

# Mechanisms of muscle wasting in sarcopenia

Vivian de Oliveira Nunes Teixeira<sup>1</sup>, Lidiane Isabel Filippin<sup>2</sup>, Ricardo Machado Xavier<sup>3</sup>

## ABSTRACT

Approximately 66% of the patients with rheumatoid arthritis (RA) have significant loss of cell mass (rheumatoid cachexia), mainly of skeletal muscle (rheumatoid sarcopenia). Sarcopenia is defined as muscle wasting associated with functional impairment. Patients with RA possess significant reduction in muscle strength, caused by muscle protein wasting, and loss of functionality. Various conditions leading to muscle wasting involve different pathways of intracellular signaling that trigger: (i) programmed cell death (apoptosis); (ii) increased protein degradation through autophagy, calcium-dependent proteases (calpains and caspases), and proteasome system; (iii) decreased satellite cell activation, responsible for muscle regeneration. This article aimed at reviewing these general mechanisms of sarcopenia and their involvement in RA. Greater knowledge of these mechanisms may lead to the development of innovative therapies to this important comorbidity.

**Keywords:** muscular atrophy, inflammation, regeneration, rheumatoid arthritis.

© 2012 Elsevier Editora Ltda. All rights reserved.

## INTRODUCTION

Rheumatoid arthritis (RA) is a systemic inflammatory disease of unknown etiology with autoimmune manifestations, characterized by symmetrical and erosive chronic synovitis, preferentially affecting peripheral joints.<sup>1</sup> In most patients, the rheumatoid factor can be detected. The prevalence of RA is approximately 0.46% in the Brazilian population<sup>2</sup> and 1% in the world population.<sup>3</sup> The disease affects mainly women aged between 30 and 60 years.

In addition to articular manifestations, RA has several systemic manifestations that significantly influence its morbidity and mortality. Rheumatoid cachexia<sup>4</sup> occurs in approximately 66% of patients with RA, being characterized by cell mass loss, predominantly in the skeletal muscle (rheumatoid sarcopenia), accompanied by maintenance or slight elevation of the fat mass (total of adipose tissue), which results in limited or no weight loss (total mass). The etiology of rheumatoid cachexia is multifactorial, including the increased production of pro-inflammatory cytokines, mainly TNF- $\alpha$  and IL-1 $\beta$ , hormonal changes, and physical inactivity. So far, no standardized

therapy has been proposed aiming specifically at that aspect of RA, and the effects of the current treatments have not been well studied.

This study aimed at reviewing the molecular mechanisms involved in sarcopenia, more specifically in rheumatoid sarcopenia. For reviewing the clinical aspects of rheumatoid sarcopenia, the article by Rocha et al.<sup>4</sup> is recommended.

## SARCOPENIA

Sarcopenia is muscle wasting associated with functional impairment. It results from several factors, such as innervation disorders, physical activity reduction, ageing, metabolic abnormalities (especially in proteins, carbohydrates, and lipids), in addition to changes in the activation of satellite cells.<sup>4,5</sup> In RA, the following are believed to play a role in the development of sarcopenia: the action of pro-inflammatory cytokines; the reduction in protein synthesis in myocytes; physical activity limitation; insulin resistance; and inadequate protein ingestion.<sup>6,7</sup>

The diagnosis of sarcopenia can be performed by use of several methods, such as nuclear magnetic resonance, computed

Received on 02/17/2011. Approved on 12/14/2011. The authors declare no conflict of interest. Financial Support: Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) – Edital Universal CNPq/Bolsa CNPq/PDJ –, and Fundo de Incentivo à Pesquisa e Eventos (FIPE).

Rheumatology Service, Hospital de Clínicas de Porto Alegre.

1. Student of the Post-Graduation Program in Medical Sciences, Universidade Federal do Rio Grande do Sul – UFRGS; Bachelor's degree in Biological Sciences, UFRGS

2. PhD in Medical Sciences, UFRGS; Professor, Centro Universitário Franciscano – UNIFRA

3. PhD in Immunology, Shimane Medical University; Professor of the Department of Internal Medicine, Medical School, UFRGS

Correspondence to: Vivian de Oliveira Nunes Teixeira. Rua Ramiro Barcelos, 2350/645 – Rio Branco. CEP: 90035-903. Porto Alegre, RS, Brasil. E-mail: viviont@gmail.com

tomography, bioimpedance, ultrasonography, total body bone densitometry, and anthropometric measures. Densitometry is often used because it provides assessment of the body composition, bone mass, lean mass, and total fat mass.<sup>8</sup> The anthropometric measures proposed by Ashwell, including waist-to-hip ratio, have also been used to assess sarcopenia.<sup>9</sup>

