

Cassava processing wastewater as a platform for third generation biodiesel production

Cristina Neves¹, Mariana Manzoni Maroneze¹, Aline Meireles dos Santos¹, Erika Cristina Francisco², Roger Wagner¹, Leila Queiroz Zepka¹, Eduardo Jacob-Lopes^{1*}

¹Federal University of Santa Maria – Dept. of Food Science and Technology, Av. Roraima, 1000 – 97105-900 – Santa Maria, RS – Brazil.

²University of Passo Fundo/FEAR, Rod. BR 285 – 99052-900 – Passo Fundo, RS – Brazil.

*Corresponding author <jacoblopes@pq.cnpq.br>

Edited by: Julio Cesar Pascale Palhares

Received July 24, 2015

Accepted October 09, 2015

ABSTRACT: This study aimed to evaluate third generation biodiesel production by microalgae *Phormidium autumnale* using cassava processing wastewater as a platform. Experiments were performed in a heterotrophic bubble column bioreactor. The study focused on the evaluation of the bioreactor (batch and fed-batch) of different operational modes and the analysis of biofuel quality. Results indicate that fed-batch cultivations improved system performance, elevating biomass and oil productions to 12.0 g L⁻¹ and 1.19 g L⁻¹, respectively. The composition of this oil is predominantly saturated (60 %) and monounsaturated (39 %), resulting in a biodiesel that complies with U.S., European and Brazilian standards. The technological route developed indicates potential for sustainable production of bulk oil and biodiesel, through the minimization of water and chemical demands required to support such a process.

Keywords: microalgae, cyanobacteria, agroindustrial wastewater, biofuel

Introduction

Increasing greenhouse gas emissions and declining fossil fuel resources are the major driving forces behind the search for sustainable renewable biofuels (Zhou et al., 2014). In this context, process integration has emerged as a way to allow for sustainable manufacturing in industrial processing (Friedler, 2010). This approach has been defined as a set of methodologies that combine several process elements to reduce energy consumption and environmental pollution (Porzio et al., 2014).

Among the most common biofuels, biodiesel is an attractive option due to its high energy density, low NO_x and SO_x emission after combustion, and its compatibility with existing vehicle engines (Rashid et al., 2014). Biofuels derived from microalgae are promising alternatives as third generation biofuels due to the unique characteristics inherent in algae, such as fast proliferation, high oil accumulation, low water consumption rates and feasibility of growing on non-arable lands (Huang et al., 2015).

In addition, microalgae are considered to have the potential to be used as biocatalysts in an integrated bioprocess (Charpentier, 2005). According to Maroneze et al. (2014), these features enable the use of agro-industrial waste as a carbon source, making it economically feasible while contributing to agroindustrial waste management.

Brazil is one of the largest global producers of cassava, whose processing to produce flour and starch gives rise to about 250-300 L wastewater per ton of processed cassava (FAO, 2001). This waste is mainly composed of organic matter and nutrients, and, furthermore, exhibits low levels of toxic compounds or microalgae growth inhibitors (Damasceno et al., 2003). It makes the cassava processing industry a potential platform for microalgae-based processes (Subhadra and Edwards, 2011).

Phormidium is a genus of filamentous, unbranched cyanobacteria, with filaments with a min diameter of 3-4 μ. Several species are known to live in extreme environments such as thermal springs, desert soils and polluted sites, which makes them robust and have simple nutritional requirements (Guiry and Guiry, 2015).

In this regard, this study aimed to evaluate third generation biodiesel production by microalgae *Phormidium autumnale* using cassava processing wastewater as a platform. The study focused on both the evaluation of different operational modes of a bioreactor (batch and fed-batch) and the analysis of biofuel quality.

