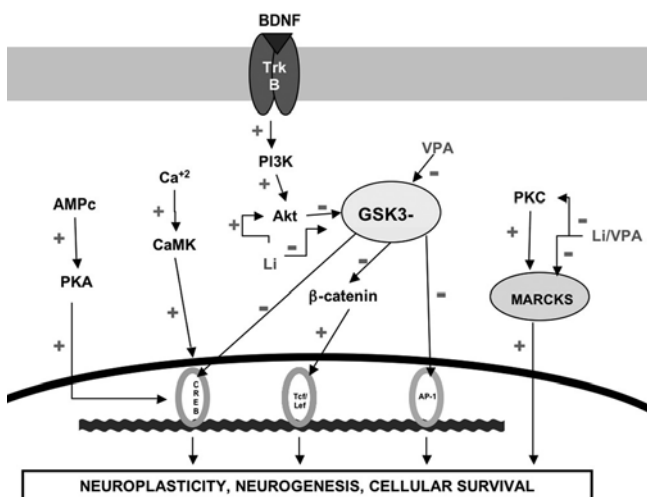


Figure 5 - Regulation of the genic expression

drugs trigger a cascade of intracellular events, capable of altering the proteic synthesis, producing reparatory effects in the synaptic plasticity and restoring the nervous transmission. It is still unknown at which point these drugs may interfere in the pathophysiological alterations of BD and stabilize the course and progression of the disease. These breakthroughs in the neurobiology of BD should be cautiously interpreted, without generalizations, as they are initial studies, which should be replicated and conducted with larger and less heterogeneous samples. Besides, abnormalities involving other pathways, such as the purinergic pathways have been also reported.¹⁰³ We hope that this new and promising research field may help the development of more efficient treatments for such a incapacitating disorder as BD.

Acknowledgements

The authors are thankful to Dr. Betina Kruter for the help in the preparation of this article.