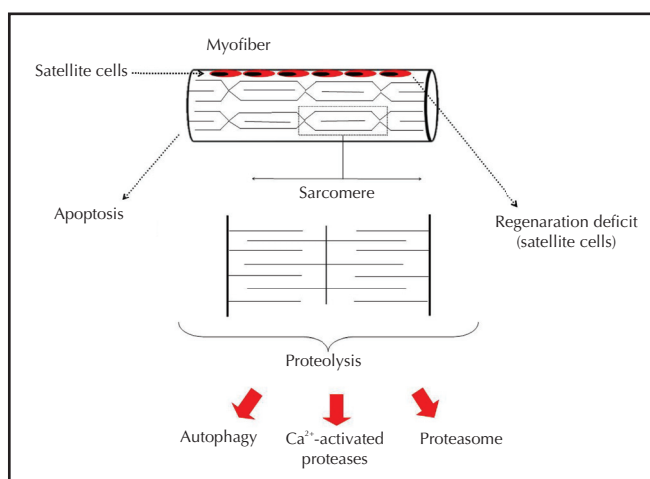
## MOLECULAR MECHANISMS INVOLVED IN SARCOPENIA

The different conditions leading to muscle wasting involve different cell signaling pathways that might lead to programmed cell death (apoptosis), increased protein breakdown, or even decreased activation of the satellite cells responsible for muscle regeneration (Figure 1). Most of our knowledge about those mechanisms derives from studies with experimental models of atrophy, such as models of denervation, hind limb unloading, disuse, fasting,<sup>10</sup> *diabetes mellitus*, and cancer,<sup>11</sup> as well as from studies with muscle biopsies of volunteer patients.<sup>12</sup>

Such mechanisms and the current knowledge about their involvement in rheumatoid sarcopenia are discussed.

### Cell mass loss

Apoptosis is an important process that occurs in multicellular organisms, both during normal development and for maintaining tissue homeostasis.<sup>13</sup> However, the role of apoptosis in post-mitotic tissues, such as skeletal muscle, has not been clarified.



**Figure 1**

Molecular mechanisms involved in sarcopenia. Muscle wasting can occur through distinct mechanisms, such as a deficiency in regeneration due to inactivity of satellite cells, apoptosis, and protein degradation pathways, such as calcium-activated proteases, proteasome, and autophagy.

The initial stage of apoptosis involves the induction of signals of cell death, which cause unbalance in the regulation of free calcium and alteration in the composition of some protein families.<sup>14</sup> After that stage, cell surface receptors or mitochondrial pathways are activated, triggering cytoplasmic and nuclear events that lead to cell death.<sup>15</sup> Caspases are the major enzymes involved in the beginning and development of apoptosis. They account for proteolytic cleavage of a wide range of cell targets,<sup>16</sup> although they do not exclusively initiate that process.<sup>17</sup>

Regarding the potential participation of apoptosis in sarcopenia, even in a model of marked atrophy, such as that of muscle denervation in mice, evidence of significant apoptosis has only been observed after two months, indicating a limited role of that mechanism in the initial stages of atrophy.<sup>18</sup>

### Muscle proteolysis

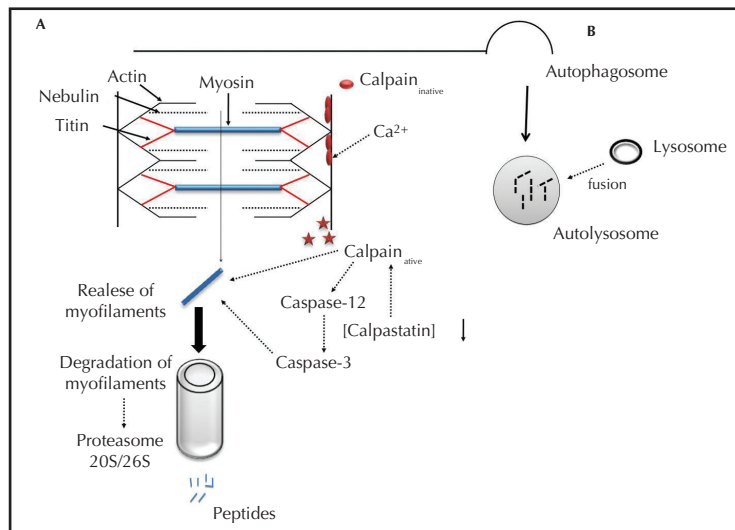
Sarcopenia is the result of unbalance between protein degradation and synthesis, although apparently the exact contribution to each of those factors varies according to the model studied.