Materials and Methods

Microorganisms and culture media

Axenic cultures of *Phormidium autumnale* were originally isolated from the Cuatro Ciénegas desert (26°59' N, 102°03' W - Mexico). Stock cultures were propagated and maintained in solidified agar-agar (20 g L⁻¹) containing synthetic Braun-Grunow medium (BG11) (Rippka et al., 1979) with the following composition (mg L⁻¹): K₂HPO₄ (30), MgSO₄ (75), CaCl₂·2H₂O (36), ammonium citrate and iron (0.6), Na₂EDTA (1), NaCl (0.72), NaNO₃ (15), citric acid (0.6), Na₂CO₃ (1500), trace metals [H₃BO₃ (2.8), MnCl₂·4H₂O (1.8), ZnSO₄·7H₂O (0.22), Na₂MoO₄·2H₂O (0.39), CoSO₄·6H₂O (0.04)]. The incubation conditions prevailing were a temperature of 20 °C, a photon flux density of 15 μmol m⁻² s⁻¹ and a photoperiod of 12 h.

Wastewater

Cassava wastewater from the flour industry (Garça, SP, Brazil) was used as a culture medium. Wastewater was collected on a monthly basis from the discharge point of the hydraulic pressing step (de-watering) for 12 months. Analyses for pH, chemical oxygen demand (COD), total nitrogen (N-TKN), total phosphorus (P-PO₄⁻³), total solids

(TS), suspended solids (SS) and volatile solids (VS) were performed following the Standard Methods for Examination of Water and Wastewater (APHA, 2005). The results are shown in Table 1. The carbon/nitrogen ratio (C/N) and the nitrogen/phosphorous ratio (N/P) were calculated through COD, N-TKN and $P-PO_4^{3-}$ and adjusted with wastewater dilution in distilled water and ammonium nitrate to obtain the C/N ratio required.

Bioreactor

Measurements were taken in a bubble column bioreactor. The system was made of borosilicate glass and had an external diameter of 12.5 cm and a height of 16 cm, resulting in a height/diameter (h/D) ratio equal to 1.28 and a nominal working volume of 2.0 L. The dispersion system of the reactor consisted of a 2.5 cm diameter air diffuser located inside the bioreactor. The airflow was monitored by a flow meter and the inlet of air and outlet of gases were filtered through filtering units made up of a polypropylene membrane with a pore diameter of 0.22 μm and total diameter of 50 mm. Two operational modes (batch and fed-batch) were considered in the experiments.

For batch cultivations, the bioreactor was fed with 2.0 L of previously sterilized cassava processing wastewater (15 psi/121 °C). The experimental conditions were as follows: initial cell concentration of 0.1 g L^{-1} , pH adjusted to 7.6, temperature of 30 °C, C/N ratio of 68 (12.0 g L^{-1} of organic carbon), constant aeration of 1VVM (volume of air per volume of wastewater per minute), and absence of light.

The same experimental conditions were used in the fed-batch cultivations. For the fed-batch culture, the feeding strategy was based on use of the cassava processing wastewater and cassava starch as carbon sources. The feed was added in pulses, when organic carbon concentration reached 6.0 g L^{-1} , with organic carbon content being adjusted to 12.0 g L^{-1} . This procedure was repeated until the culture reached a stationary phase.

The experiments were performed twice, and in duplicate for each operational mode. Therefore, kinetic data refer to the mean value of four repetitions.

Sampling and analytical methods

Samples of 20 mL were collected from the bioreactor aseptically in a laminar flow hood every 24 h during the growth phase of the microorganism. All analyses were performed twice, and in duplicate for each experimental condition.

Cell biomass was gravimetrically evaluated by filtering 10 mL of culture medium through a 0.45 μm membrane filter, drying at 60 °C until constant weight. Organic carbon concentration was expressed in terms of chemical oxygen demand (COD) and analyzed according to the closed reflux colorimetric method of Standard Methods for the Examination of Water and Wastewater (APHA, 2005). The filtered fraction of the gravimetric procedure was used for the evaluation of organic carbon concentration. At the end of the process, the biomass was separated from the culture medium by decantation, followed by centrifugation, drying and milling.