Sponsoring and Conflict of Interest: Inexistent

Receive in 01/05/2004
Accepted in 04/27/2004

References

- Manji HK, Lenox RH. Signaling: cellular insights into the pathophysiology of bipolar disorder. *Biol Psychiatry*. 2000;48(6):518-30.
- Zandi PP, Willour VL, Huo Y, Chellis J, Potash JB, MacKinnon DF, et al. Genome scan of a second wave of NIMH genetics initiative bipolar pedigrees: chromosomes 2, 11, 13, 14, and X. *Am J Med Genet*. 2003;119B(1):69-76.
- Soares JC, Mann JJ. The anatomy of mood disorders - review of structural neuroimaging studies. *Biol Psychiatry*. 1997;41(1):86-106.
- Rajkowska G. Cell pathology in bipolar disorder. *Bipolar Disord*. 2002;4(2):105-16.
- Vawter MP, Freed WJ, Kleinman JE. Neuropathology of bipolar disorder. *Biol Psychiatry*. 2000;48(6):486-504.
- Ghaemi SN, Boiman EE, Goodwin FK. Kindling and second messengers: an approach to the neurobiology of recurrence in bipolar disorder. *Biol Psychiatry*. 1999;45(2):137-44.
- Lenox RH, Frazer A. Mechanism of action of antidepressants and mood stabilizers. In: Davis KL, Charney D, Coyle JT, Nemeroff C, editors. *Neuropsychopharmacology: the fifth generation of progress*. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 1139-63.
- Blumberg HP, Stern E, Martinez D, Ricketts S, de Assis J, White T, et al. Rostral and orbital prefrontal cortex dysfunction in the manic state of bipolar disorder. *Am J Psychiatry*. 1999;156(12):1986-8.
- Drevets WC, Price JL, Simpson Jr JR, Todd RD, Reich T, Vannier M, et al. Subgenual prefrontal cortex abnormalities in mood disorders. *Nature*. 1997;386(6627):824-7.
- Altshuler LL, Conrad A, Hauser P, Li XM, Guze BH, Denikoff K, et al. Reduction of temporal lobe volume in bipolar disorder: a preliminary report of magnetic resonance imaging. *Arch Gen Psychiatry*. 1991;48(5):482-3.
- Hauser P, Altshuler LL, Berrettini W, Dauphinais ID, Gelernter J, Post RM. Temporal lobe measurement in primary affective disorder by magnetic resonance imaging. *J Neuropsychiatry Clin Neurosci*. 1989;1(2):128-34.
- Elkis H, Friedman L, Wise A, Meltzer HY. Meta-analyses of studies of ventricular enlargement and cortical sulcal prominence in mood disorders. *Arch Gen Psychiatry*. 1995;52(9):735-46.
- Sheline Yi, Gado MH, Price JL. Amygdala core nuclei volumes are decreased in recurrent major depression. *Neuroreport*. 1998;9(9):2023-8.
- Pearlson GD, Barta PE, Powers RE, Menon RR, Richards SS, Aylward EH, et al. Medial and superior temporal gyral volumes and cerebral asymmetry in schizophrenia versus bipolar disorder. *Biol Psychiatry*. 1997;41(1):1-4.
- Strakowski SM, DelBello MP, Sax KW, Zimmerman ME, Shear PK, Hawkins JM, et al. Brain magnetic resonance imaging of structural abnormalities in bipolar disorder: a pilot study. *Arch Gen Psychiatry*. 1999;56(3):254-60.
- Altshuler LL, Bartzokis G, Grieder T, Curran J, Mintz J. Amygdala enlargement in bipolar disorder and hippocampal reduction in schizophrenia: an MRI study demonstrating neuroanatomic specificity. *Arch Gen Psychiatry*. 1998;55(7):663-4.
- McDonald WM, Tupler LA, Marsteller FA, Figiel GS, DiSouza S, Nemeroff CB, et al. Hyperintense lesions on magnetic resonance images in bipolar disorder. *Biol Psychiatry*. 1999;45(8):965-71.