Some proteolytic systems have been described as participating in muscle degradation, and the following are examples: autophagy; calcium-activated proteases, such as calpain and caspases; and the ubiquitin-proteasome system (Figure 1).<sup>19,20</sup>

In *in vivo* experimental models and in humans, there is no consensus about the relative importance of the different protein degradation pathways. Purintrapiban et al.<sup>20</sup> have studied the role of these different mechanisms of proteolysis in muscle cell culture. The inhibition of the calpain, proteasome, and lysosome enzymatic systems caused a 20%, 62%, and 40% reduction in total protein degradation, respectively. However, the participation of each of those pathways varies significantly, depending on the clinical situation involved, such as denervation, immobilization, malignant cachexia, and chronic inflammation.<sup>19-21</sup>

### Autophagy

Autophagy is a very old mechanism of cell survival that allows cell self-consumption during periods of extreme nutritional deprivation.<sup>22</sup> Such process occurs with the consumption of cytoplasmic components, such as cytosol and cell organelles, and is lysosome dependent. During autophagy, double-membrane vesicles (the autophagosomes) form around large part of the cytoplasm or whole organelles, sequestering the protein substrates in the vacuolar system. Then, the autophagosome fuses with the lysosome, forming an autolysosome, and then, the substrates are hydrolyzed by lysosomal hydrolases<sup>23</sup>



**Figure 2**

Muscle proteolysis pathways. A: activation of calcium-activated proteases. Calpains cleave proteins that anchor the actin-myosin complex, releasing those proteins to be degraded by another cell proteolysis system (proteasome). B: the autophagy system, in which the cytoplasmic constituents are isolated and degraded in an autolysosome.

(Figure 2). Such hydrolases are physically isolated from the cytoplasmic constituents by the lysosomal membrane, and, thus, have greater capacity to degrade cytoplasmic components, as compared with myofibrillar components.<sup>24</sup>

*In vitro*<sup>25</sup> and *in vivo*<sup>26</sup> studies have evidenced the presence of autophagosomes in muscle fibers of myotube culture and in mice. An *in vitro* study with myocyte culture under amino acid restriction has shown that the acceleration of protein catabolism was mainly due to autophagy induction.<sup>25</sup> In an *in vivo* study, Mizushima et al.<sup>26</sup> have shown, through the observation of overexpression of microtubule-associated protein 1 light chain 3 (LC3), the activation of the autophagy system in skeletal muscle of fasting mice. LC3 is essential to maintain membrane integrity and cell growth, and is overexpressed, along with other genes involved with autophagy and muscle wasting, in different models of atrophy,<sup>27,28</sup> in addition to being an indicator of autophagic activity.<sup>29</sup>

Despite the existence of distinct mechanisms of sarcopenia, the pathways that activate the autophagy and ubiquitin-proteasome systems are common. Both pathways involve the forkhead box O3 (FOXO3) transcription factor and the nuclear transcription factor kappa-B (NF- $\kappa$ B). FOXO3 is translocated to the nucleus in the absence of stimuli of protein synthesis,<sup>30</sup> while NF- $\kappa$ B is translocated in the presence of inflammation.<sup>31</sup> FOXO3 has been identified as a critical factor for controlling muscle autophagy,<sup>32</sup> and several genes of autophagy are regulated by that transcription factor.<sup>30</sup>

### Calcium-activated proteases: calpain and caspases

The calpain system is a protein-degradation pathway of eukaryotic cells composed of two enzymes (calpains) and calpastatin.

Such proteases are calcium-dependent, non-lysosomal cysteine proteases,<sup>33</sup> and have an endogenous inhibitor, calpastatin, which regulates their activity<sup>21</sup> (Figure 2).

Calpains cannot degrade proteins into amino acids or small peptides and do not catalyze the degradation of the complex of sarcoplasmic proteins. Although they do not directly degrade muscle contractile proteins, calpains cleave the proteins that anchor the actin-myosin complex, releasing the protein components of the sarcomere to be degraded by another cell proteolysis system.<sup>20,34</sup> The substrates of calpain include titin, nebulin, desmin and filamin, proteins that anchor the sarcomere,<sup>33,35</sup> and also troponin and tropomyosin of the sarcomere,<sup>33,36</sup> which release the actin-myosin complex.

Activation of the calpain system has already been shown in several situations of muscle atrophy, such as during prolonged periods of inactivity,<sup>33</sup> ageing, dystrophies, and other pathologies that accompany muscle wasting.<sup>24</sup>

Caspases are non-calcium-dependent cytoplasmic cysteine-proteases that can cleave other proteins after an aspartic acid residue, an uncommon specificity among proteases.<sup>24</sup>

Caspase-3 seems to be able to degrade the actin-myosin complex. Du et al.<sup>37</sup> have shown that purified and activated caspase-3 can cleave actin, breaking the muscle actin-myosin complex, releasing those proteins to be degraded by other proteolytic complexes<sup>16</sup> (Figure 2). Although activated in muscle wasting, the real role of those caspases is still controversial.

### Proteasome

Another proteolytic system related to sarcopenia and currently considered one of the most important is the ubiquitin-proteasome system. This highly conserved system is the major

machinery of non-lysosomal protein degradation in eukaryotic cells<sup>38</sup> (Figure 2).