Total lipid concentration of the biomass was determined gravimetrically by the modified Bligh and Dyer method (1959), using the ratio between methanol, chloroform and distilled water of 2:1:0.8 (v/v/v). The method of Hartman and Lago (1976) was used to saponify and esterify the dried lipid extract to obtain the fatty acid methyl esters (biodiesel). Fatty acid composition was determined using a gas chromatograph. Fatty acid methyl esters were identified by comparison of retention times with the authentic standards and quantified through area normalization by the Chromatography Station T2100p (Plus Edition) v 9.04 software.

The fuel properties of biodiesel (ester content, EC; cetane number, CN; iodine value, II; degree of unsaturation, DU; saponification value, SV; long-chain saturated factor, LCSF; cold filter plugging point, CFPP; cloud point, CP; allylic position equivalents, APE; bis-allylic position equivalents, BAPE; oxidation stability, OS; higher heating value, HVV; kinematic viscosity, μ and kinematic density, ρ) were determined by the BiodieselAnalyzer® 1.1 software, which estimates biodiesel properties based on the fatty acid profile of the parent oil, through a system of empirical equations (Talebi et al., 2014).

Results and Discussion

The performance parameters of the process in different operating modes are shown in Table 2. The fed-batch strategies intensified the conversion of organic carbon into microalgal biomass. Five feeding pulses with cassava processing wastewater improved the performance of the bioreactor, reaching maximum cell densities of 12.0 g L^{-1} . In this operational condition, an average rate of organic carbon consumption of 0.105 $\text{g L}^{-1} \text{h}^{-1}$, a biomass yield coefficient of 0.39 $\text{g}_{\text{biomass}} \text{g}_{\text{carbon}}^{-1}$, and a global organic carbon conversion of 78 % were obtained, in a residence time of 336 h (Figure 1). The discontinuous feeding in the bioreactor

Table 1 – Composition of cassava processing wastewater.

Parameter	Value
pH	5.47 ± 0.05
COD (g L^{-1})	24.0 ± 0.35
N-TKN (g L^{-1})	0.25 ± 0.01
$P-PO_4^{3-}$ (g L^{-1})	0.16 ± 0.01
TS (g L^{-1})	35.40 ± 0.28
SS (g L^{-1})	25.31 ± 0.22
VS (g L^{-1})	10.09 ± 0.13
C/N	96.0 ± 4.24
N/P	1.50 ± 0.07

COD: chemical oxygen demand; N-TKN: total nitrogen; $P-PO_4^{3-}$: total phosphorus; TS: total solids; SS: suspended solids; VS: volatile solids; C/N: carbon/nitrogen ratio; N/P: nitrogen/phosphorus ratio.

kept the organic carbon concentration between 6 and 12 g L⁻¹, providing cells with sufficient substrate to extend their growth during the cultivation cycle, reducing the restriction to a minimum and increasing biomass yield. This range of organic carbon is associated with the period of the highest volumetric growth rates.

Comparatively, the biomass productions obtained are higher than those reported by Lu et al. (2010) and Vidotti et al. (2014) who reported values of 4.2 g L⁻¹ and 3.5 g L⁻¹ for *C. protothecoides* and *C. vulgaris*, respectively, in cultivations of cassava processing wastewater. In addition, Bastos et al. (2014) reported maximum cell densities of 0.80 g L⁻¹ in the cultivation of cyanobacteria *A. microscopica Nageli* in rice processing wastewater.

In terms of bulk oil production, the feed-batch strategies reduced the lipid content of the biomass (9-10 %) in comparison with batch cultivations (14 %). However, as microalgal oils are intracellular products, the best conditions for oil production have to combine biomass production and lipid content. Thus, lipid productions of 1.19 g L⁻¹ were evidenced in feed-batch cultivations with cassava processing wastewater. A faster product formation rate implies higher produc-

Table 2 – Kinetic parameters for the process in different operating modes.

