

PERORAL SUSCEPTIBILITY OF *Aedes albifasciatus* AND *Culex pipiens* COMPLEX MOSQUITOES (DIPTERA: CULICIDAE) FROM ARGENTINA TO WESTERN EQUINE ENCEPHALITIS VIRUS

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AVILES, G. et al. Peroral susceptibility of *Aedes albifasciatus* and *Culex pipiens* complex mosquitoes (Diptera: Culicidae) from Argentina to western equine encephalitis virus. Rev. Saúde públ., S. Paulo, 24: 265-9, 1990.

ABSTRACT. The transmission cycle of western equine encephalitis (WEE) virus in South America is unknown. A WEE virus strain was isolated from *Aedes albifasciatus* in Argentina during the WEE epizootic of 1982-83. Also, *Culex pipiens* from Argentina was reported to be able to transmit WEE virus experimentally, but other results indicate that *Cx. pipiens* from the USA is refractory to this virus. We determined the susceptibility of Argentina strains of *Ae. albifasciatus* and *Culex pipiens* complex mosquitoes to infection by WEE virus by the oral route. Adult females were fed on chicks infected with a WEE virus strain isolated in Cordoba Province, Argentina, or were fed on a blood/virus suspension. Each mosquito ingested between $10^{1.6}$ to $10^{6.4}$ vero cell plaque-forming units of virus. Each of 28 *Ae. albifasciatus* was positive for virus from the fourth day postfeeding, and there was evidence for virus replication. In contrast, 0/44 *Cx. p. quinquefasciatus* and only 1/15 *Cx. p. pipiens* was positive. *Aedes albifasciatus* is susceptible to infection by WEE virus and should be considered a potential vector of this virus in Argentina. Both subspecies of *Cx. pipiens* are refractory to peroral infection by WEE virus and probably do not play a role in the WEE virus cycle in Argentina.

KEYWORDS: *Aedes*, microbiology. *Culex*, microbiology. Encephalitis virus, western equine, pathogenicity.

INTRODUCTION

Western equine encephalitis (WEE) virus causes epidemics and epizootics in North and South America. In the Northern Hemisphere, *Culex tarsalis* and *Aedes melanimon* mosquitoes are important vectors^{7,8,15}. In South America, severe aperiodic equine epizootics have been recognized since 1908 in the temperate zone of Argentina and Uruguay, although limited outbreaks in humans have been reported only in Southern Argentina¹⁶. In these areas the natural transmission cycle is unknown.

One hundred and forty-nine thousand mosquitoes collected in Santa Fe Province during the large WEE epizootic of 1982-83, yielded four WEE virus strains from *Ae. albifasciatus*, *Anopheles albitarsis*, *Mansonia* species and *Psorophora pallescens*¹³. The mosquitoes were predominantly *Ae. albifasciatus* (42.8%) and species of *Culex* (*Culex*) (34.4%). In the south, where human cases

had occurred, 474 mosquitoes (70% *Ae. albifasciatus* and 30% *Cx. species*) were tested for virus with negative results.

Villa et al.¹⁷ reported that *Cx. pipiens* from Buenos Aires Province, Argentina, were able to transmit an Argentine strain of WEE virus under experimental conditions. The virus strain had undergone several laboratory passages in guinea pigs and chick embryos, and a high dose of virus in the blood meal was used. In contrast, attempts to experimentally transmit California and Washington strains of WEE virus by *Cx. pipiens* in the USA were unsuccessful¹⁵. Subsequent studies showed that USA strains of *Cx. pipiens* were essentially refractory to peroral infection with WEE virus^{8,9}.

Culex pipiens is represented by two subspecies in Argentina; *Cx. p. quinquefasciatus* occurs in Central and Northern Argentina and *Cx. p. pipiens* in Southern Argentina. Hybrids are present in the zone where the subspecies overlap^{2,10}. The objec-

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tive of the present work was to determine whether Argentine *Cx. pipiens* complex mosquitoes and *Ae. albifasciatus* were susceptible to infection by WEE virus by the oral route.