The ubiquitin-proteasome system is responsible for processing and degrading cell proteins essential for the regulation of development, differentiation, proliferation, apoptosis, signal transduction, and immune and inflammatory response, governing, thus, basic cell processes.<sup>39,40</sup>

Cell proteins destined for degradation by proteasome should be properly labeled with a covalent bond of multiple ubiquitin monomers, peptides composed of 76 amino acids. Ubiquitin can be conjugated with specific protein substrates, a process that requires three enzymes (Figure 3): E1, an ubiquitin-activating enzyme; E2, an ubiquitin-conjugating enzyme; and E3, an ubiquitin ligase. Initially, E1 is activated and, in an energy-dependent reaction, transfers, through E2, ubiquitin to E3, which catalyzes ubiquitin binding to the protein, labeling it for degradation.<sup>41</sup> This process of degradation of polyubiquitinated proteins occurs in the proteasome (20S or 26S), which is a complex composed of one or three large enzymes with the function of degrading unnecessary or damaged cell proteins.<sup>19</sup>

The E3 enzymes provide specificity to the target protein for degradation. Hundreds of different E3 have already been identified, and each one seems to modulate the ubiquitination of a group of protein substrates.<sup>41</sup> In the skeletal muscle, two

specific E3 related to the atrophy process – MAF-bx (muscle atrophy F-box) or atrogin-1, and MuRF-1 (Muscle Ring Finger-1) – were identified.<sup>42</sup> A third E3 ubiquitin ligase, NEDD-4, has been reported, and it seems to facilitate muscle atrophy in denervation and hind limb unloading models.<sup>43</sup>

MuRF-1 is an E3 ubiquitin ligase recognized as a marker of the muscle atrophy process in several experimental models.<sup>44</sup> This protein can bind to titin of the M line,<sup>45</sup> the third in amount among muscle proteins (10%).<sup>46</sup>

Some studies have reported the increased expression of subunits of the proteasome and ubiquitinating enzymes during muscle atrophy,<sup>47</sup> as well as the increase in the expression of E3 ligases in models of denervation, immobilization, food restriction, *diabetes mellitus*, and uremia.<sup>44</sup> Such studies have suggested that muscle wasting is related with the activity of MuRF-1 and atrogin-1 E3 ligases.<sup>44</sup> In murine models, proteasome inhibition can reduce protein degradation during atrophy,<sup>48</sup> indicating an important role of the ubiquitin-proteasome pathway in sarcopenia. Such results, however, cannot be extrapolated to humans.<sup>49</sup> Biological and synthetic inhibitors of proteasome can inhibit the cell cycle and induce apoptosis, preferentially in neoplastic cells.<sup>50</sup>

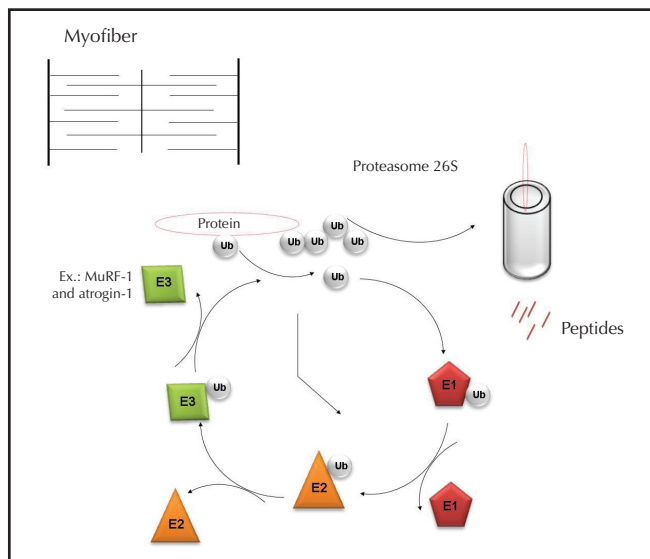
The role of that pathway in human muscle wasting has been reviewed by Murton et al.,<sup>12</sup> who have suggested that the activation of ubiquitin-ligases, MAF-bx/atrogin-1 and MuRF-1, occurs mainly during inflammatory processes.

Muscle atrophy due to the overexpression of the ubiquitin-proteasome system seems to involve different pathways. Some studies have shown that signaling through the NF- $\kappa$ B pathway, which induces the expression of genes related to the sarcopenia process, such as MuRF-1 and MAF-bx, in addition to pro-inflammatory cytokines. The activation of the NF- $\kappa$ B pathway is involved in the muscle atrophy caused by disuse and cachexia, although its mechanisms have not been completely clarified.<sup>51</sup> There is evidence of the involvement of oxidative stress in such activation.<sup>52</sup> In addition to the NF- $\kappa$ B pathway, the increased expression of MuRF-1 and MAF-bx<sup>31</sup> can also occur via FOXO3<sup>30</sup> and myogenin.<sup>53</sup>

### Satellite cells

In addition to protein degradation, deficiencies in the process of muscle regeneration might also be involved in sarcopenia (Figure 1).