Parameter	Operational mode		
	Batch	Fed-Batch (wastewater)	Fed-Batch (Starch)
X _{max} (g L ⁻¹)	5.2 ± 0.07	12.0 ± 0.02	8.2 ± 0.05
r _s (g L ⁻¹ h ⁻¹)	0.102 ± 0.00	0.105 ± 0.00	0.098 ± 0.00
Y _{X/S} (g g ⁻¹)	0.52 ± 0.00	0.39 ± 0.00	0.28 ± 0.00
X (%)	66.1 ± 0.7	78.7 ± 1.9	73.99 ± 0.9
Lipid (%)	14.0 ± 0.4	10.0 ± 0.2	9.0 ± 0.1
P _L (g L ⁻¹)	0.73 ± 0.00	1.19 ± 0.01	0.74 ± 0.01
RT (h)	120 ± 0.00	336 ± 0.00	360 ± 0.00

X_{max}: maximum cell density; r_s: average rate of organic carbon conversion; Y_{X/S}: biomass yield coefficient; X: global organic carbon conversion; P_L: lipid production; RT: residence time.

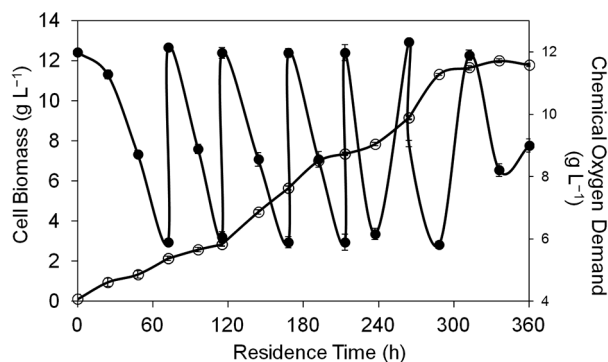


Figure 1– Cell biomass and substrate consumption dynamics in feed-batch cultivations (feeding pulses with cassava processing wastewater).

tions and corresponding reductions in plant operating time and operating cost, for an existing plant. By contrast, in order for a new plant to be built, the increase in response rates implies an increase in production, which can be achieved by means of a smaller bioreactor; as a result, less capital investment is required (Rosso et al., 2015).

The composition of this oil (Table 3) indicated eight different compounds, with oleic acid (33 %) being the main one. Microalgal oil showed a predominantly saturated (60 %) and monounsaturated (39 %) profile. The biodiesel produced from microalgal oils has the following fuel properties (Table 4): ester content of 99 %, cetane number of 58.6, iodine value of 36.3 g I₂ 100 g⁻¹, degree of unsaturation of 39 %, saponification value of 266.4, long-chain saturated factor of 4 %, cold filter

Table 3 – Fatty acid profile in fed-batch (wastewater) operational mode.

Fattyacids	Percent (%)
Caproicacid (C6:0)	3.80 ± 0.0
Caprylicacid (C8:0)	26.8 ± 0.3
Lauricacid (C12:0)	0.97 ± 0.0
Myristicacid (C14:0)	0.87 ± 0.0
Palmiticacid (C16:0)	20.8 ± 0.1
Palmitoleic acid (C16:1)	5.88 ± 0.0
Stearic acid (C18:0)	6.93 ± 0.0
Oleic acid (C18:1n9c)	33.88 ± 0.7
Saturated	60.1
Monounsaturated	39.7

Table 4 – Properties of microalgal biodiesel produced in fed-batch (wastewater) operational mode and its comparison with soybean and the standards used in the US (ASTM 6751), Europe (EN 14214) and Brazil (ANP 255).

Properties	Microalgae	Soybean*	ANP 255	ASTM 6751	EN 14214
EC (%)	99.8	96.9	-	-	min 96.5
CN	58.6	49.0	min 45	min 47	min 51
IV (gI ₂ 100 g ⁻¹)	36.3	128	-	-	max 120
DU (%)	39.7	143.8	-	-	-
SV	266.4	-	-	-	-
LCSF (%)	5.55	1.6	-	-	-
CFPP (°C)	0.96	-5.0	max 19	-	-
CP (°C)	5.95	-	-	-	-
APE	33.8	-	-	-	-
BAPE	0.00	-	-	-	-
OS (h)	0.00	1.3	-	min 3	min 6
HVV	37.5	-	-	-	-
μ (mm ² s ⁻¹)	0.90	4.2	-	1.9-6.0	3.5-5.0
ρ (g cm ⁻³)	0.87	-	-	-	-