MATERIALS AND METHODS

WEE virus strain:

The Cba CIV 180 strain was isolated from the brain of a sick horse in Cordoba Province during the epizootic of 1982-83 and had undergone three passages in suckling mouse brains. The virus stock was made with mouse brains harvested 24 to 48 h after intracerebral inoculation and was prepared as a 10% W/V suspension.

Mosquitoes:

Adult *Ae. albifasciatus* females were captured by mechanical aspirator from human bait on a farm near Córdoba City (31° 22'S, 64° 14'W), Córdoba Province, Argentina. Attempts to colonize this species in the laboratory were unsuccessful.

Female *Cx. p. quinquefasciatus* of the second laboratory generation were used at 14 days of age. They originated from a single oviposition from females collected in the urban area of Córdoba City. *Culex p. pipiens* of the second and the third generations came from two ovipositions of females collected in Puerto Madryn, (42° 50'S, 65° 05'W), Chubut Province, Argentina, and were used as 5-to 10-day old females. The subspecies were determined by examining the male genitalia¹ of thirteen specimens from each colony. No intermediates between *pipiens* and *quinquefasciatus* were found.

Experimental Design:

Mosquitoes were infected by feeding on 1-to 12-day-old viremic chicks previously inoculated subcutaneously with approximately $10^{3.0}$ - $10^{3.7}$ vero cell culture plaque-forming units (PFU) of virus.

Since few *Ae. albifasciatus* fed, an additional attempt was made to expose mosquitoes to infection by the oral route by using an artificial feeding technique. A heparinized-blood/virus suspension was prepared by mixing 0.5 ml of the stock virus (titer $10^{8.0}$ PFU/0.1 ml), 3 ml of normal guinea pig blood, and 1.5 ml of Media 199. The suspension contained $10^{5.7}$ PFU/5 μ l and the mosquitoes were allowed to feed during a 2-h period. *Culex pipiens* mosquitoes were starved 24 h before the experiments were initiated.

Mosquitoes were allowed to feed on viremic chicks during a 9-to 15-h period at 10 to 24 h postinoculation. Serum specimens were taken from the chicks preexposure and postexposure. Blood was diluted 1:10 in a heparinized diluent and centri-

fuged at 4°C and 750 x g for 30 minutes. Supernatants were frozen at -80°C until tested for virus. A group of freshly fed mosquitoes also was frozen at -80°C until tested for virus and used as the sample for the first day of incubation.

The remaining fed mosquitoes were incubated in a secure room inside a laboratory with level 2 biosafety (Biosafety in Microbiological and Biomedical Laboratories, 1984), at 26°C maximum-21°C minimum for *Ae. albifasciatus*, 30°C maximum-24°C minimum for *Cx. p. quinquefasciatus* and 26°C maximum-15°C minimum for *Cx. p. pipiens*, in a humid atmosphere and at natural photoperiods with at least 11 h of full light.

Samples of 1 to 11 mosquitoes were frozen on different days of incubation for subsequent titration. They were triturated individually with 1 ml of diluent (Media 199, 20% fetal calf serum and gentamicin 5 mg%). Suspensions were centrifuged at 11,400 x g for 30 minutes at 4°C and supernatants were serially diluted in the same diluent and inoculated in 0.1-ml volumes into C1 76 vero cell culture under double agar overlay⁵. The second agar overlay was added after 48 h, and the cell cultures were examined for 3 days and characteristic plaques were counted. Virus titers of mosquitoes are expressed as PFU/mosquito. The infection rate of mosquitoes was established taking into account the number of infected mosquitoes and the total number tested from days 3 and 4 to day 21 postinfection.

RESULTS

Culex p. quinquefasciatus fed readily and survived well. *Ae. albifasciatus* and *Cx. p. pipiens* showed low feeding rates and high mortality. A total of 41 *Ae. albifasciatus*, 48 *Cx. p. quinquefasciatus*, and 21 *Cx. p. pipiens* were tested (Table 1).