Satellite cells (SC) are quiescent myogenic precursors found in the adult muscle between the basal lamina and the sarcolemma, and have some properties of stem cells.<sup>54</sup> SC can be activated in response to stimuli of growth, remodeling, or



**Figure 3** Proteasome system of muscle degradation. Ubiquitin-proteasome system is a cytoplasmic multiprotein complex that degrades proteins labeled with ubiquitin. That degradation requires the participation of three distinct proteins (E1, E2 and E3). The E3 proteins provide specificity to the proteins that will be degraded. In muscle atrophy, some E3 enzymes, such as MuRF-1 and atrogin-1, have been described.

muscle injury.<sup>55,56</sup> When activated, they enter the cell cycle, divide, differentiate into myoblasts, and fuse to form myotubes, which then develop into a new fiber or fuse with already existing muscle fibers to repair damaged myofibers and/or to increase hypertrophy of the muscle fibers.<sup>57</sup>

When activated, SC can be identified by the expression of markers, such as MyoD and myogenin, indicators of SC proliferation and differentiation, respectively.<sup>58</sup>

Some studies have shown that coculture of muscle precursors with macrophages increase proliferation and differentiation of myoblasts, suggesting the involvement of inflammatory mediators in SC activation.<sup>59</sup> Among the inflammatory mediators, TNF- $\alpha$  is increased in muscle tissue after an injury, but also seems to be involved in muscle regeneration.<sup>60,61</sup>

Our group, studying acute inflammatory processes in an experimental model of muscle injury, has demonstrated the important involvement of the local production of nitric oxide in SC proliferation and differentiation.<sup>56,62</sup>

However, little is known about the pathway through which sarcopenia is activated and which initial stimulus triggers the activation of SC in the presence of chronic inflammatory process. There is an apparent contradiction between the increased activation of those regenerative cells and the final result, which is muscle atrophy. Further studies are required to clarify whether that SC activation, which probably occurs as an attempt to regenerate the atrophic muscle, is not sufficient to compensate protein loss, or whether myogenesis is not completed due to, for example, apoptosis.

---

## SARCOPENIA IN RHEUMATOID ARTHRITIS

Despite the progress in understanding the molecular mechanisms that lead to muscle atrophy in several situations, rheumatoid sarcopenia is still rarely studied. Functionally, patients with RA have a significant reduction in muscle strength, but the muscle contractile velocity and properties remain unaltered.<sup>6</sup> Such data demonstrate that the impact of the disease occurs through protein loss, affecting mainly in-parallel sarcomeres, preserving the number of in-series sarcomeres.

Data on muscle wasting pathways in RA, especially involving apoptosis, are scarce. So far, studies on individuals with RA or on animal models of chronic arthritis demonstrating the real role of apoptosis in muscle wasting are still lacking. Studies carried out in our laboratory have not shown apoptotic bodies or labeling with caspase-3 in the gastrocnemius muscle of mice with collagen-induced arthritis (CIA) (unpublished data), suggesting that such mechanism does not play a striking role in rheumatoid sarcopenia.

Similarly, studies on experimental models or patients with RA assessing the participation of mechanisms of autophagy, activation of calpains, and caspases are still lacking.

Regarding the proteasome pathway, the increase in E3 ubiquitin ligases associated with muscle proteolysis has already been identified in the skeletal muscle of murine arthritis models.<sup>63,64</sup> However, the other components of the ubiquitin-proteasome pathway, such as ubiquitin and proteasomal subunits, and the disease stage at which atrophy develops are yet to be studied. In the ubiquitin-proteasome pathway, an increase in the expression of MuRF-1 and MAF-bx<sup>31</sup> has been observed via NF- $\kappa$ B, FOXO3,<sup>30</sup> and myogenin,<sup>53</sup> but such data have not been confirmed in muscles of individuals with arthritis. Thus, although it is the most studied proteolytic pathway in general, its importance is yet to be confirmed in patients with chronic arthropathy.

Finally, in the atrophy of the gastrocnemius muscle in the model of Freund's adjuvant-induced arthritis (AIA), Castellero et al.<sup>64</sup> have shown the activation and proliferation of SC by use of their myogenin and MyoD markers. Such findings require confirmation in other experimental models, as well as in studies with patients.

---

## CONCLUSION

We have discussed how several intracellular pathways are involved, in an inter-related manner, with the process of muscle wasting. Such pathways, comprising mechanisms of cell apoptosis, proteolysis of myofibrils, and alteration in cell regeneration through SC, have been actively studied in several clinical and experimental conditions. Those mechanisms are not uniformly present in those conditions, and their relative importance varies significantly according to the clinical situation. Thus, the best preventive and therapeutic management might not also be the same for all situations of muscle atrophy.

Despite the significant muscle wasting that occurs in most patients with RA, with a significant socioeconomic and functional impact on that population, so far no standardized therapy has been proposed for that complication. There are few studies assessing the impact of current therapies on muscle wasting.<sup>4</sup> Similarly, as already discussed, studies on the participation of several pathways that lead to muscle atrophy and regeneration in either experimental models or in patients with chronic arthropathies are scarce. Further studies on those topics are required, because greater understanding about the mechanisms of modulation between muscle catabolism and anabolism will result in the development of innovative and more effective therapeutic strategies and better quality of life for the patients.