- Loeber RT, Sherwood AR, Renshaw PF, Cohen BM, Yurgelun-Todd DA. Differences in cerebellar blood volume in schizophrenia and bipolar disorder. *Schizophr Res*. 1999;37(1):81-9.
- Stoll AL, Renshaw PF, Yurgelun-Todd D, Cohen BM. Neuroimaging in bipolar disorder: what have we learned? *Biol Psychiatry*. 2000;48(6):505-17. Erratum in: *Biol Psychiatry*. 2001;49(1):80.
- Öngür D, Drevets WC, Price JL. Glial reduction in the subgenual prefrontal cortex in mood disorders. *Proc Natl Acad Sci USA*. 1998;95(22):13290-5.
- Rajkowska G, Halaris A, Selemon LD. Reductions in neuronal and glial density characterize the dorsolateral prefrontal cortex in bipolar disorder. *Biol Psychiatry*. 2001;49(9):741-52.
- Cotter D, Mackay D, Landau S, Kerwin R, Everall I. Reduced glial cell density and neuronal size in the anterior cingulate cortex in major depressive disorder. *Arch Gen Psychiatry*. 2001;58(6):545-53.
- Uranova N, Orlovskaya D, Vikhrevva O, Zimina I, Kolomeets N, Vorstrikov V, et al. Electron microscopy of oligodendroglia in severe mental illness. *Brain Res Bull*. 2001;55(5):597-610.
- Benes FM, Kwok EW, Vincent SL, Todtenkopf MS. A reduction of non-pyramidal cells in sector CA2 of schizophrenics and manic-depressives. *Biol Psychiatry*. 1998;44(2):88-97.
- Benes FM, Vincent SL, Todtenkopf MS. The density of pyramidal and nonpyramidal neurons in anterior cingulate cortex of schizophrenic and bipolar subjects. *Biol Psychiatry*. 2001;50(6):395-406.
- Baumann B, Danos P, Krell D, Diekmann S, Wurthmann C, Bielau H, et al. Unipolar-bipolar dichotomy of mood disorder in supported by noradrenergic brainstem system morphology. *J Affect Disord*. 1999;54(1-2):217-24.
- Young LT, Warsh JJ, Kish SJ, Shannak K, Hornykeiwicz O. Reduced brain 5-HT and elevated NE turnover and metabolites in bipolar affective disorder. *Biol Psychiatry*. 1994;35(2):121-7.
- Magistretti PJ, Ransom BR. Astrocytes. In: Davis KL, Charney D, Coyle JT, Nemeroff C, editors. *Neuropsychopharmacology: the fifth generation of progress*. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 133-45.
- Eastwood SL, Harrison PJ. Hippocampal synaptic pathology in schizophrenia, bipolar disorder, and major depression: a study of complexin mRNAs. *Mol Psychiatry*. 2000;5(4):425-32.
- Rosoklija G, Toomayan G, Ellis SP, Keilp J, Mann JJ, Latov N, et al. Structural abnormalities of subicular dendrites in subjects with schizophrenia and mood disorders. *Arch Gen Psychiatry*. 2000;57(4):349-56.
- Avissar S, Schreiber G, Danon A, Belmaker RH. Lithium inhibits adrenergic and cholinergic increases in GTP binding in rat cortex. *Nature*. 1988;331(6155):440-2.
- Mork A, Geisler A. The effects of lithium in vitro and ex vivo on adenylylase cyclase in brain are exerted by distinct mechanisms. *Neuropharmacology*. 1989;28(3):307-11.
- Young LT, Li PP, Kish SJ, Siu KP, Warsh JJ. Postmortem cerebral cortex Gs alpha-subunit levels are elevated in bipolar in bipolar affective disorder. *Brain Res*. 1991;553(2):323-6.
- Young LT, Li PP, Kish SJ, Siu KP, Kamble A, Hornykiewicz O, et al. Cerebral