TABLE 1

Feeding rates and survival of *Aedes albifasciatus* and two subspecies of *Culex pipiens* during infection experiments with WEE virus.

Mosquito species/subspecies	No. of feeding trials	Number exposed	Number fed	Number survived and tested for virus
<i>Aedes albifasciatus</i>	13	651	86	41
<i>Cx. p. quinquefasciatus</i>	1	60	52	48
<i>Cx. p. pipiens</i>	2	85	25	21

Virus titers of the infective meals (chick bloods and viral suspension) and individual *Ae. albifasciatus* titers are shown in Table 2. *Ae. albi-*

fasciatus sampled on the first day had virus titers of $10^{3.9}$ to $10^{6.6}$ PFU/mosquito. All *Ae. albifasciatus* tested after the fourth day were positive with virus titers of $10^{3.3}$ to $10^{6.7}$ PFU/mosquito.

TABLE 2

Cba CIV 180 WEE virus titers in the infective meals and in individual *Aedes albifasciatus* mosquitoes infected by peroral route.

Infective meal virus pre-post exposure period (log PFU /5 µl)	Virus titers (log PFU/mosquito) of individual mosquitoes per day of extrinsic incubation								
	1	2	4	5	6	8	9	11	13
1.6 - 2.7	4.4		5.3			6.7			
			6.7			5.8			
			6.0						
			4.6						
3.9 - 3.9	3.9		5.5						
			5.7						
			6.6						
4.1 - 4.4				6.3					4.5
				6.5					4.5
									5.6
4.7 - 4.9				4.5					
				5.5					
5.0 - 5.4	5.3			3.5					
				5.9					
4.6 - 5.7	3.9							5.6	
	4.8								
5.4 - 5.7	5.8		6.3					6.0	
	5.8		6.5						
			6.6						
5.7*	5.8		5.3		5.0				
5.0 - 5.8		4.9				3.3			
5.2 - 6.0	4.8							5.0	
5.5 - 6.3	6.6								
	5.9								
3.7 - 6.4	6.5							5.3	

n 12 1 11 6 1 3 3 1 3
 x 5.3 5.9 5.3 5.2 5.4 4.8

* viral suspension

Culex p. quinquefasciatus were fed an infective meal that titered $10^{4.3}$ to $10^{4.5}$ PFU/ml. Four freshly-fed females contained $10^{2.2}$ to $10^{2.5}$ PFU/mosquito immediately after feeding. Thereafter, none of 44 females tested on days 3 through 20 postfeeding were positive for virus. *Culex p. pipiens* were fed on an infective meal that titered $10^{5.5}$ to $10^{7.5}$ PFU/ml. Six freshly-fed females contained $10^{3.1}$ to $10^{4.1}$ PFU/mosquito immediately after feeding. Thereafter, only 1 of 15 mosquitoes tested on days 4 through 21 postfeeding was positive for virus. The positive female was tested on day 4 postfeeding and contained $10^{2.1}$ PFU of virus.

Using an estimated blood meal volume of 5 µl,

it was calculated that each *Ae. albifasciatus* ingested between $10^{1.6}$ to $10^{6.4}$ PFU, each *Cx. p. quinquefasciatus* between $10^{2.0}$ to $10^{2.2}$ PFU, and each *Cx. p. pipiens* between $10^{3.1}$ to $10^{5.1}$ PFU of virus.

DISCUSSION

Virus titers in freshly-fed mosquitoes (day one of incubation) represent remnants of ingested virus and correlate with titers of the infective meals. The results of Villa et al.¹⁷ suggested that WEE virus could infect and be transmitted by *Cx. pipiens* after ingestion of large doses of virus ($10^{7.3}$ LD₅₀/5 µl for 3-week-old mice). According to our results, Argentinian *Cx. pipiens* of both subspecies have a low capacity to become infected with Argentinian WEE virus, and the infection threshold is $>10^3$ PFU for *Cx. p. quinquefasciatus* and $>10^2$ PFU for *Cx. p. pipiens*. These titers are sufficient to infect known vector species of *Culex* and *Aedes*^{4,6}. The lack of isolations of WEE virus from *Culex* in nature during WEE epizootics in Argentina, combined with our results, suggest that *Cx. pipiens* does not play an important role in the WEE virus cycle in Argentina. Our results concerning the vector competence of *Cx. pipiens* for WEE virus agree with those from the USA^{8,9,15}.