EC: ester content; CN: cetane number; IV: iodine value; DU: degree of unsaturation; SV: saponification value; LCSF: long-chain saturated factor; CFPP: cold filter plugging point; CP: cloud point; APE: allylic position equivalents; BAPE: bis-allylic position equivalents; OS: oxidation stability; HVV: higher heating value; μ: kinematic viscosity; ρ: kinematic density. *Knothe (2005).

plugging point at 5.5 °C, cloud point at 0.96 °C, allylic position equivalents of 33.8, bis-allylic position equivalents of zero, oxidation stability of 0 h, higher heating value of 37.5, kinematic viscosity of 0.90 mm² s⁻¹ and kinematic density of 0.87 g cm⁻³. All these parameters, with the exception of oxidative stability and viscosity, comply with the limits established by U.S., European, and Brazilian standards (ASTM, 2002; UNE-EN, 2003; ANP, 2003), and are comparable to soybean biodiesel (Knothe, 2005). Non-conformities can be circumvented by the addition of antioxidants and additives or the blending of biodiesel with petrodiesel, which improves the quality properties of the biofuel (Hui, 2006; Chu et al., 2013).

Finally, if the values of water, organic carbon, nitrogen and phosphorus are considered, the cassava processing wastewater has a potential value of 12.5 USD m⁻³, based on equivalents of industrial water, industrial glucose, ammonium nitrate and sodium phosphate (USDA, 2015). These compounds are the feedstocks commonly used for heterotrophic microalgae cultures, demonstrating substantial potential for exploration of this waste to support the production of bulk oil and biodiesel by microalgae.

Conclusion

Cassava processing wastewater seems a good culture medium for supporting the growth and single-cell oil production of *Phormidium autumnale*. The high lipid production capacity potential obtained is interesting for the generation of quality biodiesel that meets or surpasses the most stringent US, European and Brazilian fuel standard requirements.

Acknowledgments

Funding for this research was provided by the *Brazilian National Council for Scientific and Technological Development* (CNPq).