## REFERENCES

## REFERÊNCIAS

1. Kinne RW, Brauer R, Stuhlmuller B, Palombo-Kinne E, Burmester GR. Macrophages in rheumatoid arthritis. *Arthritis Res* 2000; 2(3):189–202.
2. Senna ER, De Barros AL, Silva EO, Costa IF, Pereira LV, Ciconelli RM *et al.* Prevalence of rheumatic diseases in Brazil: a study using the COPCORD approach. *J Rheumatol* 2004; 31(3):594–7.
3. Lee DM, Weinblatt ME. Rheumatoid arthritis. *Lancet* 2001; 358(9285):903–11.
4. Rocha Omd, Batista AdAP, Maestá N, Burini RC, Laurindo IMM. Sarcopenia da caquexia reumatoide: conceituação, mecanismos, consequências clínicas e tratamentos possíveis. *Rev Bras Reumatol* 2009; 49(3):288–301.
5. Doherty TJ. Invited review: Aging and sarcopenia. *J Appl Physiol* 2003; 95(4):1717–27.
6. Matschke V, Murphy P, Lemmey AB, Maddison P, Thom JM. Skeletal Muscle Properties in Rheumatoid Arthritis Patients. *Medicine & Science in Sports & Exercise* 2010; 42(12):2149–55.
7. Rall LC, Roubenoff R. Rheumatoid cachexia: metabolic abnormalities, mechanisms and interventions. *Rheumatology* 2004; 43(10):1219–23.
8. Silva TAA, Frisoli Junior A, Pinheiro MM, Szejnfeld VL. Sarcopenia associada ao envelhecimento: aspectos etiológicos e opções terapêuticas. *Rev Bras Reumatol* [Review] 2006; 46(6):391–7.
9. Ashwell M, Chinn S, Stalley S, Garrow JS. Female fat distribution – A simple classification based on 2 circumference measurements. *International Journal of Obesity* 1982; 6(2):143–52.
10. Calura E, Cagnin S, Raffaello A, Laveder P, Lanfranchi G, Romualdi C. Meta-analysis of expression signatures of muscle atrophy: gene interaction networks in early and late stages. *BMC Genomics* 2008;9.
11. DeBoer MD. Animal models of anorexia and cachexia. *Expert Opinion on Drug Discovery* [Review] 2009; 4(11):1145–55.
12. Murton AJ, Constantin D, Greenhaff PL. The involvement of the ubiquitin proteasome system in human skeletal muscle remodelling and atrophy. *Biochimica Et Biophysica Acta-Molecular Basis of Disease* 2008; 1782(12):730–43.
13. Dupont-Versteegden EE. Apoptosis in skeletal muscle and its relevance to atrophy. *World Journal of Gastroenterology* 2006; 12(46):7463–6.
14. Primeau AJ, Adhietty PJ, Hood DA. Apoptosis in heart and skeletal muscle. *Canadian Journal of Applied Physiology-Revue Canadienne De Physiologie Appliquee* 2002; 27(4):349–95.
15. Cande C, Vahsen N, Garrido C, Kroemer G. Apoptosis-inducing factor (AIF): caspase-independent after all. *Cell Death and Differentiation* 2004; 11(6):591–5.
16. Earnshaw WC, Martins LM, Kaufmann SH. Mammalian caspases: Structure, activation, substrates, and functions during apoptosis. *Annual Review of Biochemistry* 1999; 68:383–424.
17. Garrido C, Kroemer G. Lifes smile, deaths grin: vital functions of apoptosis-executing proteins. *Current Opinion in Cell Biology*. 2004; 16(6):639-46.
18. Bruusgaard JC, Gundersen K. In vivo time-lapse microscopy reveals no loss of murine myonuclei during weeks of muscle atrophy. *Journal of Clinical Investigation* 2008; 118(4):1450–7.
19. Hasselgren PO, Wray C, Mammen J. Molecular regulation of muscle cachexia: It may be more than the proteasome. *Biochemical and Biophysical Research Communications* 2002; 290(1):1–10.
20. Purintrapiban J, Wang MC, Forsberg NE. Degradation of sarcomeric and cytoskeletal proteins in cultured skeletal muscle cells. *Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology* 2003; 136(3):393–401.
21. Powers SK, Kavazis AN, DeRuisseau KC. Mechanisms of disuse muscle atrophy: role of oxidative stress. *American Journal of Physiology-Regulatory Integrative and Comparative Physiology* 2005; 288(2):R337–R44.
22. Lum JJ, DeBerardinis RJ, Thompson CB. Autophagy in metazoans: Cell survival in the land of plenty. *Nature Reviews Molecular Cell Biology* 2005; 6(6):439–48.
23. Bechet D, Tassa A, Taillandier D, Cornbaret L, Attaix D. Lysosomal proteolysis in skeletal muscle. *International Journal of Biochemistry & Cell Biology* 2005; 37(10):2098–114.