The demonstrated susceptibility of *Ae. albifasciatus* to infection with WEE virus by the oral route is further evidence for considering this species to be a potential vector of WEE virus in Argentina. *Aedes albifasciatus* has been found naturally infected and it was an abundant mosquito during the 1982-83 epizootic¹³. This mosquito feeds mainly on mammals, especially bovines and equines^{11,12,14}. It has been suggested that *Ae. albifasciatus* may serve as a vector in a WEE virus transmission cycle involving the European hare (*Lepus europaeus*) in Argentina¹⁴. This would correspond to a similar cycle involving *Aedes melanimon* and *Lepus californicus* in California⁷. Further incrimination of *Ae. albifasciatus* as a vector of WEE virus must await more definitive experiments on vector competence that include determination of transmission efficiency. This is difficult to accomplish because the species has not been colonized, and it is difficult to get field-collected specimens to feed, survive, oviposit, and refeed in the laboratory.

ACKNOWLEDGMENTS

To Drs. Mireya de Brewer, Biol. Liliana Bufa and Biol. Walter Almiron, for their assistance in identifying *Culex pipiens* complex mosquitoes to subspecies. To the Centre National Patagonico (CONICET), especially to Dr. Jose Luis Garrido for facilitating our work in Chubut Province, and for the economic assistance to CONICET, CONICOR (Consejo de Investigaciones de Córdoba) y SE.NA.SA. (Servicio Nacional de Sanidad Animal).

AVILES, G. et al. Suscetibilidade por via oral dos mosquitos *Aedes albifasciatus* e do complexo *Culex pipiens* (Diptera: Culicidae) da Argentina ao vírus da encefalite equina tipo oeste. Rev. Saúde públ., S. Paulo, 24: 265-9, 1990.

RESUMO: Desconhece-se o ciclo de transmissão da encefalite equina tipo oeste (WEE) na América do Sul. Uma cepa do vírus foi isolada na Argentina, durante a epizootia de 1982-1983, a partir de *Aedes albifasciatus*. Sob o ponto de vista experimental, o *Culex pipiens* da Argentina revelou-se capaz de transmitir o vírus WEE, porém outros resultados têm indicado que o *Cx. pipiens* dos Estados Unidos é refratário a esse vírus. Assim, procurou-se determinar a suscetibilidade de cepas argentinas de *Ae. albifasciatus* e complexo *Culex pipiens*, à infecção do vírus WEE por via oral. As fêmeas adultas foram alimentadas em pintos infectados com cepa do vírus isolada na Província de Córdoba, Argentina, ou então alimentadas em suspensão do vírus e sangue. Cada mosquito ingeriu entre $10^{1,6}$ e $10^{6,4}$ unidades virais formadoras de placas de cultura de célula ("vero cell"). Cada um dos 28 *Ae. albifasciatus* mostrou-se a partir do quarto dia pós-prandial e houve evidência de replicação viral. Em contraposição, 0/44 *Cx. p. quinquefasciatus* e apenas 1/15 *Cx. p. pipiens* revelou-se positivo. *Aedes albifasciatus* é suscetível à infecção pelo vírus WEE e deveria ser considerado vetor potencial desse agente na Argentina. Ambas subespécies de *Cx. pipiens* são refratárias à infecção por via oral e provavelmente não desempenham papel do ciclo do vírus WEE na Argentina.

DESCRITORES: *Aedes*, microbiologia. *Culex*, microbiologia. Vírus da encefalite equina ocidental, patogenicidade.

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Received in 26/4/1990

Accepted in 18/6/1990