References

- American Public Health Association [APHA]. 2005. Standard Methods for the Examination of Water and Wastewater. 20ed. APHA, Washington, DC, USA.
- American Society for Testing and Materials [ASTM]. 2002. ASTM 6751 - Standard Specification for Biodiesel Fuel (B100) Blend Stock for Distillate Fuels. ASTM, West Conshohocken, PA, USA.
- Bastos, R.G.; Bonini, M.A.; Zepka, L.Q.; Jacob-Lopes, E.; Queiroz, M.I. 2014. Treatment of rice parboiling wastewater by cyanobacterium *Aphanothece microscopica Nægeli* with potential for biomass products. *Desalination and Water Treatment* 6: 1-7.
- Bligh, E.G.; Dyer, J.W. 1959. A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology* 37: 911-917.
- Charpentier, J.C. 2005. Process intensification by miniaturization. *Chemical Engineering & Technology* 107: 3-17.
- Chu, J.M.; Xu, X.Q.; Zhang, Y.L. 2013. Production and properties of biodiesel produced from *Amygdalus pedunculata* Pall. *Bioresource Technology* 134: 374-376.
- Damasceno, S.; Cereda, M.P.; Pastore, G.M.; Oliveira, J.G. 2003. Production of volatile compounds by *Geotrichum fragrans* using cassava wastewater as substrate. *Process Biochemistry* 39: 411-414.
- European Standard [EN]. 2003. UNE-EN 14214 - Automotive Fuels - Fatty Acid Methyl Esters (FAME) for Diesel Engine - Requirements and Test Methods. European Standard, Pilsen, Czech Republic.
- Food and Agriculture Organization [FAO]. 2001. An Assessment of the Impact of Cassava Production and Processing on the Environment and Biodiversity. FAO, Rome, Italy.
- Friedler, F. 2010. Process integration, modeling and optimization for energy saving and pollution reduction. *Applied Thermal Engineering* 30: 2270-2280.
- Guiry, M.D.; Guiry, G.M. 2015. Algae base: world-wide electronic publication. National University of Ireland, Galway. Available at: <http://www.algaebase.org/> [Accessed Jun. 24, 2015]
- Hartman, L.; Lago, R.C.A. 1976. Rapid preparation of fatty acids methyl esters. *Laboratory Practice* 22: 475-476.
- Huang, J.; Xia, J.; Jiang, W.; Li, Y.; Li, J. 2015. Biodiesel production from microalgae oil catalyzed by a recombinant lipase. *Bioresource Technology* 180: 47-53.
- Hui, Y.H. 2006. *Handbook of Food Science, Technology and Engineering*. CRC Press, Boca Raton, FL, USA.
- Knothe, G. 2005. Dependence of biodiesel fuel properties on the structure of fatty acid alkyl esters. *Fuel Processing Technology* 86: 1059-1070.
- Lu, Y.; Zhai, Y.; Liu, M.; Wu, Q. 2010. Biodiesel production from algal oil using cassava (*Manihot esculenta* Crantz) as feedstock. *Journal of Applied Phycology* 22: 573-578.
- Maroneze, M.M.; Menezes, C.R.; Barin, J.S.; Queiroz, M.I.; Zepka, L.Q.; Jacob-Lopes, E. 2014. Treatment of cattle-slaughterhouse wastewater and the reuse of sludge for biodiesel production by microalgal heterotrophic bioreactors. *Scientia Agricola* 71: 521-524.
- National Petroleum Agency [ANP]. 2003. ANP 255 - Provisional Brazilian Biodiesel Standard. ANP, Brasília, DF, Brazil.
- Porzio, G.F.; Colla, V.; Matarese, N.; Nastasi, G.; Branca, T.A.; Amato, A.; Fornai, B.; Vannucci, M.; Bergamasco, M. 2014. Process integration in energy and carbon intensive industries: an example of exploitation of optimization techniques and decision support. *Applied Thermal Engineering* 70: 1148-1155.
- Rashid, N.; Rehman, M.S.U.; Sadiq, M.; Mahmood, T.; Han, J.I. 2014. Current status, issues and developments in microalgae derived biodiesel production. *Renewable and Sustainable Energy Reviews* 40: 760-778.
- Rippka, R.; Deruelles, J.; Waterbury, J.B.; Herdman, M.; Stainer, R.Y. 1979. Generic assignments, strain histories and properties of pure cultures of cyanobacteria. *Journal of General Microbiology* 111: 1-61.

- Roso, G.R.; Queiroz, M.I.; Streit, N.; Menezes, C.R.; Zepka, L.Q.; Jacob-Lopes, E. 2015. The bioeconomy of microalgal carotenoid-rich oleoresins produced in agroindustrial biorefineries. *Journal of Chemical Engineering & Process Technology* 6: 1-7.
- Subhadra, B.G.; Edwards, M. 2011. Co-product market analysis and water foot print of simulated commercial algal biorefineries. *Applied Energy* 88: 3515-3523.
- Talebi, A.F.; Tabatabaei, M.; Chisti, Y. 2014. Biodiesel analyzer: a user-friendly software for predicting the properties of prospective biodiesel. *Biofuel Research Journal* 2: 55-57.
- United States Department of Agricultural [USDA]. 2015. Economic Research Service. Available at: <http://www.ers.usda.gov/> [Accessed Jul. 15, 2015]
- Vidotti, A.D.S.; Coelho, R.S.; Franco, L.M.; Franco, T.T. 2014. Miniaturized culture for heterotrophic microalgae using low cost carbon sources as a tool to isolate fast and economical strains. *Chemical Engineering Transactions* 38: 325-330.
- Zhou, W.; Chen, P.; Min, M.; Ma, X.; Wang, J.; Griffith, R.; Hussain, F.; Peng, P.; Xie, Q.; Li, Y.; Shi, J.; Meng, J.; Ruan, R. 2014. Environment-enhancing algal biofuel production using wastewater. *Renewable and Sustainable Energy Reviews* 36: 256-269.