- cortex Gs alpha protein levels and forskolin-stimulated cyclic AMP formation are increased in bipolar affective disorder. *J Neurochem.* 1993;61(3):890-8.
35. Friedman E, Wang HY. Receptor-mediated activation of G proteins is increased in postmortem brains of bipolar affective disorder subjects. *J Neurochem.* 1996;67(3):1145-52.
36. Dowlatshahi D, MacQueen GM, Wang JF, Reiaich JS, Young LT. Protein-coupled cyclic AMP signaling in post mortem brain of subjects with mood disorders: effects of diagnosis, suicide, and treatment at time of death. *J Neurochem.* 1999;73(3):1121-6.
37. Schreiber G, Avissar S, Danon A, Belmaker RH. Hyperfunctional G proteins in mononuclear leukocytes in patients with mania. *Biol Psychiatry.* 1991;29(3):273-80.
38. Spleiss O, Van Calker D, Scharer L, Adamovic K, Berger M, Gebicke-Haerter PJ. Abnormal G protein alpha(s)- and alpha(i2)- subunit mRNA expression in bipolar affective disorder. *Mol Psychiatry.* 1998;3(6):512-20.
39. Young LT, Li PP, Kamble A, Siu KP, Warsh JJ. Mononuclear leukocyte levels of G proteins in depressed patients with bipolar disorder and major depressive disorder. *Am J Psychiatry.* 1994;151(4):594-6.
40. Avissar S, Nechamkin Y, Barki-Harrington L, Roitman G, Schreiber G. Differential G protein measures in mononuclear leukocytes of patients with bipolar mood disorder are state dependent. *J Affect Disord.* 1997;43(2):85-93.
41. Mitchell PB, Manji HK, Chen G, Jolkovsky L, Smith-Jackson E, Denicoff K, et al. High levels of Gs alpha in platelets of euthymic patients with bipolar affective disorder. *Am J Psychiatry.* 1997;154(2):218-23.
42. Manji HK, Chen G, Shimon H, Hsiao JK, Potter WZ, Belmaker RH. Guanine nucleotide-binding proteins in bipolar affective disorder: effects of long-term lithium treatment. *Arch Gen Psychiatry.* 1995;52(2):135-44.
43. Alda M, Keller D, Grof E, Turecki G, Cavazzoni P, Duffy A, et al. Is lithium response related to G(s) alpha levels in transformed lymphoblasts from subjects with bipolar disorder? *J Affect Disord.* 2001;65(2):117-22.
44. Avissar S, Barki-Harrington L, Nechamkin Y, Roitman G, Schreiber G. Reduced beta-adrenergic receptor-coupled Gs protein function and Gs alpha immunoreactivity in mononuclear leukocytes of patients with depression. *Biol Psychiatry.* 1996;39(9):755-60.
45. Ebstein RP, Lerer B, Shapira B, Shemesh Z, Moscovich DG, Kindler S. Cyclic AMP second-messenger signal amplification in depression. *Br J Psychiatry.* 1988;152:665-9.
46. Rahman S, Li PP, Young LT, Kofman O, Kish SJ, Warsh JJ. Reduced [3H]cyclic AMP binding in postmortem brain from subjects with bipolar affective disorder. *J Neurochem.* 1997;68(1):297-304.
47. Chang A, Li PP, Warsh JJ. Altered cAMP-dependent protein kinase subunit immunolabeling in postmortem brain from patients with bipolar affective disorder. *J Neurochem.* 2003;84(4):781-91.
48. Fields A, Li PP, Kish SJ, Warsh JJ. Increased cyclic AMP-dependent protein kinase activity in postmortem brain from patients with bipolar affective disorder. *J Neurochem.* 1999;73(4):1704-10.
49. Perez J, Tardito D, Mori S, Racagni G, Smeraldi E, Zanardi R. Altered Rap1 endogenous phosphorylation and levels in platelets from patients with bipolar disorder. *J Psychiatr Res.* 2000;34(2):99-104.
50. Perez J, Zanardi R, Mori S, Gasperini M, Smeraldi E, Racagni G. Abnormalities of cAMP-dependent endogenous phosphorylation in platelets from patients with bipolar disorder. *Am J Psychiatry.* 1995;152(8):1204-6.
51. Newman ME, Belmaker RH. Effects of lithium in vitro and ex vivo on components of the adenylate cyclase system in membranes from the cerebral cortex of the rat. *Neuropharmacology.* 1987;26(2-3):211-7.
52. Newman M, Klein E, Birmaher B, Feinsod M, Belmaker RH. Lithium at therapeutic concentrations inhibits human brain noradrenaline-sensitive cyclic AMP accumulation. *Brain Res.* 1983;278(1-2):380-1.
53. Masana MI, Bitran JA, Hsiao JK, Potter WZ. In vivo evidence that lithium inactivates G[i] modulation of adenylate cyclase in brain. *J Neurochem.* 1992;59(1):200-5.