24. Goll DE, Neti G, Mares SW, Thompson VF. Myofibrillar protein turnover: The proteasome and the calpains. *J Anim Sci* 2008; 86(14 suppl):E19–35.
25. Mordier S, Deval C, Bechet D, Tassa A, Ferrara M. Leucine limitation induces autophagy and activation of lysosome-dependent proteolysis in C2C12 myotubes through a mammalian target of rapamycin-independent signaling pathway. *Journal of Biological Chemistry* 2000; 275(38):29900–6.
26. Mizushima N, Yamamoto A, Matsui M, Yoshimori T, Ohsumi Y. In vivo analysis of autophagy in response to nutrient starvation using transgenic mice expressing a fluorescent autophagosome marker. *Molecular Biology of the Cell* 2004; 15(3):1101–11.
27. Lecker SH, Jagoe RT, Gilbert A, Gomes M, Baracos V, Bailey J *et al.* Multiple types of skeletal muscle atrophy involve a common program of changes in gene expression. *Faseb Journal* 2004; 18(1):39–51.
28. Satchek JM, Hyatt JPK, Raffaello A, Jagoe RT, Roy RR, Edgerton VR *et al.* Rapid disuse and denervation atrophy involve transcriptional changes similar to those of muscle wasting during systemic diseases. *Faseb Journal* 2007; 21(1):140–55.
29. Nakatogawa H, Suzuki K, Kamada Y, Ohsumi Y. Dynamics and diversity in autophagy mechanisms: lessons from yeast. *Nature Reviews Molecular Cell Biology*. 2009; 10(7):458–67.
30. Mammucari C, Milan G, Romanello V, Masiero E, Rudolf R, Del Piccolo P *et al.* FOXO3 controls autophagy in skeletal muscle in vivo. *Cell Metabolism* 2007; 6(6):458–71.
31. Li H, Malhotra S, Kumar A. Nuclear factor-kappa B signaling in skeletal muscle atrophy. *Journal of Molecular Medicine-Jmm* 2008; 86(10):1113–26.
32. Wohlgemuth SE, Seo AY, Marzetti E, Lees HA, Leeuwenburgh C. Skeletal muscle autophagy and apoptosis during aging: Effects of calorie restriction and life-long exercise. *Experimental Gerontology* 2010; 45(2):138–48.
33. Goll DE, Thompson VF, Li HQ, Wei W, Cong JY. The calpain system. *Physiological Reviews* 2003; 83(3):731–801.
34. Koh TJ, Tidball JG. Nitric oxide inhibits calpain-mediated proteolysis of talin in skeletal muscle cells. *American Journal of Physiology-Cell Physiology* 2000; 279(3):C806–C12.
35. Goll DE, Dayton WR, Singh I, Robson RM. Studies of the alpha-actinin actin interaction in the Z-disk by using calpain. *Journal of Biological Chemistry* 1991; 266(13):8501–10.
36. Goll DE, Thompson VF, Taylor RG, Zaleska T. Is calpain activity regulated by membranes and autolysis or by calcium and calpastatin. *Bioessays* 1992; 14(8):549–56.
37. Du J, Wang XN, Miereles C, Bailey JL, Debigare R, Zheng B *et al.* Activation of caspase-3 is an initial step triggering accelerated muscle proteolysis in catabolic conditions. *Journal of Clinical Investigation* 2004; 113(1):115–23.
38. Naujokat C, Fuchs D, Berges C. Adaptive modification and flexibility of the proteasome system in response to proteasome inhibition. *Biochimica Et Biophysica Acta-Molecular Cell Research* 2007; 1773:1389–97.
39. Naujokat C, Hoffmann S. Role and function of the 26S proteasome in proliferation and apoptosis. *Laboratory Investigation* 2002; 82(8):965–80.
40. Wolf DH, Hilt W. The proteasome: a proteolytic nanomachine of cell regulation and waste disposal. *Biochimica Et Biophysica Acta-Molecular Cell Research* 2004; 1695(1–3):19–31.
41. Li YP, Chen YL, Li AS, Reid MB. Hydrogen peroxide stimulates ubiquitin-conjugating activity and expression of genes for specific E2 and E3 proteins in skeletal muscle myotubes. *American Journal of Physiology-Cell Physiology* 2003; 285(4):C806–C12.
42. Hasselgren PO, Wray C, Mammen J. Molecular regulation of muscle cachexia: it may be more than the proteasome. *Biochem Biophys Res Commun* 2002; 290(1):1–10.
43. Koncarevic A, Jackman RW, Kandarian SC. The ubiquitin-protein ligase Nedd4 targets Notch1 in skeletal muscle and distinguishes the subset of atrophies caused by reduced muscle tension. *Faseb Journal* 2007; 21(2):427–37.
44. Bodine SC, Latres E, Baumhueter S, Lai VK, Nunez L, Clarke BA *et al.* Identification of ubiquitin ligases required for skeletal muscle atrophy. *Science* 2001; 294(5547):1704–8.
45. Centner T, Yano J, Kimura E, McElhinny AS, Pelin K, Witt CC *et al.* Identification of muscle specific ring finger proteins as potential regulators of the titin kinase domain. *J Mol Biol* 2001; 306(4):717–26.
46. Wang K, McClure J, Tu A. Titin: major myofibrillar components of striated muscle. *Proc Natl Acad Sci U S A* 1979; 76(8):3698–702.
47. Lecker SH, Solomon V, Mitch WE, Goldberg AL. Muscle protein breakdown and the critical role of the ubiquitin-proteasome pathway in normal and disease states. *Journal of Nutrition* 1999; 129(1):227S–37S.
48. Tawa NE Jr, Odessey R, Goldberg AL. Inhibitors of the proteasome reduce the accelerated proteolysis in atrophying rat skeletal muscles. *J Clin Invest* 1997; 100(1):197–203.
49. Rennie MJ, Selby A, Atherton P, Smith K, Kumar V, Glover EL *et al.* Facts, noise and wishful thinking: muscle protein turnover in aging and human disuse atrophy. *Scandinavian Journal of Medicine & Science in Sports* 2010; 20(1):5–9.
50. Rajkumar SV, Richardson PG, Hideshima T, Anderson KC. Proteasome inhibition as a novel therapeutic target in human cancer. *Journal of Clinical Oncology* 2005; 23(3):630–9.
51. Jackman RW, Kandarian SC. The molecular basis of skeletal muscle atrophy. *American Journal of Physiology-Cell Physiology* 2004; 287(4):C834–C43.
52. Filippin LI, Vencelino R, Marroni NP, Xavier RM. Influência de processos redox na resposta inflamatória da artrite reumatoide. *Rev Bras Reumatol* 2008; 48(1):17–24.
53. Moresi V, Williams AH, Meadows E, Flynn JM, Potthoff MJ, McAnally J *et al.* Myogenin and Class II HDACs Control Neurogenic Muscle Atrophy by Inducing E3 Ubiquitin Ligases. *Cell* 2010; 143(1):35–45.
54. Cassano M, Quattrocchi M, Crippa S, Perini I, Ronzoni F, Sampaolesi M. Cellular mechanisms and local progenitor activation to regulate skeletal muscle mass. *Journal of Muscle Research and Cell Motility* 2009; 30(7–8):243–53.
55. Morgan JE, Partridge TA. Muscle satellite cells. *Int J Biochem Cell Biol* 2003; 35(8):1151–6.
56. Filippin LI, Cuevas MJ, Lima E, Marroni NP, Gonzalez-Gallego J, Xavier RM. Nitric oxide regulates the repair of injured skeletal muscle. *Nitric Oxide* 2011; 24(1):43–9.
57. Yamada M, Sankoda Y, Tatsumi R, Mizunoya W, Ikeuchi Y, Sunagawa K *et al.* Matrix metalloproteinase-2 mediates stretch-induced activation of skeletal muscle satellite cells in a nitric oxide-dependent manner. *Int J Biochem Cell Biol* 2008; 40(10):2183–91.

58. Berkes CA, Tapscott SJ. MyoD and the transcriptional control of myogenesis. *Seminars in Cell & Developmental Biology* 2005; 16(4-5):585-95.
59. Massimino ML, Rapizzi E, Cantini M, DallaLibera L, Mazzoleni F, Arslan P *et al.* ED2+ macrophages increase selectively myoblast proliferation in muscle cultures. *Biochemical and Biophysical Research Communications* 1997; 235(3):754-9.
60. Chen SE, Jin BW, Li YP. TNF-alpha regulates myogenesis and muscle regeneration by activating p38 MAPK. *American Journal of Physiology-Cell Physiology* 2007; 292(5):C1660-C71.
61. Warren GL, Hulderman T, Jensen N, McKinstry M, Mishra M, Luster MI *et al.* Physiological role of tumor necrosis factor alpha in traumatic muscle injury. *Faseb Journal* 2002; 16(10):1630-+.
62. Filippin LI, Moreira AJ, Marroni NP, Xavier RM. Nitric oxide and repair of skeletal muscle injury. *Nitric Oxide-Biology and Chemistry* 2009; 21(3-4):157-63.
63. Granado M, Martin AI, Priego T, Lopez-Calderon A, Villanua MA. Tumour necrosis factor blockade did not prevent the increase of muscular muscle RING finger-1 and muscle atrophy F-box in arthritic rats. *Journal of Endocrinology* 2006; 191(1):319-26.
64. Castellero E, Martin AI, Lopez-Menduina M, Granado M, Villanua MA, Lopez-Calderon A. IGF-I system, atrogenes and myogenic regulatory factors in arthritis induced muscle wasting. *Molecular and Cellular Endocrinology* 2009; 309(1-2):8-16.