54. Chen G, Pan B, Hawver D, Wright C, Potter WZ, Manji HK. Attenuation of cyclic AMP production by carbamazepine. *J Neurochem.* 1996;67(5):2079-86.
55. Elphick M, Anderson SM, Hallis KF, Grahame-Smith DG. Effects of carbamazepine on 5-hydroxytryptamine function in rodents. *Psychopharmacology. (Berl.)* 1990;100(1):49-53.
56. Post RM, Ballenger JC, Uhde TW, Smith C, Rubinow DR, Bunney WE Jr. Effect of carbamazepine on cyclic nucleotides in CSF of patients with affective illness. *Biol Psychiatry.* 1982;17(9):1037-45.
57. Mathews R, Li PP, Young LT, Kish SJ, Warsh JJ. Increased G alpha q/11 immunoreactivity in postmortem occipital cortex from patients with bipolar affective disorder. *Biol Psychiatry.* 1997;41(6):649-56.
58. Jope RS, Song L, Li PP, Young LT, Kish SJ, Pacheco MA, et al. The phosphoinositide signal transduction system is impaired in bipolar affective disorder brain. *J Neurochem.* 1996;66(6):2402-9.
59. Shimon H, Agam G, Belmaker RH, Hyde TM, Kleinman JE. Reduced frontal cortex inositol levels in postmortem brain of suicide victims and patients with bipolar disorder. *Am J Psychiatry.* 1997;154(8):1148-50.
60. Brown AS, Mallinger AG, Renbaum LC. Elevated platelet membrane phosphatidylinositol-4,5-biphosphate in bipolar mania. *Am J Psychiatry.* 1993;150(8):1252-4.
61. Soares JC, Dippold CS, Wells KF, Frank E, Kupfer DJ, Mallinger AG. Increased platelet membrane phosphatidylinositol-4,5-biphosphate in drug-free bipolar patients. *Neurosci Lett.* 2001;299(1-2):150-2.
62. Soares JC, Chen G, Dippold CS, Wells KF, Frank E, Kupfer DJ, et al. Concurrent measures of protein kinase C and phosphoinositides in lithium-treated bipolar patients and healthy individuals: a preliminary study. *Psychiatry Res.* 2000;95(2):109-18.
63. Soares JC, Mallinger AG, Dippold CS, Frank E, Kupfer DJ. Platelet membrane phospholipids in euthymic bipolar disorder patients: are they affected by lithium treatment? *Biol Psychiatry.* 1999;45(4):453-7.
64. Soares JC, Dippold CS, Mallinger AG. Platelet membrane phosphatidylinositol-4,5-biphosphate alterations in bipolar disorder - evidence from a single case study. *Psychiatry Res.* 1997;69(2-3):197-202.
65. Sherman WR, Gish BG, Honchar MP, Munsell LY. Effects of lithium on phosphoinositide metabolism in vivo. *Fed Proc.* 1986;45(11):2639-46.
66. Moore GJ, Bechuk JM, Parrish JK, Faulk MW, Arfken CL, Strahl-Bevaqua J, et al. Temporal dissociation between lithium-induced changes in frontal lobe myo-inositol and clinical response in manic-depressive illness. *Am J Psychiatry.* 1999;156(12):1902-8.
67. Wang HY, Friedman E. Enhanced protein kinase C activity and translocation in bipolar affective disorders brains. *Biol Psychiatry.* 1996;40(7):568-75.
68. Wang HY, Markowitz P, Levinson D, Undie AS, Friedman E. Increased membrane-associated protein kinase C activity and translocation in blood platelets from bipolar affective disorder patients. *J Psychiatr Res.* 1999;33(2):171-9.
69. Friedman E, Hoau YW, Levinson D, Connell TA, Singh H. Altered platelet protein kinase C activity in bipolar affective disorder, manic episode. *Biol Psychiatry.* 1993;33(7):520-5.
70. Manji HK, Etcheberrigaray R, Chen G, Olds JL. Lithium dramatically decreases membrane-associated protein kinase C in the hippocampus: selectivity for the alpha isozyme. *J Neurochem.* 1993;61(6):2303-10.
71. Chen G, Rajkowska G, Du F, Seraji-Bozorgzad N, Manji HK. Enhancement of hippocampal neurogenesis by lithium. *J Neurochem.* 2000;75(4):1729-34.
72. Bechuk JM, Arfken CL, Dolan-Manji S, Murphy J, Hasanat K, Manji HK. A preliminary investigation of a protein kinase C inhibitor in the treatment of acute mania. *Arch Gen Psychiatry.* 2000;57(1):95-7.
73. Gould TD, Manji HK. The Wnt signaling pathway in bipolar disorder. *Neuroscientist.* 2002;8(5):497-511.
74. Grimes CA, Jope RS. The multifaceted roles of glycogen synthase kinase 3b in cellular signaling. *Prog Neurobiol.* 2001;65(4):391-426. Erratum in: *Prog Neurobiol.* 2001;65(5):497.
75. Johnson GV, Hartigan JA. Tau protein in normal in Alzheimer's disease

- brain: an update. *J Alzheimers Dis.* 1(4-5):329-41.
76. Lucas FR, Goold RG, Gordon-Weeks PR, Salinas PC. Inhibition of GSK-3 β leading to the loss of phosphorylated MAP-1B is an early event in axonal remodeling induced by WNT-7a or lithium. *J Cell Sci.* 1998;111(Pt 10):1351-61.
77. Hong M, Chen DC, Klein PS, Lee VM. Lithium reduces tau phosphorylation by inhibition of glycogen synthase kinase-3. *J Biol Chem.* 1997;272(40):25326-32.
78. Grimes CA, Jope RS. CREB DNA binding activity is inhibited by glycogen synthase kinase-3 beta and facilitated by lithium. *J Neurochem.* 2001;78(6):1219-32.
79. He B, Meng YH, Mivechi NF. Glycogen synthase kinase 3beta and extracellular signal-regulated kinase inactivate heat shock transcription factor 1 by facilitating the disappearance of transcriptionally active granules after heat shock. *Mol Cell Biol.* 1998;18(11):6624-33.
80. Li X, Bijur GN, Jope RS. Glycogen synthase kinase 3-beta, mood stabilizers, and neuroprotection. *Bipolar Disord.* 2002;4(2):137-44.
81. Chuang DM, Chen RW, Chalecka-Franaszek E, Ren M, Hashimoto R, Senatorov V, et al. Neuroprotective effects of lithium in cultured cells and animal models of disease. *Bipolar Disord.* 2002;4(2):129-36.
82. Kozlovsky N, Belmaker RH, Agam G. Low GSK-3beta immunoreactivity in postmortem frontal cortex of schizophrenic patients. *Am J Psychiatry* 2000;157:831-3.
83. Lesort M, Greendorfer A, Stockmeier C, Johnson GV, Jope RS. Glycogen synthase kinase 3-beta, beta-catenin, and tau in postmortem bipolar brain. *J Neural Transm.* 1999;106(11-12):1217-22.
84. Liu M, Simon MI. Regulation by cAMP-dependent protein kinase of a G protein-mediated phospholipase C. *Nature* 1996;382(6586):83-7.
85. Dobbeling U, Berchtold MW. Down-regulation of the protein kinase A pathway by activators of protein kinase C and intracellular Ca²⁺ in fibroblasts cells. *FEBS Lett.* 1996;391(1-2):131-3.
86. Supattapone S, Danoff SK, Theibert A, Joseph SK, Steiner J, Snyder SH. Cyclic AMP-dependent phosphorylation of a brain inositol triphosphate receptor decreases its release of calcium. *Proc Natl Acad Sci USA.* 1988;85(22):8747-50.
87. Dubovsky SL, Murphy J, Thomas M, Rademacher J. Abnormal intracellular calcium ion concentration in platelets and lymphocytes of bipolar patients. *Am J Psychiatry.* 1992;149(1):118-20.
88. Okamoto Y, Kagaya A, Shinno H, Motohashi N, Yamawaki S. Serotonin-induced platelet calcium mobilization is enhanced in mania. *Life Sci.* 1995;56(5):327-32.
89. Kusumi I, Koyama T, Yamashita I. Trombin-induced platelet calcium mobilization is enhanced in bipolar disorder. *Biol Psychiatry.* 1992;32(8):731-4.
90. Suzuki K, Kusumi I, Sasaki Y, Koyama T. Serotonin-induced platelet intracellular calcium mobilization in various psychiatric disorders: is it specific to bipolar disorder? *J Affect Disord.* 2001;64(2-3):291-6.
91. El Khoury AE, Petterson U, Kallner G, Aberg-Wistedt A, Stain-Malmgren R. Calcium homeostasis in long-term lithium-treated women with bipolar affective disorder. *Prog Neuropsychopharmacol Biol Psychiatry.* 2002;26(6):1063-9.
92. Emamghoreishi M, Li PP, Shlichter L, Parikh SV, Cooke R, Warsh JJ. Associated disturbances in calcium homeostasis and G protein-mediated camp signaling in bipolar I disorder. *Biol Psychiatry.* 2000;48(7):665-73.
93. Chen B, Dowlatshahi D, McQueen GM, Wang GF, Young LT. Increased hippocampal BDNF immunoreactivity in subjects treated with antidepressant medication. *Biol Psychiatry.* 2001;50(4):260-5.
94. Thome J, Sakai N, Shin KH, Steffen C, Zhang YJ, Impey S, et al. cAMP response element-mediated gene transcription is up-regulated by chronic antidepressant treatment. *J Neurosci.* 2000;20(11):4030-6.
95. Mai L, Jope RS, Li X. BDNF-mediated signal transduction is modulated by GSK3beta and mood stabilizing agents. *J Neurochem.* 2002;82(1):75-83.
96. Nikolakaki E, Coffey PJ, Hemelsoet R, Woodgett JR, Defize LH. Glycogen synthase kinase phosphorylates Jun family members in vitro and negatively regulates their transactivating potential in intact cells. *Oncogene.* 1993;8(4):833-40.
97. Hedgepeth CM, Conrad LJ, Zhang J, Huang HC, Lee VM, Klein PS. Activation of the Wnt pathway: a molecular mechanism of lithium action. *Dev Biol.* 1997;185(1):82-91.
98. Novak A, Dedhar S. Signaling through b-catenin and Lef/Tcf. *Cell Mol Life Sci.* 1999;56(5-6):532-7.
99. Lenox RH, Watson DG, Patel J, Ellis J. Chronic lithium administration alters a prominent PKC substrate in rat hippocampus. *Brain Res.* 1992;570(1-2):333-40.
100. Wang L, Liu X, Lenox RH. Transcriptional down-regulation of MARCKS gene expression in immortalized hippocampal cells by lithium. *J Neurochem.* 2001;79(4):816-25.
101. Manji HK, Bersudsky Y, Chen G, Belmaker RH, Potter WZ. Modulation of protein kinase C isozymes and substrates by lithium: the role of myo-inositol. *Neuropsychopharmacology.* 1996;15(4):370-81.
102. Watson DG, Watterson JM, Lenox RH. Sodium valproate down-regulates the miristoylated alanine-rich C kinase substrate (MARCKS) in immortalized hippocampal cells: a property unique to PKC-mediated mood stabilizers. *J Pharmacol Exp Ther.* 1998;285(1):307-16.
103. Machado-Vieira R, Lara DR, Souza DO, Kapczinski F. Purinergic dysfunction in mania: an integrative model. *Med Hypotheses.* 2002;58(4):297-304.

Correspondence

Benício Noronha Frey
 Rua Almirante Abreu, 108/201.
 90420-010 Porto Alegre, RS, Brazil
 Phones: + 55 51 3333.4821 / + 55 51 9916.7666
 E-mail: benicio.frey@terra.com.br
